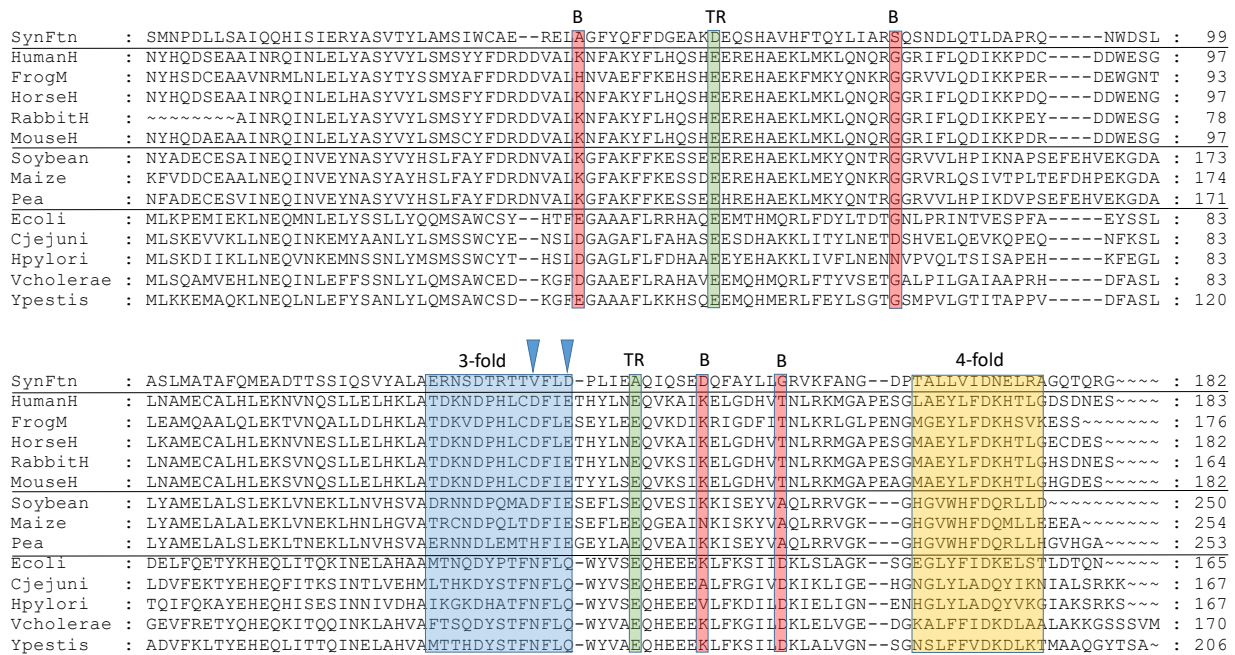


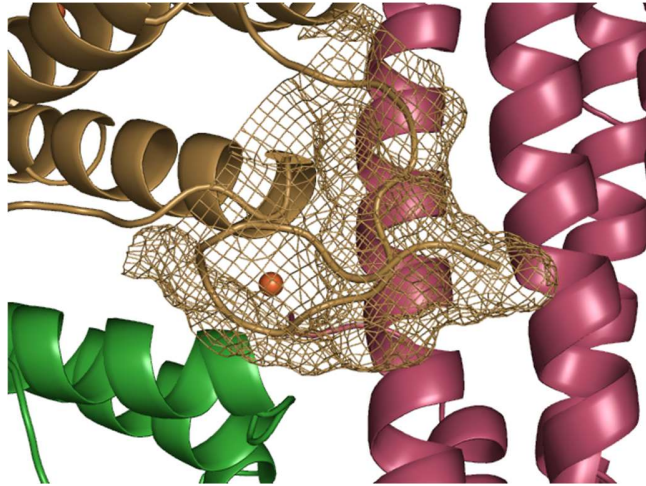
## Supporting Information

Routes of iron entry into, and exit from, the catalytic ferroxidase sites of the prokaryotic ferritin *SynFtn*

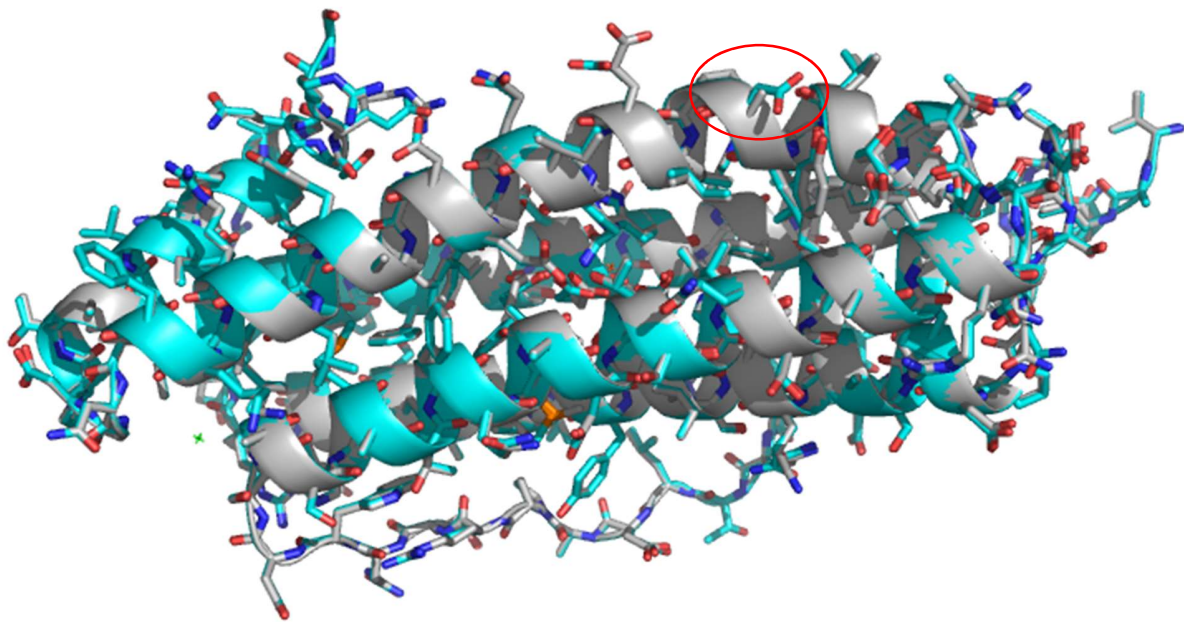
Justin M. Bradley, Jacob Pullin, Geoffrey R. Moore, Dimitri A. Svistunenko, Andrew M. Hemmings and Nick E. Le Brun



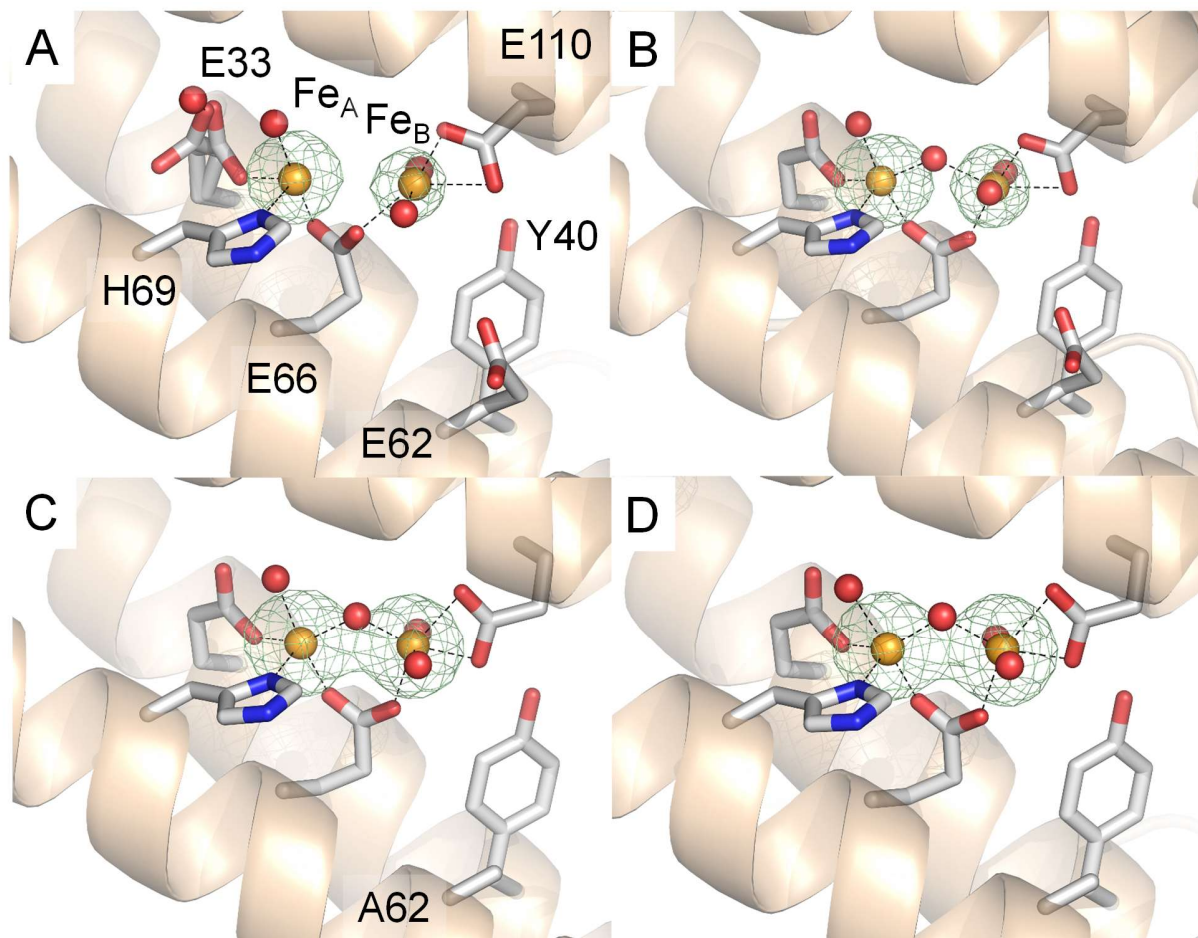
**Figure S1.** Comparison of the sequence of SynFtn with those of selected other ferritins. The area shaded in blue denotes those residues that comprise the 3-fold channel with the positions of the two conserved carboxylates of the animal proteins marked by blue triangles. The area shaded orange denotes the residues that comprise the 4-fold channel. Red shading indicates residues that line the B-channel of prokaryotic ferritins and green shading indicates the positions of the ‘transfer carboxylates’ identified in Frog M and Human H ferritin. The proteins shown are ferritin heavy chain from *Homo sapiens* (HumanH), ferritin middle subunit from *Rana catesbeiana* (FrogM), ferritin heavy chain from *Equus caballus* (HorseH), ferritin heavy chain from *Oryctolagus cuniculus* (RabbitH), ferritin heavy chain from *Mus musculus* (MouseH), ferritin 1 from *Glycine max* (Soybean), ferritin 1 from *Zea mays* (Maize), ferritin 1 from *Pisum sativum* (Pea), non-heme ferritin FtnA from *Escherichia coli* (Ecoli), non-heme ferritin from *Campylobacter jejuni* (Cjejuni), non-heme ferritin from *Helicobacter pylori* (Hpylori), ferritin from *Vibrio cholera* (Vcholerae) and ferritin from *Yersinia pestis* (Ypestis).



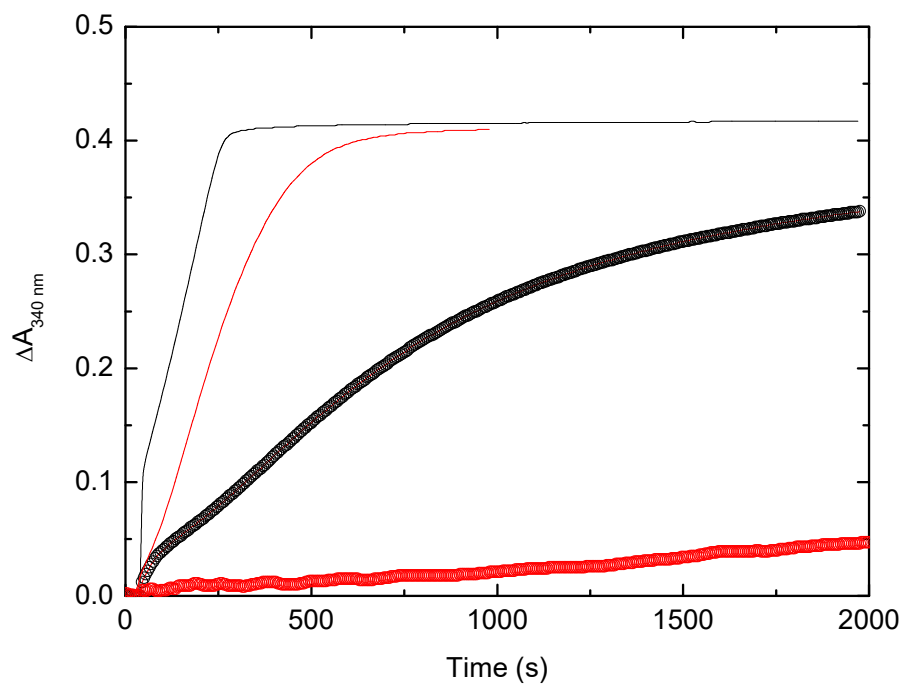
**Figure S2.** The B-channel of *SynFtn* is capped by an N terminal extension of the peptide chain. The image shows the B-channel of *SynFtn* formed at the intersection of three subunit monomers (PDB entry 6GKA) overlaid with an orange sphere at the position at which an iron ion was observed in the structure of E44Q *PmFtn* soaked overnight in a  $\text{Fe}^{2+}$  containing solution (PDB entry 4ZKH). The brown mesh represents the surface of the N terminal of *SynFtn*, which is an extension relative to *E. coli* FtnA, the most extensively characterized of the prokaryotic Ftn proteins.



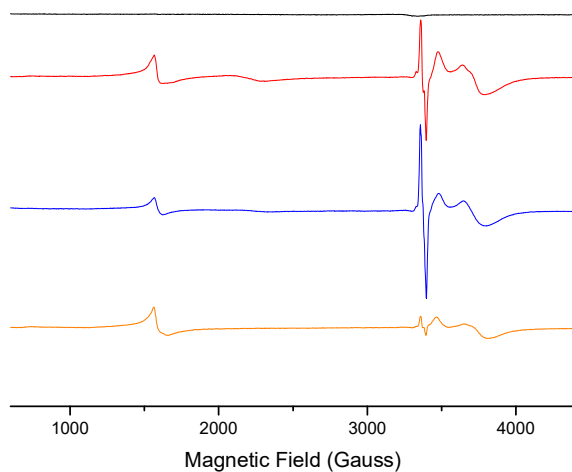
**Figure S3.** Comparison of the crystal structure of wild type and variant D137A SynFtn. Oxygen atoms coloured red, nitrogen blue, sulphur orange and carbon cyan (wild type) or white (D137A). The position of the mutated residue is marked by the red ellipse.



**Figure S4.** The ferroxidase center of Fe<sup>2+</sup>-soaked SynFtn variants. The di-iron site of D137A SynFtn following the soaking of crystals in a 5 mM Fe<sup>2+</sup> solution for either 2 min (A) or 20 min (B). (C-D) as (A-B) but for crystals of E62A SynFtn. Carbon is shown in white, nitrogen in blue, oxygen in red and iron as bright orange spheres with associated water molecules or (hydr)oxide ions as red spheres. The light green mesh in each of panels A to D shows the anomalous difference Fourier map calculated from data collected at the iron K-edge and contoured at 8  $\sigma$ .

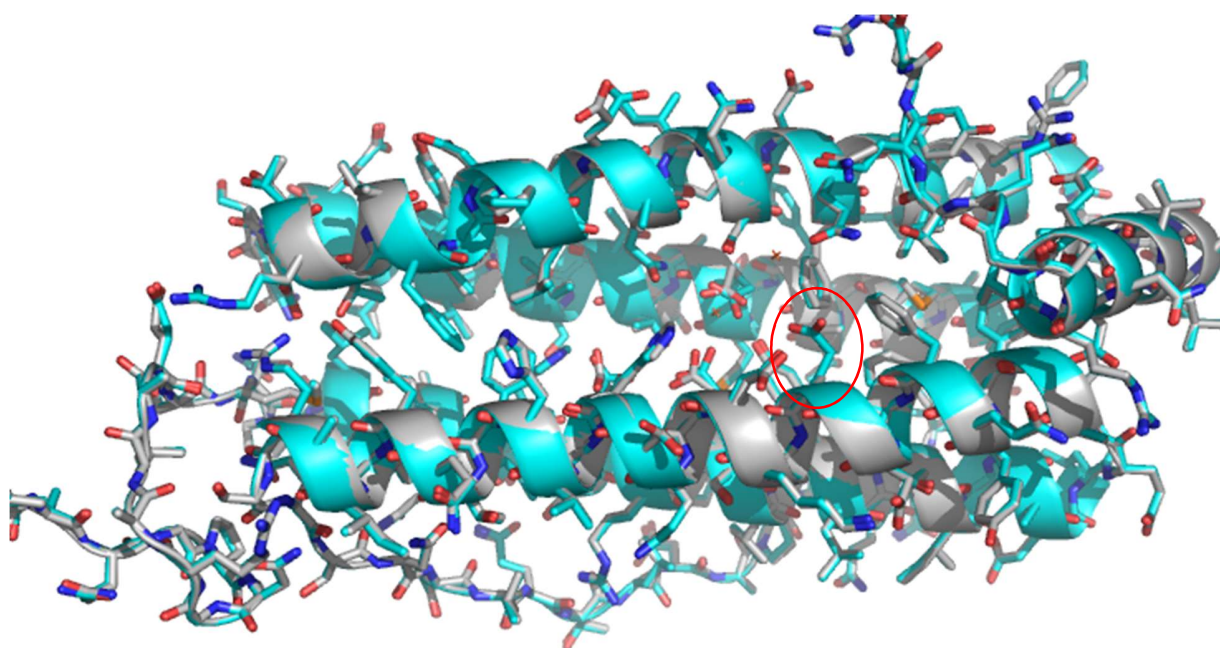


**Figure S5.** Inhibition of *SynFtn* by  $\text{Zn}^{2+}$ . The increase in absorbance at 340 nm as a function of time following the addition of 400 equivalents of  $\text{Fe}^{2+}$  to  $0.5 \mu\text{M}$  protein (24mer). The response of wild type protein is shown in black and variant D137A in red, solid lines represent data for addition of  $\text{Fe}^{2+}$  to apo proteins and open circles for addition of  $\text{Fe}^{2+}$  to protein pre-incubated with 96 equivalents of  $\text{Zn}^{2+}$  (4  $\text{Zn}^{2+}$ /monomer). All measurements were carried out in MES pH 6.5 at 25 °C.



**Figure S6.** EPR properties of *SynFtn* and variants . Full sweep EPR spectra of: wild type *SynFtn* prior to the addition of  $\text{Fe}^{2+}$  (black); wild type *SynFtn* frozen 9 s after the addition of 72 equivalents of  $\text{Fe}^{2+}$  (red); variant E62A *SynFtn* frozen 12 s after the addition of 72 equivalents of  $\text{Fe}^{2+}$  (blue); and, variant D137A *SynFtn* frozen 20 s after the addition of 72 equivalents of  $\text{Fe}^{2+}$  (orange). Note that the intensity of the radical signal centred on 3386 G is variable between preparations and does not correlate with the E62A and D137A substitutions.





**Figure S7.** Comparison of the crystal structure of wild type and variant E62A SynFtn. Oxygen atoms coloured red, nitrogen blue, sulphur orange and carbon cyan (wild type) or white (E62A). The position of the mutated residue is marked by the red ellipse.



**Table S1.** *SynFtn* D137A mutant. Data collection and refinement statistics.

<b>PDB code</b>	<b>6SOM</b>	<b>6SON</b>	<b>6SOO</b>
<b>Fe<sup>2+</sup> Soak (min)</b>	<b>0</b>	<b>2</b>	<b>20</b>
<b>Wavelength</b>	0.9763	0.9795	0.9795
<b>Resolution range</b>	40.55 - 2.15 (2.23 - 2.15)	51.07 - 1.60 (1.66 - 1.60)	44.12 - 1.57 (1.63 - 1.57)
<b>Space group</b>	F 4 3 2	F 4 3 2	F 4 3 2
<b>Unit cell</b>	176.8 176.8 176.8 90 90 90	176.9 176.9 176.9 90 90 90	176.5 176.5 176.5 90 90 90
<b>Total reflections</b>	223008 (22708)	609753 (61303)	852797 (83644)
<b>Unique reflections</b>	13408 (1302)	31808 (3112)	33428 (3301)
<b>Multiplicity</b>	16.6 (17.4)	19.2 (19.7)	25.5 (25.3)
<b>Completeness (%)</b>	99.81 (100.00)	99.93 (100.00)	99.96 (99.97)
<b>Mean I/sigma(I)</b>	10.25 (1.47)	26.10 (1.58)	27.06 (1.27)
<b>Wilson B-factor</b>	37.51	30.84	29.22
<b>R-merge</b>	0.1991 (1.465)	0.05239 (1.774)	0.06179 (2.264)
<b>R-meas</b>	0.2055 (1.509)	0.05386 (1.821)	0.06308 (2.31)
<b>R-pim</b>	0.05005 (0.3592)	0.01231 (0.4085)	0.01252 (0.4565)
<b>CC1/2</b>	0.997 (0.693)	1 (0.644)	1 (0.605)
<b>CC*</b>	0.999 (0.905)	1 (0.885)	1 (0.868)
<b>Reflections used in refinement</b>	13406 (1302)	31807 (3112)	33427 (3300)
<b>Reflections used for R-free</b>	677 (65)	1539 (153)	1671 (165)
<b>R-work</b>	0.1855 (0.2561)	0.1582 (0.2303)	0.1652 (0.2448)
<b>R-free</b>	0.2397 (0.3310)	0.1890 (0.2805)	0.1957 (0.2966)

<b>CC(work)</b>	0.957 (0.859)	0.968 (0.868)	0.959 (0.853)
<b>CC(free)</b>	0.946 (0.752)	0.948 (0.810)	0.956 (0.768)
<b>Number of non-hydrogen atoms</b>	1514	1630	1583
<b>Macromolecules</b>	1393	1416	1399
<b>Ligands</b>	1	3	2
<b>Solvent</b>	120	211	182
<b>Protein residues</b>	178	178	178
<b>RMS(bonds)</b>	0.006	0.005	0.005
<b>RMS(angles)</b>	0.71	0.69	0.67
<b>Ramachandran favored (%)</b>	98.86	100.00	100.00
<b>Ramachandran allowed (%)</b>	1.14	0.00	0.00
<b>Ramachandran outliers (%)</b>	0.00	0.00	0.00
<b>Rotamer outliers (%)</b>	0.00	0.66	0.00
<b>Clashscore</b>	1.83	1.79	0.73
<b>Average B-factor</b>	37.46	34.81	32.62
<b>Macromolecules</b>	36.95	33.02	31.18
<b>Ligands</b>	43.73	51.64	43.96
<b>Solvent</b>	43.35	46.58	43.56

Statistics for the highest-resolution shell are shown in parentheses.

**Table S2.** *SynFtn* E62A mutant. Data collection and refinement statistics.

<b>PDB code</b>	<b>6SOP</b>	<b>6SOQ</b>	<b>6SOR</b>
<b>Fe<sup>2+</sup> Soak (min)</b>	<b>0</b>	<b>2</b>	<b>20</b>
<b>Wavelength</b>	0.9795	0.9795	0.9795
<b>Resolution range</b>	40.58 - 1.93 (2.00 - 1.93)	36.13 - 1.67 (1.73 - 1.67)	39.46 - 1.74 (1.80 - 1.74)
<b>Space group</b>	F 4 3 2	F 4 3 2	F 4 3 2
<b>Unit cell</b>	176.9 176.9 176.9 90 90 90	177 177 177 90 90 90	176.5 176.5 176.5 90 90 90
<b>Total reflections</b>	128714 (12505)	211108 (13333)	194099 (16168)
<b>Unique reflections</b>	18381 (1789)	27809 (2527)	24709 (2426)
<b>Multiplicity</b>	7.0 (7.0)	7.6 (5.3)	7.9 (6.7)
<b>Completeness (%)</b>	99.82 (99.89)	98.90 (91.85)	99.87 (100.00)
<b>Mean I/sigma(I)</b>	11.11 (1.19)	22.75 (1.44)	15.09 (1.52)
<b>Wilson B-factor</b>	35.55	28.17	30.79
<b>R-merge</b>	0.1016 (1.327)	0.04735 (0.9856)	0.06909 (0.9414)
<b>R-meas</b>	0.1098 (1.435)	0.05083 (1.093)	0.07412 (1.021)
<b>R-pim</b>	0.04087 (0.5372)	0.01811 (0.4579)	0.02631 (0.3896)
<b>CC1/2</b>	0.998 (0.49)	0.999 (0.496)	0.998 (0.581)
<b>CC*</b>	0.999 (0.811)	1 (0.814)	1 (0.857)
<b>Reflections used in refinement</b>	18379 (1789)	27808 (2526)	24708 (2426)
<b>Reflections used for R-free</b>	931 (90)	1398 (120)	1248 (119)
<b>R-work</b>	0.1550 (0.2735)	0.1511 (0.2555)	0.1575 (0.2304)
<b>R-free</b>	0.1998 (0.3269)	0.1830 (0.3370)	0.1873 (0.2812)

<b>CC(work)</b>	0.967 (0.802)	0.969 (0.810)	0.968 (0.844)
<b>CC(free)</b>	0.964 (0.755)	0.945 (0.645)	0.966 (0.836)
<b>Number of non-hydrogen atoms</b>	1583	1637	1600
<b>Macromolecules</b>	1395	1398	1395
<b>Ligands</b>	1	4	4
<b>Solvent</b>	187	235	201
<b>Protein residues</b>	178	178	178
<b>RMS(bonds)</b>	0.007	0.007	0.007
<b>RMS(angles)</b>	1.01	1.09	1.02
<b>Ramachandran favored (%)</b>	99.43	99.43	100.00
<b>Ramachandran allowed (%)</b>	0.57	0.57	0.00
<b>Ramachandran outliers (%)</b>	0.00	0.00	0.00
<b>Rotamer outliers (%)</b>	0.00	0.00	0.00
<b>Clashscore</b>	1.46	2.18	1.46
<b>Average B-factor</b>	37.11	30.13	33.13
<b>Macromolecules</b>	35.79	28.09	31.68
<b>Ligands</b>	39.32	30.23	32.88
<b>Solvent</b>	46.95	42.27	43.21

Statistics for the highest-resolution shell are shown in parentheses.

**Table S3.** Refined fractional occupancies of metal binding sites in *SynFtn* mutants. Soak solutions comprised the well solution with 5 mM Fe<sup>2+</sup> and the pH adjusted to 6.5. Crystals were soaked for either 2 or 20 min prior to freezing.

<i>SynFtn</i> variant	Soak/ min	Fractional occupancy <sup>a</sup>		
		Site A	Site B	Site 3FC
D137A	2	0.50	0.32	0
	20	0.56	0.54	0
E62A	2	0.74	0.51	0.26
	20	0.92	0.88	0.33

<sup>a</sup>Sites A and B constitute the ferroxidase centre. Site 3FC represents the iron binding site which lies on the symmetry axis in the 3-fold channel of the iron-soaked wild type ferritin structure (PDB entry 3OUY), and has a maximal occupancy of 0.33