Author Queries

Journal: Philosophical Transactions of the Royal Society B

Manuscript: rstb20110415

Q1 Please check the edit of affiliations.

Q2 Please check the representation to heading levels.

Q3 Please provide expansion of the acronym CRDS.

Q4 Please provide volume and page range for reference [41].

Q5 Please provide volume number for reference [79].

PHILOSOPHICAL TRANSACTIONS OF THE ROYAL SOCIETY B

Phil. Trans. R. Soc. B (2012) **00**, 1–12
doi:10.1098/rstb.2011.0415

Introduction

Biological sources and sinks of nitrous oxide and strategies to mitigate its emissions

Andrew J. Thomson[1,*], Georgios Giannopoulos[1], Jules Pretty[3], Elizabeth M. Baggs[2] and David J. Richardson[1]

[1]*School of Biological Sciences, Norwich Research Park, University of East Anglia, Norwich NR4 7TJ, UK*
[2]*School of Biological Sciences, University of Aberdeen, Aberdeen AB24 3UU, UK*
[3]*School of Biological Sciences, University of Essex, Wivenhoe Park, Essex CO4 3SQ, UK*

Nitrous oxide (N_2O) is a powerful atmospheric greenhouse gas and cause of ozone layer depletion. Global emissions continue to rise. More than two-thirds of these emissions arise from bacterial and fungal denitrification and nitrification processes in soils, largely as a result of the application of nitrogenous fertilizers. This article summarizes the outcomes of an interdisciplinary meeting, 'Nitrous oxide (N_2O) the forgotten greenhouse gas', held at the Kavli Royal Society International Centre, from 23 to 24 May 2011. It provides an introduction and background to the nature of the problem, and summarizes the conclusions reached regarding the biological sources and sinks of N_2O in oceans, soils and wastewaters, and discusses the genetic regulation and molecular details of the enzymes responsible. Techniques for providing global and local N_2O budgets are discussed. The findings of the meeting are drawn together in a review of strategies for mitigating N_2O emissions, under three headings, namely: (i) managing soil chemistry and microbiology, (ii) engineering crop plants to fix nitrogen, and (iii) sustainable agricultural intensification.

Keywords: nitrous oxide; denitrification; greenhouse gas; climate change; mitigating emissions

1. INTRODUCTION

Nitrous oxide (N_2O) is a colourless, non-toxic gas, commonly known as laughing gas. Since its discovery over 200 years ago, it has found use both as an anaesthetic and a fuel additive. However, in 1908, the invention of the Haber–Bosch process, allowing the abiological reduction of atmospheric nitrogen to ammonia (NH_3; called nitrogen fixation), gave rise to the introduction of synthetic nitrogen-based fertilizers that has enabled dramatic increases in intensive farming. This, in turn, has led to increasing N_2O emissions from the increased presence of reactive nitrogen in soil [1,2]. The deposition of nitrogen from motor vehicles, especially near busy roads, means that fossil fuels are also a major contributor to soil nitrogen levels [3]. The return of animal waste to soil and wastewater treatment further contribute to N_2O emissions [4,5]. The cumulative effect over the past century has been an estimated approximately 20 per cent increase in atmospheric N_2O concentration that is still increasing at a rate of 0.2–0.3% yr^{-1} [6]. More than two-thirds of these emissions come from bacterial and fungal respiratory processes in soils, broadly termed denitrification and nitrification [1,2]. Figure 1 illustrates the proportions of total global nitrous oxide emitted by various sources, including human activities.

N_2O is a powerful greenhouse gas (GHG) with an atmospheric lifetime of 114 years [7]. Although N_2O only accounts for around 0.03 per cent of total GHG emissions, it has an almost 300-fold greater potential for global warming effects, based on its radiative capacity, compared with that of carbon dioxide (CO_2) [7]. Hence, when the impact of individual GHGs on global warming is expressed in terms of the Intergovernmental Panel on Climate Change approved unit of CO_2 equivalents, N_2O accounts for approximately 10 per cent of total emissions [6].

In the stratosphere, the main sink for N_2O, ultraviolet photochemistry oxidizes NO_x [8]. Today, N_2O is a major cause of ozone layer depletion [9]. Since 1997, many of the non-biological emissions of N_2O, for example, those associated with the transport industry, have been systematically lowered, whereas emissions from agriculture are essentially unchanged [7]. Although the 1997 Kyoto Protocol set emission limitations and reduction obligations, with respect to a basket of six gases, including N_2O, on its signatories this Protocol expires in 2012. It is crucial that its successor is able to address fully the issue of soil-derived N_2O

* Author for correspondence (a.thomson@uea.ac.uk).

One contribution of 12 to a Theo Murphy Meeting Issue 'Nitrous oxide: the forgotten greenhouse gas'.

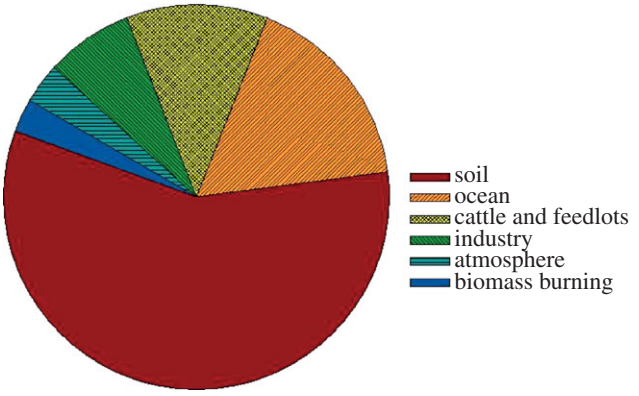

Figure 1. Proportions of total global nitrous oxide emitted by various sources and human activities. Adapted from data in the Contribution of Working group III to the fourth assessment report of the intergovernmental panel on climate change, 2007. Eds B. Metz, O. R. Davidson, P .R. Bosch, R. Dave and L. A. Meyer. Cambridge, UK; New York, NY: Cambridge University Press.

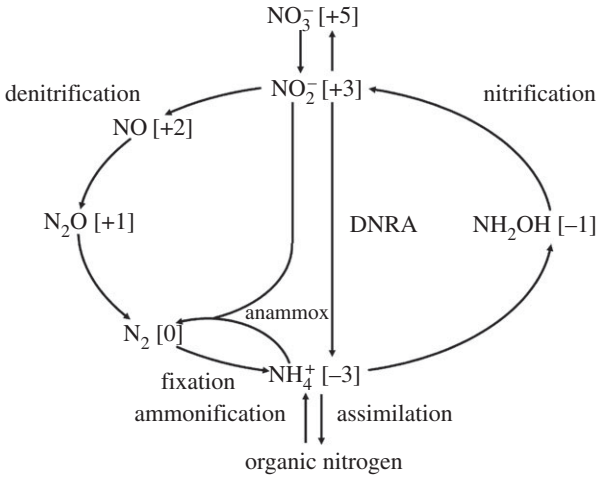

Figure 2. The microbiological nitrogen cycle. Shown are the several microbial processes that respire or assimilate nitrogen (the oxidation states of N are given in parentheses). The name of each process is indicated. Nitrous oxide (N_2O) is an intermediate in denitrification. The anammox reaction, used in wastewater treatment plants, is the catabolism between ammonia and nitrite to yield nitrogen gas, e.g. $NH_4^+ + NO_2^- \rightarrow N_2 + 2H_2O$.

emissions. Because of the ongoing decline of chlorofluorocarbons and the continuous increase of N_2O in the atmosphere, the contributions of N_2O to both the greenhouse effect and ozone depletion will be even more pronounced in the twenty-first century [9,10].

The availability of nitrogen (nitrate or ammonium) as well as phosphorus (phosphate) and potassium is crucial determinants of globally sustainable crop yields. There is widespread nitrogen and phosphate deficiency and thus potential yields are often not reached. This deficiency is particularly acute in the developing world where the need to apply nitrogen fertilizer or encourage biological nitrogen fixation will certainly increase. In these systems, the primary aim is food security, but with it will, undoubtedly, come yet further increases in N_2O emissions. Thus, the environmental damage from the further intensification of agriculture will increase more rapidly unless means can be found to mitigate the emissions of biologically derived N_2O [11].

Between 23 and 24 May 2011, a residential scientific meeting, entitled 'Nitrous oxide (N_2O) the forgotten greenhouse gas', was held at the Kavli Royal Society International Centre, Chicheley Hall, Buckinghamshire, UK. The objective of this meeting was to bring together scientists from a wide range of disciplines, including biochemists, chemists, molecular biologists, geneticists, microbiologists, soil scientists, ecologists and environmental scientists to discuss four areas, namely: (i) biological sources of N_2O emissions and the consequent problems; (ii) biological production and consumption of N_2O; (iii) measuring and modelling N_2O balances; finally (iv) strategies for mitigating N_2O emissions. The papers published in this themed volume of *Philosophical Transactions of the Royal Society B* were presented and discussed at this meeting.

This paper provides an introduction and background to the nature of the problem of the biological sources of N_2O, exploring the biological sources and sinks of N_2O from different environments such as oceans, soils and wastewaters, and describes the genetic regulation and molecular details of the enzymes responsible. The techniques for measuring and assessing the amounts of N_2O to provide global and local budgets are discussed. A summary is provided of the main conclusions reached by the papers in this issue. Finally, these findings are drawn together in a discussion of strategies to mitigate N_2O release.

2. THE NITROGEN CYCLE

Nitrogen gas (N_2), present at 78.08 per cent (v/v) in the atmosphere, possesses one of the most stable chemical linkages known, namely, a chemical triple bond that requires almost 10^3 kJ M^{-1} of energy to break into its component N atoms. The triple bond of N_2 also has a very high-energy barrier towards breaking, necessitating the use of highly effective catalysts, or enzymes, to speed up the scission process. All biological organisms require nitrogen to synthesize amino acids, proteins, nucleic acids and many additional cofactors. The total nitrogen combined in biology originates from the atmosphere to where it is ultimately returned as the gas, N_2. Figure 2 shows the best known, arguably, of all elemental cycles, the nitrogen (or N−) cycle. Nitrogen is driven through all its accessible redox states from the most strongly reduced state, as $[NH_3]$, in the −3 oxidation state, to the most highly oxidized state, nitrate ion, $[NO_3]^-$, in the +5 oxidation state. Various species with intermediate oxidation states are produced such as nitrite ion, $[NO_2]^-$, the gases nitric oxide, [NO] and nitrous oxide $[N_2O]$. They arise through the actions of a number of biological processes the most prominent of which are termed nitrogen fixation, nitrification, dissimilatory nitrate reduction to ammonia (DNRA, or nitrate ammonification), anaerobic ammonia oxidation (Anammox) and denitrification. Ammonium ion, $[NH_4]^+$, availability is the net result of immobilization, mineralization and nitrification

Table 1. The genes and enzymes that carry out the bacterial nitrogen cycle.

transformation	genes	encoding enzyme	references
$N_2 \rightarrow NH_3$	*nifHDK*	nitrogenase	[13]
$NO_3^- \rightarrow NO_2^-$	*narG*	dissimilatory nitrate reductase	[14]
$NO_2^- \rightarrow NO$	*nirS*, *nirK*	nitrite reductase haem cd_1 and copper nitrite reductase	[15]
$NO \rightarrow N_2O$	*norCB*	nitric oxide reductase	[16,17]
$N_2O \rightarrow N_2$	*nosZ*	nitrous oxide reductase	[18–20]
NH_4^+ oxidation	*amo*, *hao*	ammonia monooxygenase, hydroxylamine oxidoreductase	[21,22]
NO_3^- assimilation	*narB*, *nasA*	assimilatory nitrate reductase	[23]
NO_2^- assimilation	*Nir*	assimilatory nitrite reductase	[24]
NH_3 assimilation	*glnA*	glutamine synthetase	[25]
organic N metabolism	*ure*	urease	[26]

[12]. Table 1 lists all the enzymes and the genes that carry out the nitrogen cycle.

Atmospheric N_2 is fixed into NH_3 only by free-living and symbiotic bacteria and archaea (diazotrophs). Nitrogenase is the universal catalyst that breaks the triple bond to produce NH_3. There are three known variants of the nitrogenase enzyme, all possessing complex, unique iron and sulphur clusters one of which contains an additional metal ion, being molybdenum, iron or vanadium, in each variant. The ammonium ion can be oxidized to the nitrate ion $[NO_3]^-$ in a three-step process called nitrification, the first of which is catalysed by the enzyme, ammonia monooxygenase (AMO). $[NO_2]^-$ and $[NO_3]^-$ ions generated from nitrification may then be reduced either during DNRA or denitrification.

The main routes of N loss are by soil erosion, leaching, ammonia volatilization, ammonia oxidation and denitrification. Approximately, 62 per cent of total global N_2O emissions is thought to be emitted from natural and agricultural soils (6 and 4.2 Tg N yr^{-1}, respectively) [27,28] mainly owing to bacterial denitrification and ammonia oxidation, the first step in nitrification [2,29]. The other third of N_2O emissions comes from the ocean via nitrification and denitrification [30]. Further anthropogenic sources of N_2O include the production nitric acid, power plants (fossil fuelled) and vehicle emissions [10]. These emissions are responsible for an 18 per cent increase of atmospheric N_2O since the early 1900s [1] and are still increasing at a rate of 0.25 per cent per year [1,7].

Denitrification is the stepwise reduction of $[NO_3]^-$ to N_2 by four enzymes each generating intermediate products, namely, nitrite ion $[NO_2]^-$, NO and N_2O. $[NO_3]^-$ can also be reduced during nitrate ammonification to $[NH_4]^+$ via $[NO_2]^-$, with N_2O being produced. Anammox is the process by which $[NO_2]^-$ is reduced to N_2 using $[NH_4]^+$ as an electron donor. The ability to denitrify is phylogenetically diverse, and can even be undertaken by microbes traditionally classified as belonging to a different functional group. For example, ammonia-oxidizing bacteria are also able to denitrify, reducing $[NO_2]^-$ ion, sometimes referred to as nitrifier denitrification. N_2O is also produced as a by-product during ammonia oxidation, the first step of nitrification. Two further major biological processes of nitrogen transformation are immobilization (or assimilation), the uptake of nitrogen by micro-organisms and its conversion to organic nitrogen, and mineralization or ammonification, the conversion of organic nitrogen to $[NH_4]^+$ [2,31].

While there are several enzymological pathways in fungi and bacteria that generate N_2O, there is only one enzyme known that converts N_2O to gaseous nitrogen, N_2, namely, nitrous oxide reductase (N_2OR) [2]. Failure of this enzyme to operate leads, for example, to the termination of the bacterial denitrification process at N_2O rather than N_2. This may be the key to the understanding of, and possible intervention in, the increased emissions of N_2O as intensification of agriculture has tended to take place through the increased application of nitrogenous fertilizers [18].

3. NITROUS OXIDE EMISSION FACTORS

The upward trend in the atmospheric concentration of N_2O over the 140 years between 1860 and 2000 from all sources has been well documented. Smith *et al.* [32] summarize the historical evidence and have now been able to account satisfactorily for the rises. They compare the amounts of new reactive N entering agricultural systems globally with the total emission of N_2O, expressing the ratio of these two as an N_2O emission factor (EF). This reactive N includes N newly fixed as synthetic fertilizer, and biologically fixed N, and also N mineralized from soil organic matter (SOM) when natural land is converted to agriculture [5] and NO_x deposition. The historical upward trend observed in the atmospheric concentration of N_2O can then be very closely matched with an overall EF close to 4 per cent. Thus, they have clearly shown that agriculture is the activity mainly responsible for the additional N_2O emissions over the last century and a half. They also apply their methodology to analyse N_2O emissions arising from biofuel production and reach the conclusion that, when rapeseed and corn, which require nitrogenous fertilizer, are used to produce biodiesel and bioethanol, the N_2O emitted could cause as much, or more, global warming as that avoided by replacement of the fossil fuel by biofuel. It is, therefore, important to avoid biofuel production based on crops with a high N demand but to use those that can be grown with little, or no, fertilizer N requirement such as willow and *Miscanthus*, the so-called 'second generation' biofuel crops.

Skiba *et al.* [33] argue that, especially in the agricultural sector, an EF can be too simplistic to reflect local

variations in climate, ecosystems and management, and should not, therefore, be used to take account of the effects of any mitigation strategies. This paper examines deviations of observed N_2O emissions from those calculated using the simple EF for all anthropogenic sources and strongly advocates the need to adopt specific EFs that reflect regional variability in climate, soil type and management. Although they can show how bottom-up emission inventories can be verified by top-down modelling they conclude that, in spite of the wealth of N_2O emission measurements of the last 20 years, there are still not enough long-term datasets to provide the information needed to design EFs for different climate zones or soil types.

4. BIOLOGICAL PRODUCTION AND CONSUMPTION OF NITROUS OXIDE

(a) *Enzymological aspects*

N_2O is produced by both fungi and certain classes of bacteria, including those living in soils and in the oceans, as part of their respiratory processes to generate energy. A recent study revealed that archaeal nitrification is dominating the N_2O production in the ocean [34]. The possible contribution of the archaea to N_2O production in terrestrial systems, however, is as yet unknown.

Shoun & Tanimoto [35] were the first to identify fungal (eukaryotes) denitrifying activities previously thought to be restricted only to bacteria (prokaryotes). Shoun *et al.* [36] review the fungal denitrification system. It comprises a copper-containing nitrite reductase (NirK) and a cytochrome P450 nitric oxide reductase (P450nor) that together reduces nitrite to N_2O. The system is localized in mitochondria that are also able to function during anaerobic respiration. Some fungal systems use dissimilatory and assimilatory nitrate reductases to denitrify nitrate. Phylogenetic analysis of *nirK* genes showed that the fungal denitrifying system has the same ancestor as the bacterial counterpart, and thus probably originates from the proto-mitochondrion. Fungal denitrification is often accompanied by co-denitrification, in which a hybrid N_2O species is formed upon the combination of the nitrogen atoms of nitrite with nitrogen donors such as amines and imines. The final product of fungal denitrification is N_2O, because the enzyme N_2OR is absent. Hence, fungal denitrification, under certain conditions, is expected to be a major source of N_2O emissions. Shoun notes that acidification of environments, for example, by acid rain and excessive use of ammonia fertilizer, promote fungal activity resulting in further increases in N_2O emissions. Prendergast-Miller *et al.* [37] have recently shown Ectomycorrhizal fungal spp. to possess the ability to produce N_2O, suggesting that they may have a significant, but as yet unexplored, role in N_2O production in forest ecosystems. Recent advances in isotopomer approaches promise the ability to be able to estimate the partition between fungal and bacterial N_2O production *in situ*, and to allow estimates of the significance of fungal denitrification across a range of ecosystems [38].

The major contributor to the biological production of N_2O in many environments is the respiratory NO reductase (NOR) found in denitrifying bacteria and in some ammonia-oxidizing organisms. Recently, the molecular structure of this enzyme, the bacterial nitric oxide reductase cNOR from *Pseudomonas aeruginosa*, has been solved by Shiro *et al.* [39]. Since 1971, the NO reduction activities of the bacterial membrane-bound NORs have been reported for many bacteria. Although there can be wide variations in the electron-donating moiety, the structures of the catalytic domains are invariant consisting of 12 transmembrane helices that bind one low-spin haem plus a high-spin haem that is adjacent to a non-haem iron centre, called Fe_B. The dinuclear pair (the haem iron and Fe_B) binds and activates two NO molecules forming the N–N bond of N_2O. Shiro *et al.* [39] discuss a number of possible mechanisms for this reaction.

An intriguing evolutionary aspect of this study is confirmation of the long suspected close structural similarity between NOR and the main subunit of aerobic and micro-aerobic cytochrome oxidases (COX) that reduce oxygen to water in an energy conserving reaction that is tightly linked to the translocation of protons across a membrane. In this case, the high-spin haem is adjacent to a copper ion (Cu_B) that has replaced the Fe_B. These structural differences between cNOR and COX observed in the catalytic centre, and the delivery pathway of the catalytic protons, clearly reflect the functional differences between these respiratory enzymes. NOR, and hence N_2O production, is thought to have preceded COX, and oxygen reduction, on the evolutionary timescale, consistent with the dramatic rise of oxygen in the Earth's atmosphere around 3.5 Ga [40].

Another source of nitrous oxide is from nitrate-ammonifying (DNRA) bacteria [41]. It is now recognized that DNRA bacteria such as *Salmonella* and *Escherichia coli* can produce NO as a side product of nitrate metabolism. This endogenous NO can lead to de-repression of genes encoding systems that are concerned with the detoxification of NO and the repair of proteins potentially damaged by this cytotoxin. One regulator that mediates this de-repression is the NO-binding protein NsrR. In *E. coli*, NsrR regulates some 20 genes, including the flavohaemoglobin (Hmp) which converts NO to N_2O under anoxic conditions [42].

In contrast to the multiplicity of mechanisms by which N_2O can be generated, only a single dominant sink for N_2O is known, the respiratory N_2O reductase (N_2OR) typically found in denitrifying bacteria that reduce N_2O to N_2. N_2OR is a homo-dimeric protein containing two structurally distinct copper cofactors per monomer that are crucial for activity, namely: Cu_Z and Cu_A [43]. These copper cofactors are inserted only into the apo-protein when it has been translocated from the cytoplasm to the periplasm [44]. Hence, severe copper depletion can lead to enzyme inactivation [45]. In N_2OR itself, the catalytic state seems chemically fragile. For example, it loses activity if exposed even briefly to oxygen. The fragility of N_2OR likely depends on the chemical nature of the Cu–S cluster of the catalytic centre. For many years,

biochemists have known that Cu_Z can adopt different oxidation states and stabilities, as evidenced by changes in colour, that depend on the previous history of exposure of either the cell or the enzyme itself to oxygen. The paper from Dell'Acqua *et al.* [46] presents evidence on the N_2OR purified from the marine organism *Marinobacter hydrocarbonoclasticus* that the catalytic centre, Cu_Z, can adopt different oxidation states. One form, Cu_Z*, [$1Cu^{2+}:3Cu^{+}$], is redox inert and, hence, enzymatically inactive. However, they have shown that it can be reactivated slowly by incubation for many minutes under non-physiological, highly reducing conditions. A so-called purple form in which the Cu_Z centre is in the oxidized, redox state [$2Cu^{2+}:2Cu^{+}$] is generated that can subsequently be reduced to the [$1Cu^{2+}:3Cu^{+}$] state. However, none of these redox states is a high-activity state. The high-activity state is reached only after complete reduction of the Cu_Z centre to an all-Cu(I) form [$4Cu^{+}$]. However, very recent structural evidence [47] reveals a form of the enzyme that, unexpectedly, contains the Cu_Z cluster in the form [Cu_4S_2], whereas the previous X-ray structures of the low-activity state [$1Cu^{2+}:3Cu^{+}$, S] show the cluster to contain only one sulphide ion, [Cu_4S]. One can speculate that the reductively reactivated, high-activity enzyme may well contain the [Cu_4S_2] cluster.

Within the cell, the maintenance of high-activity N_2OR, or recovery of activity, say, after transient exposure to oxygen, is likely due to ancillary proteins that insert the copper cofactors into the apo-protein and are known to be required for N_2OR activity. Thus, supply of sulphur and electrons is a requirement. In the *nos* gene cluster, there is a putative ABC transporter (possibly of sulphur), consisting of NosD, NosF and NosY [48]. In addition, the operon encodes a Cu chaperone, NosL. The membrane-bound regulator NosR, required for operon expression, appears to contain redox centres, including FeS clusters (perhaps for electron supply). Thus, the biosynthesis of N_2OR and the maintenance of its reductase activity requires these ancillary proteins. These are all points of vulnerability that can lead to inactivation of N_2OR and, hence, lead to release of gaseous N_2O. A clearer understanding of these processes, their regulation and operation will help define the optimal environmental conditions for maintenance of the activity of N_2OR and hence the encouragement of the release of N_2 rather than N_2O.

(b) *Microbiological aspects*

Of the many factors that contribute to the emission of N_2O from bacterial populations, one important determinant is the cellular abundance and another is the activities of the enzymes that produce and consume N_2O [42]. Enzyme abundance is governed by expression of the corresponding genes of regulatory systems and signal transduction pathways that respond to intra- or extracellular signals. Because N_2O is relatively inert at ambient temperature, and is not a potent toxin, micro-organisms can tolerate relatively high concentrations (millimolar). N_2O does not, therefore, appear to be a signal that regulates the expression of any of the denitrification genes. From the point of view of mitigating N_2O release from denitrification, the absence of regulation by N_2O is a significant observation, because denitrifying populations do not apparently respond to N_2O accumulation by making more of the N_2OR. The expression of the genes encoding the enzymes that produce and consume N_2O is regulated by environmental signals, typically oxygen and NO, acting through regulatory proteins, which, either directly or indirectly, control the frequency of transcription initiation. Because denitrification is an anaerobic respiration, it makes good physiological sense for denitrification genes to be upregulated by low-oxygen concentrations. NO is an intermediate of the pathway, and is somewhat toxic. Regulation of denitrification gene expression by NO is therefore presumed to be a mechanism to coordinate NO production and consumption so as to avoid its accumulation to toxic levels.

Bakken *et al.* [49] nicely expand these points. For instance, in various mutants of *Paracoccus denitrificans*, the transcription of *nosZ*, that codes for N_2OR, is equally effective with *FnrP* that responds to oxygen depletion or *NNR*, responding to NO. In *P. denitrificans*, N_2OR is expressed much earlier than nitrite reductase (NIR) and NOR in response to low oxygen. Moreover, only a fraction of the cells are able to express NIR and NOR before all the oxygen has been depleted. In contrast, nearly 100 per cent of the cells appear to express N_2OR, as judged from the rate of reduction of externally supplied N_2O. The denitrification phenotype of *P. denitrificans* at pH 7 demonstrates highly efficient reduction of NO_x all the way to N_2, with only minor emissions of either NO or N_2O. Bakken wryly observes that if the denitrifying communities of soils performed equally well, their contribution to emission of NO and N_2O would be negligible. Although the performance of *P. denitrificans* appears to be exceptional, the soil bacterium *Agrobacterium tumefaciens* is unable to reduce N_2O to N_2 because it lacks *nosZ*. Indeed, strains which lack *nosZ* occur within many genera of denitrifying prokaryotes, and if organisms with such a truncated denitrification apparatus were to dominate in soils, it would lead to high $N_2O/(N_2+N_2O)$ product ratios of denitrification. Bakken, therefore, proposes the term 'denitrification regulatory phenotype', that is a set of variables characterizing the organism's ability to perform a balanced and effective transition from oxic to anoxic respiration with only marginal emissions of intermediates. This rather detailed understanding of the bacterial nitrogen cycle to date has come from studies of Gram-negative bacteria but evidence is now appearing showing that Gram-positive bacteria, such as Bacilli, can also carry out denitrification [50,51].

5. NITROUS OXIDE EMISSIONS FROM SOILS

It is now well-recognized that microbial activity in soils is a major contributor to atmospheric loading of N_2O. Clark *et al.* [52] have assessed the influence of different long-term fertilization and cultivation treatments in a 160 year-old field experiment, comparing the potential for denitrification with the size and diversity of the soil denitrifier communities. Denitrification potential was

found to be much higher in soil from an area left to develop from arable into woodland than from a farmyard manure-fertilized arable treatment, which in turn was significantly higher than inorganic nitrogen-fertilized and unfertilized arable plots. These observations correlated with abundance of *nirK* but not *nirS* (dissimilatory nitrite reductase genes). Most genetic variation was seen in *nirK* where sequences resolved into separate groups according to soil treatment. They conclude that bacteria-containing *nirK* is most likely responsible for the increased denitrification potential associated with nitrogen and organic carbon availability in this soil. Soil physico-chemical properties (bulk density, pH, organic matter, organic C, N and C : N ratio) have an overriding influence on the potential denitrification activity resulting in increased N_2O emissions in soils with high organic matter. Significantly, there were also structural differences in denitrifier communities in soils with high N and C contents. Thus, they possess proportionally fewer copies of the N_2OR gene *nosZ*, so may be less able to close the nitrogen cycle by reducing N_2O to N_2. They also note that soil management (tillage) can lower GHG emissions.

Bakken *et al*. [49] report that, in model strains of *P. denitrificans* in pure cultures and in microbial communities extracted from soils, the $N_2O/(N_2+N_2O)$ product ratio of denitrification is controlled by pH. The ratio increases with acidity. The effect is probably due primarily to interference with the assembly of the enzyme N_2OR, rather than to the narrow pH range of the maximal activity of the enzyme. There have been many similar observations of pH effects on denitrification in soils indicating a wide generality of the phenomenon [53–58]. These findings suggest that the continuing acidification of agricultural soils through excessive use of nitrogen fertilizers, as demonstrated for China [59] will enhance N_2O emissions drastically. It is proposed that careful adjustment of pH in agricultural soils, say, by liming, should reduce N_2O emissions from slightly acid soils. This needs to be tested rigorously in field trials.

Plants themselves have a strong influence on the microbial community of the rhizosphere, where most of the N_2O generating activity occurs. The release of plant-derived low molecular weight organic compounds into the soil enhances heterotrophic activity, with denitrifiers and nitrate ammonifiers thought to compete for this carbon. Hence, N_2O production and reduction rates are often positively correlated with total carbon or soluble organic carbon availability [60,61]. There is currently interest in understanding the physiological and genetic bases underpinning the influence of plant traits in regulating N_2O emission, and the possibility that this could inform future breeding programmes to couple enhanced crop agronomic performance with environmental sustainability in terms of lowering net GHG emissions and increasing soil carbon stocks.

Denitrification enzymes require a variety of metal cofactors, including Mo, Fe, Cu and Zn. The absolute requirement of N_2OR for Cu (and sulphur) for activity, as well as the absence of any parallel pathways that can reduce N_2O, accounts for the critical role of this element in the success of this final step of denitrification. Many species of bacteria have scavenging systems, such as siderophores, excreted by cells to chelate Fe strongly in order to extract it from soils, or sequester it from the ocean, and to deliver Fe(II) to cell surface receptors for active uptake into the cell. Furthermore, Fe can also be stored within cells inside proteins, such as ferritins, for retrieval in times of external Fe stress (or to compartmentalize the Fe during dormancy to protect it from reacting with O_2, thereby generating products potentially toxic to DNA). There are no such sequestering or storage systems yet known for copper in bacteria with the exception of some methanotrophic bacteria that excrete Cu-chelating compounds [62]. Hence, copper availability to the cell depends on the concentrations of Cu in the local external environment as well as on its state of chelation within soils. Zumft [2] first showed that by growing laboratory cultures of denitrifying bacteria in Cu-deficient media high levels of N_2O emissions occur compared with those in copper sufficient media leading him to the conclusion that N_2OR is a copper-dependent enzyme.

Copper in soil is found as the water-soluble cation Cu^{2+} but in reducing soils as the insoluble ion, Cu^{1+}. Soil bacteria can take up Cu^{2+} or Cu^{+} either by energized or diffusive transport [63]. The biological availability of Cu in soils to crops is influenced by a number of factors: its chemical state, soil conditions (pH, redox, soil moisture, etc.), SOM, inputs (fertilizer, manure, animal feed, etc.), weather, crop type and maturity. Cu deficiency is often observed in alkaline soils. A negative correlation of Cu plant uptake and pH is seen in clay soils. Cu bioavailability is also lowered by adsorption of Cu on clay surfaces or, in soils with high organic matter such as humic acids, formation of metal–organic complexes [64].

However, free Cu^{2+} species can also be toxic to soil bacteria. Ore *et al.* [65] correlated copper toxicity in *Nitrosomonas europaea* to free ion metal activity in soil pore water; EC_{50} $Cu^{2+} = 2 \times 10^{-6}$ to 2×10^{-9} M. Two major uncertainties exist regarding the interaction of bacteria and free metal ions. First, not all soil bacteria have the same tolerance to free ion metals and microbial communities can adapt during long-term exposure, developing pollution-induced community tolerance and, second, it is difficult to assess which bacterial cells are exposed to the free metal ions in the soil matrix. Thus, in Cu-limiting conditions, it was recently demonstrated that the bacterium *P. denitrificans* is able to acquire Cu from the soil matrix by excreting zinc coproporphyrin III in both aerobic and anaerobic environments [66].

Cu is also a required cofactor in $[NO_2]^-$ reduction in some bacteria such as *Achromobacter xylosoxidans*. In a large-scale field study, Enwall *et al*. [67] found a positive relationship between soil Cu content and the abundance of *nirK* genes. Thus, Cu plays a key role in both NO_2^- and N_2O reduction [68]. With approximately 40 per cent of Europe's arable soils being Cu deficient (less than 2 mg Cu kg^{-1}), the potential for N_2O mitigation (with a simultaneous crop yield increase) is high. Nevertheless, above certain concentrations metals in the soil can have adverse effects on

soil nutrient cycling and soil food webs [69]. Investigating the trade-off between the effect of mineral micronutrients on N_2O soil emissions and soil ecosystem functioning (nutrient cycling) is an important aspect with practical and environmental implications yet to be explored [70].

Plants and soil microbes compete for Cu uptake. Cu is a vital micronutrient to maximize crop yield and quality. Too little (less than 2 mg kg^{-1}) or too much (greater than 30 mg kg^{-1}) Cu in soils will result in adverse effects on plant growth. Cu supplements can be applied either as soil amendments or fertilizers (e.g. in the form of pig slurry or $CuSO_4$) or foliar fertilizers (e.g. copper oxychloride) to the crops. Cu availability can also be controlled through changing SOM contents.

6. NITROUS OXIDE FROM OCEANS AND IN THE ATMOSPHERE

Oceans are an important source of N_2O. Freing *et al.* [71] present tracer data together with *in situ* measurements of N_2O to estimate the concentration and production rates of biologically produced N_2O in the ocean on a global scale. They estimate that oceanic N_2O production is dominated by nitrification with a contribution of only approximately 7 per cent from denitrification indicating that previously used approaches may have overestimated the contribution from denitrification. Continental shelf areas account for only a negligible fraction of the global production of N_2O, whereas coastal zones such as estuaries probably contribute significantly to the total oceanic emissions of N_2O because they are fertilized to an increasing degree by river run-off carrying a high load of organic nitrogen (eutrophication).

In the oceans, the estimated global annual subsurface N_2O production ranges from 3.1 ± 0.9 to 3.4 ± 0.9 Tg N yr^{-1}. The largest amount of subsurface N_2O is produced in the upper 500 m of the water column. The oxygen minimum zones of the intermediate layers (between 300 and 700 m water depth) in various regions of the ocean are expanding and have been losing oxygen during the last 50 years. This could result in an expansion of the zones supporting denitrification, probably having an impact on the production and decomposition of N_2O. Whether it would have a net positive or negative effect on N_2O production remains unclear as the net behaviour of denitrification and its controlling mechanisms are not yet fully understood.

There is also evidence that the oceans are warming. As marine autotrophic and heterotrophic processes display sensitivities to temperature (to varying degrees), ocean warming might result in changes of the bacterial community structure and hence in changes of N_2O production. Changes in ocean temperature also affect the solubility of N_2O. Rising ocean temperature is likely to result in the N_2O long-term storage capacity of the deep ocean being reduced. Oceanic N_2O sources are thus likely to vary as ongoing changes of the ocean environment such as deoxygenation, warming and eutrophication occur.

N_2O concentrations in the atmosphere are rising steadily with consequences not only for global warming but also for ozone destruction. The paper by Portmann *et al.* [72] reports the effects of N_2O, together with other gases CO_2, CH_4 and halocarbons on stratospheric ozone levels over the last 100 years and predicts its future evolution using a chemical model of the stratosphere. This model and the underlying chemistry are set out in their paper. It is concluded that, as halocarbons return toward pre-industrial levels, N_2O and CO_2 are likely to play the dominant roles in ozone depletion. They show, however, that there are nonlinear interactions between these gases that preclude the unambiguous separation of their effects on ozone. For example, the chemical destruction of O_3 by N_2O is buffered by the thermal effects of CO_2 in the middle stratosphere by approximately 20 per cent. Nonetheless, it is clear that N_2O is expected to be the largest ozone-destroying compound in the foreseeable future. Hence, successful mitigation of release of anthropogenic N_2O provides an important opportunity for reduction in future ozone depletion than any of the remaining uncontrolled halocarbon emissions.

7. NITROUS OXIDE EMISSIONS FROM WASTEWATER TREATMENT

An excellent example of the type of local analyses that can be applied to a single source of N_2O emission is provided by the paper from Law *et al.* [73] on wastewater treatment plants. Despite its relatively small contribution to the overall global GHG emissions, N_2O emissions from biological nutrient removal wastewater treatment plants can be very significant in terms of the contributions to their overall carbon footprint. N_2O emissions vary substantially depending on the design and operation of the plants, and on the flow and characteristics of wastewater. Such variations indicate that N_2O may be mitigated through engineering proper process design and operation. Preliminary strategies remain to be verified through full-scale applications. Law *et al.* note that in most wastewater treatment plants in contrast, for example, to soils where denitrification is often the primary source of N_2O, autotrophic NH_3 oxidation makes a relatively greater contribution than heterotrophic denitrification.

8. STRATEGIES FOR MITIGATING NITROUS OXIDE EMISSIONS

Evidence presented in this volume and elsewhere makes clear the damaging effects on climate of atmospheric N_2O. Therefore, strategies to ameliorate N_2O emission arising from intensive agricultural practices should be developed in order to decrease current levels of N_2O emissions and to forestall further rises predicted to occur as usage of nitrogenous fertilizer increases across the globe. Strategies that might be adopted arise from three quite different approaches: first, by managing soil chemistry and microbiology to ensure that bacterial denitrification runs to completion, generating N_2 instead of N_2O; second, by reducing dependence on fertilizers through engineering crop plants for example to fix nitrogen

themselves in order to sustain growth and yield, or by capitalizing on C–N interactions in the rhizosphere; third, by promoting sustainable agricultural intensification that is, producing more output from the same area of land while reducing the negative environmental impacts. We consider each of these strategies in turn.

(a) *Managing soil chemistry and microbiology*

It seems unlikely that it will ever be possible to develop farming practices that completely eliminate N_2O emissions from soil denitrifiers in agriculture. The ability to denitrify is phylogenetically diverse, and recent developments in techniques for quantifying N_2O production from denitrification show its occurrence to be more widespread than previously thought. However, it should be possible to mitigate N_2O emissions by using our understanding of the enzymology and microbiology of denitrification to design protocols to manipulate soil chemistry and physics and, thereby, the physiology of denitrifying bacteria to ensure that the reduction of N_2O to N_2 is, as far as possible, unconstrained.

Much evidence has been presented in the papers in this volume, and elsewhere, that it is the failure of the enzyme N_2OR to operate that curtails the denitrification process at N_2O rather running on to N_2. Two key factors that can cause this are low soil concentrations of Cu available to the bacterium and soil pH values below 7. Cu availability will depend not only on the absolute Cu concentration in the soil but also on the presence of competing chemical chelators, such as humic acids. Hence, there is the possibility of using SOM management, copper application or liming as primary controls of copper availability and pH values. Recent work investigated the effect of O_2 on NO_2^--dependent denitrification and the emission of NO, N_2O and N_2 in cultures of soil extracted bacteria [74]. There was evidence that N_2OR can be temporarily inactivated by sudden exposure to even low levels of O_2, whereas the other enzymes of denitrification continue to function. In soils, themselves, $N_2O–N_2$ ratios are higher as the soil pore O_2 concentration increases. This may, in part reflect a greater contribution of ammonia-oxidizing bacteria to N_2O emission, but could also arise from the sensitivity of N_2OR to O_2. It will be difficult in soils to show *in vivo* enzyme inactivation.

A full list of factors known to influence the ratio of N_2 to N_2O during denitrification include $[NO_3]^-$ and C availability, partial pressure of O_2, water-holding capacity, Cu availability as well as soil pH. The set of management options by which soil conditions might be manipulated either to lower emission of N_2O, or to increase its reduction to N_2 would include liming, manure addition, biochar or zeolite addition, minimal tillage, integrated fertilizer residue management, crop residue addition, as well as controlled release fertilizer, nitrification inhibitors, plant trait, plant breeding. Results reported by Bakken *et al.* [49] do indeed suggest that mitigation of N_2O emissions by increasing the pH of soils is currently a most promising management option. The pervasive effect of pH on the product stoichiometry of denitrification lies within the pH range 5–7, that of most agricultural soils. A recent paper points out the importance of assessing emissions according to the unit of product [75]. It shows very clearly the rapid increase in N_2O emissions when N fertilizer is added in excess of crop requirements. By considering agronomic conditions optimizing rather than minimizing nitrogen fertilizer application rates, N_2O emissions are reduced. A fuller discussion of all these aspects is given by Richardson *et al.* [18] which also contains descriptions of various management practices.

It may also be possible through plant breeding to manipulate denitrification through inputs into the plant rhizosphere, thereby changing the composition of plant-derived carbon flow or nitrogen uptake demand, or through crop spacing, tillage or integrated inorganic fertilizer, residue and SOM management. Breeding for plant release of biological nitrification inhibitors that block the AMO and hydroxylamine oxidoreductase pathways in ammonia-oxidizing bacteria promises to allow manipulation of soil nitrogen concentrations, and hence the soil denitrification potential. However, the effects on N_2O production are unknown. Such opportunities for managing N_2O emissions need to be considered in the light of effects on soil carbon levels and chemistry, not only because of the other key GHGs, CO_2 and CH_4, but also because of the important balance between fertilizer application increasing carbon sequestration through greater biomass production versus the undesirable alternative consequence of increased N_2O emission.

A key step in the future will be whether we can use technical advances in geochemistry and environmental biochemistry to monitor a wide set of parameters, both of the soil and the bacterial processes, in field studies so that we can take an ecosystems biology approach to allow identification, and ranking, of the various factors that regulate N_2O production and consumption. We note the recent development of field-deployable instruments capable of measuring nitrous oxide isotopic ratios, based on the principle of laser cavity ring down spectroscopy, CRDS [76]. It can measure continuously in real time the abundance of isotopically labelled $^{14}N^{15}N^{16}O$ and $^{15}N^{14}N^{16}O$ relative to $^{14}N^{14}N^{16}O$ in N_2O. Unlike mass spectrometry, this technique can distinguish between the two isotopomers $^{14}N^{15}N^{16}O$ and $^{15}N^{14}N^{16}O$. The nitrogen isotopic site preference, the difference between the isotope ratios of the central and terminal nitrogen atom, can distinguish between N_2O produced via the hydroxylamine oxidation pathway and that of nitrate reduction as well as between fungal and bacterial N_2O production.

Central to the development of appropriate mitigation practices is addressing the challenge of spatial scale. N_2O production impacts us at different spatial scales, from the cellular production to the landscape, and to the global impact of climate change and feedbacks within and between these scales. The challenge we face is in understanding phenomena of global magnitude that have their foundations at the microscale, and to formulate appropriate management practices for mitigation that are informed by regulation at the microscale. Recent efforts have demonstrated links

between microbial gene expression, environmental parameters and N_2O-genic processes at the microcosm scale, but there is still much progress to be made when relating this to processes at the macroscale, exacerbated by the high-spatial heterogeneity of N_2O emission [77]. The scaling up, or even scaling down, of N_2O producing processes in the plant–soil–microbe system is essential to inform policymakers of the environmental factors driving climate change that can be targeted for management, and may help reduce model uncertainty, which is vital for accurate prediction of emissions and for the formulation of appropriate mitigation strategies. We have invested much effort into examining the drivers of microbial activity at the rhizosphere to plot scales, but there is still uncertainty over whether this regulation is still relevant at the landscape scale, how we can extrapolate between scales, and whether the drivers of N_2O production/reduction that can be targeted for management vary depending on the spatial scale being considered. To address this will require integration of molecular, microbiology, physiology, physics, biogeochemistry and mathematical modelling approaches.

(b) *Engineering crop plants*

A recent review discusses the feasibility of, and assesses the way forward in, reducing dependence on fertilizers through engineering crop plants to fix nitrogen themselves in order to sustain growth and yield [78]. This paper drew on a meeting convened by the Bill and Melinda Gates Foundation. Three approaches were considered. The first is the development of root nodule symbioses in cereals. Legumes and actinorhizal (non-legume) plants have evolved productive nitrogen-fixing symbioses with rhizobial and *Frankia* bacteria, respectively. The main steps required to make symbiotic nitrogen-fixing cereals include engineering bacteria to recognize and infect a host cereal root cell, and having the plant subsequently establish a low-oxygen environment such as a root nodule. The second approach discussed was the application, as fertilizers, of nitrogen-fixing endophytic bacteria that form nodule-independent associations with cereal crops. Although commercial biofertilizers containing such bacteria are available, it is unclear whether the enhancement of plant growth is the result of nitrogen fixation or of bacterial molecules that act as plant growth hormones. Nevertheless, biofertilizers represent an existing, and the only currently available, technology. The third method considered was the introduction of the nitrogenase enzyme system into a plant organelle. To achieve this, the complete biosynthetic pathway of the several components of the nitrogenase enzyme must be engineered into cereals and targeted to a low-oxygen compartment within the plant. In a related approach, a recent paper has reported expression of the Nos operon proteins from *Pseudomonas stutzeri* in transgenic plants to assemble N_2OR, the objective being to bestow on plants the ability to reduce N_2O to N_2 themselves. Both the single-gene transformants (*nosZ*) and the multi-gene transformants (*nosFLZDY*) produced active recombinant N_2OR. Enzymatic activity was detected using the methyl viologen-linked enzyme assay, showing that extracts from both types of transgenic plants exhibited N_2O-reducing activity [79].

All these approaches are challenging but the rewards would be great. It has been claimed that, if the coupling of nitrogen supply and carbon metabolism could be achieved, excess nitrogen would not be lost to the environment, thereby resulting in lower N_2O emissions.

(c) *Sustainable agricultural intensification*

Agriculture contributes a disproportionate amount of GHGs with high impact on warming notably about 47 per cent and 58 per cent of total CH_4 and N_2O emissions, respectively. Of all global land area, 14 per cent is used for food production, which ties up a vast amount of carbon. Changes in agricultural practices that affect this store could have a considerable effect on global warming.

Sustainable agricultural intensification is defined as producing more output from the same area of land while reducing the negative environmental impacts and at the same time increasing contributions to natural capital and the flow of environmental services [80,81]. A sustainable production system would thus exhibit most of the following attributes:

- using crop varieties and livestock breeds with a high ratio of productivity to use of externally derived inputs;
- avoiding the unnecessary use of external inputs;
- harnessing agro-ecological processes such as nutrient cycling, biological nitrogen fixation, allelopathy, predation and parasitism;
- minimizing use of technologies or practices that have adverse impacts on the environment and human health;
- making productive use of human capital in the form of knowledge and capacity to adapt and innovate and social capital to resolve common landscape-scale problems; and
- quantifying and minimizing the impacts of system management on externalities such as GHG emissions, clean water availability, carbon sequestration, conservation of biodiversity, and dispersal of pests, pathogens and weeds.

In terms of technologies, therefore, productive and sustainable agricultural systems make the best of both crop varieties and livestock breeds and their agro-ecological and agronomic management. The pioneering rice breeder, Peter Jennings, who led early advancements in high-yielding rice varieties during the first green revolution, has argued for an 'agronomic revolution': 'It is now widely recognized that rice yield gaps result from agronomic failings, and that future yield increases depend heavily on this science. Agronomy's time has come to lift farm productivity out of stagnancy' quotation from Pretty [81]. Agronomy refers to the management of crops and livestock in their specific circumstances, and matches with the emergence of the term agro-ecology to indicate that there is a need to invest in science and practice

that gives farmers a combination of the best possible seeds and breeds and their management in local ecological contexts.

This suggests that sustainable intensification will very often involve more complex mixes of domesticated plant and animal species and associated management techniques, requiring greater skills and knowledge by farmers. To increase production efficiently and sustainably, farmers need to understand under what conditions agricultural inputs (seeds, fertilizers and pesticides) can either complement or contradict biological processes and ecosystem services that inherently support agriculture. In all cases, farmers need to see for themselves that added complexity and increased efforts can result in substantial net benefits to productivity, but they need also to be assured that increasing production actually leads to increases in income. Too many successful efforts in raising production yields have ended in failure when farmers were unable to market the increased outputs. Understanding how to access rural credit, or how to develop warehouse receipt systems and especially, how to sell any increased output, becomes as important as learning how to maximize input efficiencies or build fertile soils.

9. CONCLUSIONS

Despite decades of research on N_2O emissions, few mitigation options have been proposed or even fewer trialled. A key target should be to improve the product stoichiometry of denitrification (N_2/N_2O) in agro-ecosystems. The understanding now reached of the genetics, microbiology, enzymology and chemistry allows trials in the field to be designed. The availability of mobile monitoring systems, such as MS and CRDS, together with isotopic spiking, and coupling to molecular ecology approaches provides the means to diagnose, distinguish and quantify the pathways operating and, hence, to allow a description of the fate of applied N to be reached. This should enable the exploration of different management options to ascertain their effectiveness. Systematic studies of complex interactions in such eco-systems that are contributing globally to the release of the potent GHG N_2O are now feasible. They should be providing prescriptions for the minimization of N_2O emissions from soils under a wide variety of circumstances.

We thank the following colleagues L. Bakken, H. W. Bange, K. Goulding, P. Hirsch, N. Le Brun, I. Moura, R. Portmann, U. Skiba, K. Smith and S. Spiro for helpful discussions.

REFERENCES

1 Lassey, K. & Harvey, M. 2007 Nitrous oxide: the serious side of laughing gas. *Water Atmos.* **15**, 1–10.
2 Zumft, W. G. 1997 Cell biology and molecular basis of denitrification. *Microbiol. Mol. Biol. Rev.* **61**, 533–616.
3 NERC. 2005 Global nitrogen enrichment (GANE): reports and key findings. See www.nerc.ac.uk/research/programmes/gane/results.asp (accessed 15 December 2011)
4 Kimochi, Y., Inamori, Y., Mizuochi, M., Xu, K.-Q. & Matsumura, M. 1998 Nitrogen removal and N_2O emission in a full-scale domestic wastewater treatment plant with intermittent aeration. *J. Biosci. Bioeng.* **86**, 202–206. (doi:10.1016/s0922-338x(98)80114-1)
5 Davidson, E. A. 2009 The contribution of manure and fertilizer nitrogen to atmospheric nitrous oxide since 1860. *Nat. Geosci.* **2**, 659–662. (doi:10.1038/ngeo608)
6 Bates, B., Kundzewicz, Z., Wu, S. & Palutikof, J. 2008 *Climate change and water. Technical Paper of the Intergovernmental Panel on Climate Change*. Geneva, Switzerland: IPCC.
7 IPCC. 2007 Working group I: the physical science basis. In *IPCC fourth assessment report: climate change 2007* (eds S. Solomon, D. Qin, M. Manning, Z. Chen, M. Marquis, K. B. Averyt, M. Tignor, H. L. Miler). Cambridge, UK: Cambridge University Press.
8 Crutzen, P. & Oppenheimer, M. 2008 Learning about ozone depletion. *Clim. Change* **89**, 143–154. (doi:10.1007/s10584-008-9400-6)
9 Ravishankara, A. R., Daniel, J. S. & Portmann, R. W. 2009 Nitrous oxide (N_2O): the dominant ozone-depleting substance emitted in the 21st century. *Science* **326**, 123–125. (doi:10.1126/science.1176985)
10 IPCC. 2001 Atmospheric chemistry and greenhouse gases. In *Climate change 2001: the scientific basis. contribution of working group I to the third assessment report of the intergovernmental panel on climate change* (eds J. Houghton, Y. Ding, D. Griggs, M. Noguer, P. van der Linden, X. Dai, K. Maskell, C. A. Johnson). Cambridge, UK: Cambridge University Press.
11 2009 *Reaping the benefits: science and the sustainable intensification of global agriculture*. London, UK: The Royal Society.
12 Bothe, H., Ferguson, S. J. & Newton, W. E. 2006 *Biology of the nitrogen cycle*. Amsterdam, The Netherlands: Elsevier.
13 Day, D. A., Poole, P. S., Tyerman, S. D. & Rosendahl, L. 2001 Ammonia and amino acid transport across symbiotic membranes in nitrogen-fixing legume nodules. *Cell. Mol. Life Sci.* **58**, 61–71. (doi:10.1007/pl00000778)
14 Bertero, M. G., Rothery, R. A., Palak, M., Hou, C., Lim, D., Blasco, F., Weiner, J. H. & Strynadka, N. C. 2003 Insights into the respiratory electron transfer pathway from the structure of nitrate reductase A. *Nat. Struct. Biol.* **10**, 681–687. (doi:10.1038/nsb969)
15 Adman, E. T. 1995 A taste of copper. *Nat. Struct. Biol.* **2**, 929–931. (doi:10.1038/nsb1195-929)
16 Hino, T., Matsumoto, Y., Nagano, S., Sugimoto, H., Fukumori, Y., Murata, T., Iwata, S. & Shiro, Y. 2010 Structural basis of biological N_2O generation by bacterial nitric oxide reductase. *Science* **330**, 1666–1670. (doi:10.1126/science.1195591)
17 Watmough, N. J., Field, S. J., Hughes, R. J. L. & Richardson, D. J. 2009 The bacterial respiratory nitric oxide reductase. *Biochem. Soc. Trans.* **37**, 200–300. (doi:10.1042/BST0370200)
18 Richardson, D., Felgate, H., Watmough, N., Thomson, A. & Baggs, E. 2009 Mitigating release of the potent greenhouse gas N_2O from the nitrogen cycle: could enzymic regulation hold the key? *Trends Biotechnol.* **27**, 388–397. (doi:10.1016/j.tibtech.2009.03.009)
19 Scala, D. J. & Kerkhof, L. J. 1998 Nitrous oxide reductase (*nosZ*) gene-specific PCR primers for detection of denitrifiers and three *nosZ* genes from marine sediments. *FEMS Microbiol. Lett.* **162**, 61–68. (doi:10.1016/s0378-1097(98)00103-7)
20 Brown, K., Djinovic-Carugo, K., Haltia, T., Cabrito, I., Saraste, M., Moura, J. G., Moura, I., Tegoni, M. & Cambillau, C. 2000 Revisiting the catalytic CuZ cluster of nitrous oxide (N_2O) reductase. *J. Biol. Chem.* **275**, 41 133–41 136. (doi:10.1074/jbc.M008617200)

21 Rotthauwe, J. H., Witzel, K. P. & Liesack, W. 1997 The ammonia monooxygenase structural gene amoa as a functional marker: molecular fine-scale analysis of natural ammonia-oxidizing populations. *Appl. Environ. Microbiol.* **63**, 4704–4712.

22 Arp, D. J., Sayavedra-Soto, L. A. & Hommes, N. G. 2002 Molecular biology and biochemistry of ammonia oxidation by *Nitrosomonas europaea*. *Arch. Microbiol.* **178**, 250–255. (doi:10.1007/s00203-002-0452-0)

23 Allen, A. E., Booth, M. G., Frischer, M. E., Verity, P. G., Zehr, J. P. & Zani, S. 2001 Diversity and detection of nitrate assimilation genes in marine bacteria. *Appl. Environ. Microbiol.* **67**, 5343–5348. (doi:10.1128/AEM.67.11.5343-5348.2001)

24 Ohashi, Y., Shi, W., Takatani, N., Aichi, M., Maeda, S. I., Watanabe, S., Yoshikawa, H. & Omata, T. 2011 Regulation of nitrate assimilation in cyanobacteria. *J. Exp. Botany* **62**, 1411–1424. (doi:10.1093/jxb/erq427)

25 Nolden, L., Farwick, M., Krämer, R. & Burkovski, A. 2001 Glutamine synthetases of *Corynebacterium glutamicum*: transcriptional control and regulation of activity. *FEMS Microbiol. Lett.* **201**, 91–98. (doi:10.1016/s0378-1097(01)00244-0)

26 Ciurli, S., Marzadori, C., Benini, S., Deiana, S. & Gessa, C. 1996 Urease from the soil bacterium *Bacillus pasteurii*: immobilization on Ca-polygalacturonate. *Soil Biol. Biochem.* **28**, 811–817. (doi:10.1016/0038-0717(96)00020-x)

27 Skiba, U. & Smith, K. A. 2000 The control of nitrous oxide emissions from agricultural and natural soils. *Chemosphere Glob. Change Sci.* **2**, 379–386. (doi:10.1016/S1465-9972(00)00016-7)

28 Smith, P. *et al.* 2008 Greenhouse gas mitigation in agriculture. *Phil. Trans. R. Soc. B* **363**, 789–813. (doi:10.1098/rstb.2007.2184)

29 Okereke, G. U. 1993 Growth yield of denitrifiers using nitrous oxide as a terminal electron acceptor. *World J. Microbiol. Biotechnol.* **9**, 59–62. (doi:10.1007/bf00656518)

30 Bange, H. W., Freing, A., Kock, A. & Löscher, C. R. 2010 Marine pathways to nitrous oxide (N_2O). In *Nitrous oxide and climate change* (ed. K. Smith), pp. 36–62. London, UK: Earthscan.

31 Brussaard, L. *et al.* 1997 Biodiversity and ecosystem functioning in soil. *Ambio* **26**, 563–70.

32 Smith, K., Mosier, A., Crutzen, P. & Winiwarter, W. 2011 The role of N_2O derived from biofuels (and from agriculture in general) in Earth's climate. *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0313)

33 Skiba, U. *et al.* 2011 UK emissions of the greenhouse gas nitrous oxide. *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0356)

34 Santoro, A. E., Buchwald, C., McIlvin, M. R. & Casciotti, K. L. 2011 Isotopic signature of N_2O produced by marine ammonia-oxidizing archaea. *Science* **333**, 1282–1285. (doi:10.1126/science.1208239)

35 Shoun, H. & Tanimoto, T. 1991 Denitrification by the fungus *Fusarium oxysporum* and involvement of cytochrome P450 in the respiratory nitrite reduction. *J. Biol. Chem.* **266**, 11 078–11 082.

36 Shoun, H., Fushinobu, S., Jiang, L., Kim, S.-W. & Wakagi, T. 2011 Fungal denitrification and nitric oxide reductase cytochrome P450nor. *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0335)

37 Prendergast-Miller, M. T., Baggs, E. M. & Johnson, D. 2011 Nitrous oxide production by the ectomycorrhizal fungi *Paxillus involutus* and *Tylospora fibrillosa*. *FEMS Microbiol. Lett.* **316**, 31–35. (doi:10.1111/j.1574-6968.2010.02187.x)

38 Sutka, R. L., Adams, G. C., Ostrom, N. E. & Ostrom, P. H. 2008 Isotopologue fractionation during N_2O production by fungal denitrification. *Rapid Commun. Mass Spectrom.* **22**, 3989–3996. (doi:10.1002/rcm.3820)

39 Shiro, Y., Sugimoto, H., Tosha, T., Nagano, S. & Hino, T. 2011 Structural basis for nitrous oxide generation by bacterial nitric oxide reductases. *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0310.R2)

40 Ananyev, G. M., Zaltsman, L., Vasko, C. & Dismukes, G. C. 2001 The inorganic biochemistry of photosynthetic oxygen evolution/water oxidation. *Biochim. Biophys. Acta Bioenerg.* **1503**, 52–68. (doi:10.1016/s0005-2728(00)00215-2)

41 Stremińska, M. A., Felgate, H., Rowley, G., Richardson, D. J. & Baggs, E. M. 2011 Nitrous oxide production in soil isolates of nitrate-ammonifying bacteria. *Environ. Microbiol. Rep.* (doi:10.1111/j.1758-2229.2011.00302.x) **Q4**

42 Spiro, S. 2011 Nitrous oxide production and consumption: regulation of gene expression by gas-sensitive transcription factors. *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0309)

43 Brown, K., Tegoni, M., Prudencio, M., Pereira, A. S., Besson, S., Moura, J. J., Moura, I. & Cambillau, C. 2000 A novel type of catalytic copper cluster in nitrous oxide reductase. *Nat. Struct. Mol. Biol.* **7**, 191–195. (doi:10.1038/73288)

44 Simon, J., Einsle, O., Kroneck, P. M. H. & Zumft, W. G. 2004 The unprecedented nos gene cluster of *Wolinella succinogenes* encodes a novel respiratory electron transfer pathway to cytochrome c nitrous oxide reductase. *FEBS Lett.* **569**, 7–12. (doi:10.1016/j.febslet.2004.05.060)

45 Zumft, W. G. & Matsubara, T. 1982 A novel kind of multi-copper protein as terminal oxidoreductase of nitrous oxide respiration in *Pseudomonas perfectomarinus*. *FEBS Lett.* **148**, 107–112. (doi:10.1016/0014-5793(82)81253-2)

46 Dell'Acqua, S., Pauleta, S., Moura, J. & Moura, I. 2011 Biochemical characterization of the purple form of *Marinobacter hydrocarbonoclasticus* nitrous oxide reductase. *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0311.R1)

47 Pomowski, A., Zumft, W. G., Kroneck, P. M. H. & Einsle, O. 2011 N_2O binding at a [4Cu:2S] copper-sulphur cluster in nitrous oxide reductase. *Nature* **477**, 234–237. (doi:10.1038/nature10332)

48 Honisch, U. & Zumft, W. G. 2003 Operon structure and regulation of the nos gene region of *Pseudomonas stutzeri*, encoding an ABC-Type ATPase for maturation of nitrous oxide reductase. *J. Bacteriol.* **185**, 1895–1902. (doi:10.1128/jb.185.6.1895-1902.2003)

49 Bakken, L., Bergaus, L., Liu, B. & Frostegård, Å. 2011 Regulation of denitrification at the cellular level: a clue to the understanding of N_2O emissions from soils. *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0321)

50 Verbaendert, I., Boon, N., De Vos, P. & Heylen, K. 2011 Denitrification is a common feature among members of the genus Bacillus. *Syst. Appl. Microbiol.* **34**, 385–391. (doi:10.1016/j.syapm.2011.02.003)

51 Jones, C. M., Welsh, A., Throbäck, I. N., Dörsch, P., Bakken, L. R. & Hallin, S. 2011 Phenotypic and genotypic heterogeneity among closely related soil-borne N_2- and N_2O-producing Bacillus isolates harboring the nosZ gene. *FEMS Microbiol. Ecol.* **76**, 541–552. (doi:10.1111/j.1574-6941.2011.01071.x)

52 Clark, I., Buchkina, N., Jhurreea, D., Goulding, K. & Hirsch, P. 2011 Impacts of nitrogen application rates on the activity and diversity of denitrifying bacteria in the Broadbalk wheat experiment. *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0314)

53 Kučera, I., Matyášek, R. & Dadák, V. 1986 The influence of pH on the kinetics of dissimilatory nitrite

rstb20110415—31/1/12—17:03–Copy Edited by: J. Tamilselvam

reduction in *Paracoccus denitrificans*. *Biochim. Biophys. Acta Bioenerg.* **848**, 1–7. (doi:10.1016/0005-2728(86)90153-2)

54 Baumann, B., van der Meer, J. R., Snozzi, M. & Zehnder, A. J. B. 1997 Inhibition of denitrification activity but not of mRNA induction in *Paracoccus denitrificans* by nitrite at a suboptimal pH. *Antonie van Leeuwenhoek* **72**, 183–189. (doi:10.1023/a:1000342125891)

55 Bergaust, L., Mao, Y., Bakken, L. R. & Frostegård, Å. 2010 Denitrification response patterns during the transition to anoxic respiration and post-transcriptional effects of suboptimal pH on nitrogen oxide reductase in *Paracoccus denitrificans*. *Appl. Environ. Microbiol.* **76**, 6387–6396. (doi:10.1128/aem.00608-10)

56 ŠImek, M. & Cooper, J. E. 2002 The influence of soil pH on denitrification: progress towards the understanding of this interaction over the last 50 years. *Eur. J. Soil Sci.* **53**, 345–354. (doi:10.1046/j.1365-2389.2002.00461.x)

57 Šimek, M., Jíšová, L. & Hopkins, D. W. 2002 What is the so-called optimum pH for denitrification in soil? *Soil. Biol. Biochem.* **34**, 1227–1234. (doi:10.1016/s0038-0717(02)00059-7)

58 Thomsen, J. K., Geest, T. & Cox, R. P. 1994 Mass spectrometric studies of the effect of pH on the accumulation of intermediates in denitrification by *Paracoccus denitrificans*. *Appl. Environ. Microbiol.* **60**, 536–541.

59 Guo, J. H. *et al.* 2010 Significant acidification in major Chinese croplands. *Science* **327**, 1008–1010. (doi:10.1126/science.1182570)

60 Philippot, L., Hallin, S., Börjesson, G. & Baggs, E. M. 2009 Biochemical cycling in the rhizosphere having an impact on global change. *Plant Soil* **321**, 61–81. (doi:10.1007/s11104-008-9796-9)

61 Morley, N. & Baggs, E. M. 2010 Carbon and oxygen controls on N_2O and N_2 production during nitrate reduction. *Soil Biol. Biochem.* **42**, 1864–1871. (doi:10.1016/j.soilbio.2010.07.008)

62 Kim, H. J., Graham, D. W., DiSpirito, A. A., Alterman, M. A., Galeva, N., Larive, C. K., Asunskis, D. & Sherwood, P. M. A. 2004 Methanobactin, a copper-acquisition compound from methane-oxidizing bacteria. *Science* **305**, 1612–1615. (doi:10.1126/science.1098322)

63 Andreazza, R., Okeke, B., Pieniz, S., Brandelli, A., Lambais, M. & Camargo, F. 2010 Bioreduction of Cu(II) by cell-free copper reductase from a copper resistant *Pseudomonas sp.* NA. *Biol. Trace Elem. Res.* **143**, 1–11. (doi:10.1007/s12011-010-8899-3)

64 Alloway, B. J. (ed.) 2008 *Micronutrient deficiencies in global crop production*. Heidelberg, Germany: Springer.

65 Ore, S., Mertens, J., Brandt, K. K. & Smolders, E. 2010 Copper toxicity to bioluminescent *Nitrosomonas europaea* in soil is explained by the free metal ion activity in pore water. *Environ. Sci. Technol.* **44**, 9201–9206. (doi:10.1021/es1026294)

66 Anttila, J. *et al.* 2011 Is coproporphyrin III a copper-acquisition compound in *Paracoccus denitrificans*? *Biochim. Biophys. Acta Bioenerg.* **1807**, 311–318. (doi:10.1016/j.bbabio.2010.12.014)

67 Enwall, K., Throback, I. N., Stenberg, M., Soderstrom, M. & Hallin, S. 2010 Soil resources influence spatial patterns of denitrifying communities at scales compatible with land management. *Appl. Environ. Microbiol.* **76**, 2243–2250. (doi:10.1128/aem.02197-09)

68 Bru, D., Ramette, A., Saby, N. P. A., Dequiedt, S., Ranjard, L., Jolivet, C., Arrouays, D. & Philippot, L. 2011 Determinants of the distribution of nitrogen-cycling microbial communities at the landscape scale. *ISME J.* **5**, 532–542. (doi:10.1038/ismej.2010.130)

69 Kandeler, F., Kampichler, C. & Horak, O. 1996 Influence of heavy metals on the functional diversity of soil microbial communities. *Biol. Fert. Soils* **23**, 299–306. (doi;10.1007/bf00335958)

70 Ruyters, S., Mertens, J., T'Seyen, I., Springael, D. & Smolders, E. 2010 Dynamics of the nitrous oxide reducing community during adaptation to Zn stress in soil. *Soil Biol. Biochem.* **42**, 1581–1587. (doi:10.1016/j.soilbio.2010.05.036)

71 Freing, A., Wallace, D. & Bange, H. 2011 Global oceanic production of nitrous oxide (N_2O). *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0360)

72 Portmann, R., Daniel, J. & Ravishankara, A. 2011 Ozone depletion due to N_2O: influences of other gases. *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0377)

73 Law, Y., Ye, L., Pan, Y. & Yuan, Z. 2011 Nitrous oxide emissions from wastewater treatment processes. *Phil. Trans. R. Soc. B* **367**. (doi:10.1098/rstb.211.0317)

74 Morley, N., Baggs, E. M., Dörsch, P. & Bakken, L. 2008 Production of NO, N_2O and N_2 by extracted soil bacteria, regulation by NO_2^- and O_2 concentrations. *FEMS Microbiol. Ecol.* **65**, 102–112. (doi:10.1111/j.1574-6941.2008.00495.x)

75 van Groenigen, J. W., Velthof, G. L., Oenema, O., van Groenigen, K. J. & van Kessel, C. 2010 Towards an agronomic assessment of N_2O emissions: a case study for arable crops. *Eur. J. Soil Sci.* **61**, 903–913. (doi:10.1111/j.1365-2389.2009.01217.x)

76 Crosson, E., Balslev-Clausen, D. & Dore, J. (eds). 2011 A new analyzer to measure the abundance of $^{14}N^{15}N^{16}O$ and $^{15}N^{14}N^{16}O$ relative to $^{14}N^{14}N^{16}O$ to help elucidate microbial N_2O dynamics in terrestrial ecosystems. In *EGU General Assembly, 3–8 April 2011*. Vienna, Austria: European Geosciences Union.

77 Standing, D., Baggs, E. M., Wattenbach, M., Smith, P. & Killham, K. 2007 Meeting the challenge of scaling up processes in the plant–soil–microbe system. *Biol. Fert. Soils* **44**, 245–257. (doi:10.1007/s00374-007-0249-z)

78 Beatty, P. H. & Good, A. G. 2011 Future prospects for cereals that fix nitrogen. *Science* **333**, 416–417. (doi:10.1126/science.1209467)

79 Wan, S., Mottiar, Y., Johnson, A. M., Goto, K. & Altosaar, I. 2011 Expression of the Nos operon proteins from *Pseudomonas stutzeri* in transgenic plants to assemble nitrous oxide reductase. *Transgenic Res.* 1–11. (doi:10.1007/s11248-011-9555-1) Q5

80 Pretty, J., Toulmin, C. & Williams, S. 2011 Sustainable intensification in African agriculture. *Int. J. Agric. Sustainability* **9**, 5–24. (doi:10.3763/ijas.2010.0583)

81 Pretty, J. 2008 Agricultural sustainability: concepts, principles and evidence. *Phil. Trans. R. Soc. B* **363**, 447–65. (doi:10.1098/rstb.2007.2163)

rstb20110415—31/1/12—17:03–Copy Edited by: J. Tamilselvam