# Drivers of Southern Ocean food web structure and impacts of environmental change

Patrick Eskuche-Keith



Thesis submitted in fulfilment of the requirements of the University of Essex for the degree of Doctor of Philosophy

> School of Life Sciences University of Essex July 2024



"It seems to me that the natural world is the greatest source of excitement; the greatest source of visual beauty; the greatest source of intellectual interest. It is the greatest source of so much in life that makes life worth living".

Sir David Attenborough

# 1 Abstract

2 The Southern Ocean is experiencing major environmental and ecological changes which 3 could drastically alter communities and impact ecosystem functioning. We still have a poor 4 understanding of the structure of Southern Ocean food webs and their likely responses to 5 ongoing and future changes, which limits our ability to develop and implement effective 6 management and conservation strategies. This thesis employs multiple approaches to 7 investigate several aspects relating to the structure and dynamics of Southern Ocean food 8 webs. First, the links between morphological traits and trophic niches are explored within the 9 demersal fish community of the subantarctic island of South Georgia, highlighting the role of 10 functional traits in driving community structure. Second, functional traits including body size, 11 mobility, foraging habitat and feeding mode are used to identify the drivers of stabilising sub-12 structures (modularity) across regional food webs. This leads to the hypothesis that habitat 13 heterogeneity is a major determinant of the distribution of modules within networks. Third, 14 an extensive dataset of mesopelagic fish and zooplankton samples from across a latitudinal 15 temperature gradient is used to determine the impact of warming on predator-prey body mass 16 ratios (PPMR). This reveals that ongoing environmental change may reorganise the size-17 structure of Southern Ocean ecosystems, with implications for their stability. Fourth, the 18 possible consequences of ongoing baleen whale population recovery for competitor 19 biomasses are explored using the Ecopath framework, with the conclusion that strong trade-20 offs between conservation objectives are likely unless substantial increases in suitable 21 primary production occur. This thesis finishes with a synthesis of these new insights into the 22 structure and dynamics of Southern Ocean food webs and discusses the major future 23 directions for food web research more generally.

# 24 Contributions statement

I hereby declare that the chapters and associated papers that make up this thesis are the result of my own work. My supervisory team (Drs Eoin O'Gorman, Simeon Hill, Michelle Taylor and Phil Hollyman) provided conceptual guidance and editorial feedback on drafts. Various external collaborators (Drs Lucía López-López, Ryan Saunders, Geraint Tarling, Benjamin Rosenbaum and Martin Collins) also provided invaluable help with various aspects of this thesis, including contributing data, aiding in fieldwork and providing feedback on manuscript drafts.

32

## 33 Acknowledgements

34 Writing this thesis broadened my appreciation for the interconnectedness of the natural world

35 and has strengthened my passion for understanding how the interactions between organisms

36 mediate the patterning of biodiversity and maintenance of ecosystem processes. Pursuing this

37 PhD has brought me into contact with amazing people and allowed me to experience mind-

38 blowing places, and for that I will be ever grateful.

This thesis has been a long journey and I've only made it through thanks to the support and encouragement of my supervisors, friends and family. First and foremost, thank you to my supervisory team, Dr Eoin O'Gorman, Dr Simeon Hill, Dr Michelle Taylor and Dr Phil Hollyman. Your feedback and guidance helped me develop my skills and independence as a researcher, and I am hugely grateful for your encouragement throughout this project. Thank you for all of the opportunities you created, from conducting fieldwork to collaborations with other researchers and publishing articles – it's been a blast!

46 Much of this thesis came about through collaborations with incredible researchers at various

47 institutes. In no particular order, thanks to Dr Ryan Saunders, Prof. Geraint Tarling and Dr

48 Martin Collins (British Antarctic Survey), Dr Benjamin Rosenbaum (iDiv), and Dr Lucía

49 López-López (IEO). Thanks also to the UK NERC and ARIES DTP for funding my project.

50 The British Antarctic Survey also provided logistical support to facilitate the fieldwork

51 underlying my first chapter, and the EU COST Action Sea-Unicorn funded the research stay

52 in Santander which ultimately led to the development of my third chapter.

53 Having a strong support network is key to success and I was lucky enough to have an 54 excellent group of friends to see me through. You are too many to thank individually so here 55 it goes in no particular order: Amy, Howard, Alex (thanks for all the shenanigans and for 56 introducing me to the bliss that is freediving), Zelin, Mike, Alice (I hope you cure your plant 57 obsession soon, for all our sakes), Peter, Lucy, Haleigh, Nonny, Jake, Hugo and Joe. Thanks 58 also to the various friends I made at BAS (Paul, maybe one day you'll have a functional air 59 mattress for me) and at KEP. In the words of a wise man, keep crushing it! Turns out that 60 light in the tunnel isn't a train after all.

61 To my parents Hugh and Lisa, you got me to this point through thick and thin and I hope I've

62 made you proud. Maybe now I'll finally get a real job. And to my wonderful partner Melissa,

63 thanks for sticking it out with me, I'm so proud of what you have achieved in the meantime.

64 Here's to the future!

# **Contents**

66	1 General Introduction9
67	1.1 Biodiversity and ecosystem stability9
68	1.2 Food web models11
69	1.3 The drivers of food web structure15
70	1.4 Food web stability16
71	1.5 The Southern Ocean18
72	1.6 Thesis outline
73	2 Morphological traits distinguish feeding guilds in a Southern Ocean fish community25
74	2.1 Introduction25
75	2.2 Materials and methods:
76	2.2.1 Sample collection:
77	2.2.2 Morphological measurements and stomach contents dissection:
78	2.2.3 Identification of feeding guilds
79	2.2.4 Predator-prey size relationships
80	2.2.5 Morphological trait distributions
81	2.3 Results
82	2.3.1 Species trait relationships
83	2.3.2 Feeding guilds
84	2.3.3 Distinguishing feeding guilds with functional traits
85	2.4 Discussion
86	2.4.1 Size-based feeding
87	2.4.2 Functional traits and feeding guilds
88	2.4.3 Further considerations
89	2.4.4 Conclusion
90	3 Trophic structuring of modularity alters energy flow through marine food webs44
91	3.1 Introduction

92	3.2 Materials and methods4	16
93	3.2.1 Study systems	6
94	3.2.2 Module identification4	17
95	3.2.3 Functional traits4	18
96	3.2.4 Statistical analysis4	19
97	3.3 Results	50
98	3.3.1 Module identification	51
99	3.3.2 Module topology	53
100	3.3.3 Functional traits	55
101	3.4 Discussion	58
102	3.4.1 Drivers of structural differences	59
103	3.4.2 Implications for food web stability6	51
104	3.4.3 Further considerations	52
105	3.4.4 Conclusion6	53
106	4 Temperature alters the predator-prey size relationships and size-selectivity of	
106 107	4 Temperature alters the predator-prey size relationships and size-selectivity of Southern Ocean fish6	64
107	Southern Ocean fish6	54
107 108	Southern Ocean fish	54 58
107 108 109	Southern Ocean fish	54 58 58
107 108 109 110	Southern Ocean fish	54 58 58
107 108 109 110 111	Southern Ocean fish.       6         4.1 Introduction.       6         4.2 Materials and methods       6         4.2.1 Fish sampling.       6         4.2.2 Zooplankton sampling       6	54 58 58 59
107 108 109 110 111 112	Southern Ocean fish	54 58 59 59 70
107 108 109 110 111 112 113	Southern Ocean fish	54 58 58 59 70 72
107 108 109 110 111 112 113 114	Southern Ocean fish.64.1 Introduction.64.2 Materials and methods64.2.1 Fish sampling.64.2.2 Zooplankton sampling64.2.3 Environmental covariates.64.2.4 Statistical analyses74.3 Results and discussion7	54 58 59 59 70 72 5 <b>f</b>
107 108 109 110 111 112 113 114 115	Southern Ocean fish	54 58 59 59 70 72 5 <b>f</b> 30
<ol> <li>107</li> <li>108</li> <li>109</li> <li>110</li> <li>111</li> <li>112</li> <li>113</li> <li>114</li> <li>115</li> <li>116</li> </ol>	Southern Ocean fish	54 58 59 59 70 72 <b>51</b> <b>30</b> 30

120	5.2.2 Regional Ecopath models
121	5.2.3 Model standardisation
122	5.2.4 Catch-derived estimates of plausible whale biomass
123	5.2.5 Estimates of baleen whale Q/B92
124	5.2.6 Model balancing
125	5.2.7 Perturbation scenarios
126	5.3 Results
127	5.3.1 Initial balanced model ensembles95
128	5.3.2 Catch-derived total whale biomass
129	5.3.3 Perturbation scenarios
130	5.4 Discussion
131	6 General discussion110
132	6.1 Chapter contributions110
133	6.1.1 Chapter 2: Morphological traits distinguish feeding guilds in a Southern Ocean fish
134	community110
135	6.1.2 Chapter 3: Trophic structuring of modularity alters energy flow through marine
136	food webs
137	6.1.3 Chapter 4: Temperature alters the predator-prey size relationships and size-
138	selectivity of Southern Ocean fish
139 140	6.1.4 Chapter 5: Trade-offs between the recovery of Southern Ocean baleen whales and conservation of their competitors
141	6.2 Future Directions
142	6.2.1 Using functional traits to explain and predict food web structure
143	6.2.2 Understanding temporal and spatial variability in food web structure
144	6.2.3 Using bioenergetics to gain mechanistic understanding of food web dynamics 122
144	6.2.4 Summary
145	References
147	Appendix A: Supplementary material for chapter 2161

148	A1: Supplementary figures	
149	A2: Supplementary tables	162
150	Appendix B: Supplementary material for chapter 3	170
151	B1: Supplementary methods and results	170
152	B2: Supplementary figures	175
153	Appendix C: Supplementary material for chapter 4	
154	C1: Supplementary figures	
155	C2: Supplementary tables	190
156	Appendix D: Supplementary material for chapter 5	214
157	D1: Supplementary methods	214
158	6.2.5 Re-aggregation of functional groups	214
159	6.2.6 Automated balancing routine:	218
160	D2: Supplementary figures	
161	D3: Supplementary tables	235
162		
163		
164		
165		
166		
167		
168		
169		
170 171		
171		
173		

# 174 **1 General Introduction**

175 Our oceans are experiencing unprecedented environmental and ecological changes. These 176 include climate-driven warming of waters (including more frequent, stronger marine heatwaves), melting of sea ice, and changes in primary productivity (Cooley et al. 2022). 177 178 Additionally, the growing human population is resulting in ever-increasing demand for food, 179 driving the over-exploitation of marine resources (Costello et al. 2020). These stressors often 180 do not act in isolation, but instead interact additively and synergistically to increase the 181 vulnerability of species and communities (Gissi et al. 2021). It is predicted that climate 182 change will soon become the single greatest driver of global biodiversity loss (Newbold 183 2018; He et al. 2019). A primary response of marine ectotherms to changing conditions 184 (particularly temperature rises) is to shift their distribution to track favourable conditions, and 185 regional species turnover rates are far greater within the marine environment than on land 186 (Blowes et al. 2019). Species that are unable to track favourable conditions may experience 187 physiological impacts resulting in fundamental changes such as decreases in size-at-age, 188 changes to fecundity, or even local extinctions (Nikolaou and Katsanevakis 2023; Niu et al. 189 2023). Such shifts in the distribution and local abundance of marine species will alter 190 regional community composition, with impacts on ecosystem structure.

191 Polar regions are particularly under threat, as many endemic species have limited capacity to 192 undergo compensatory distribution shifts due to the shrinking of suitable habitat (such as sea 193 ice or waters within their thermal tolerance range) or the presence of physical barriers (e.g. 194 the Antarctic continental shelf). Polar ecosystems will thus be increasingly impacted both by 195 abiotic and biotic factors, as environmental changes drive the decline of native species and 196 influxes of sub-polar species alter the composition of regional species assemblages. Such 197 biodiversity changes and homogenisation are of great concern from both conservation and 198 management perspectives, as they may increase the vulnerability of ecosystems to further 199 changes and could disrupt ecosystem functioning and the maintenance of ecosystem services 200 (Olden et al. 2004).

# 201 1.1 Biodiversity and ecosystem stability

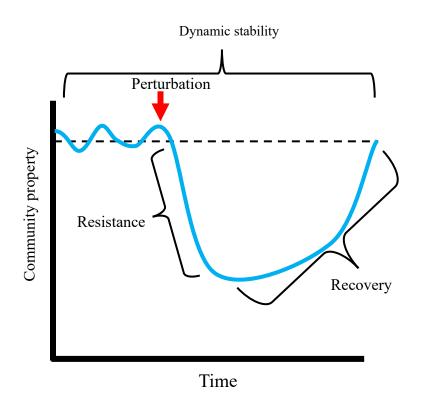
202 Recognition of ongoing biodiversity change has led to a focus on the links between diversity

and the dynamics and stability of communities and maintenance of ecosystem processes.

- 204 Ecological stability has numerous definitions (as reviewed by Ives and Carpenter 2007), the
- 205 most common being the overall temporal variability or amplitude of fluctuations in the focal

206 community properties (dynamic stability), the extent to which discrete perturbations alter 207 these properties (resistance), and the ability or rate at which the properties return to 208 equilibrium (recovery) (McCann 2000; Ives and Carpenter 2007) (Figure 1.1). Species 209 diversity may enhance community stability through mechanisms such as the 'portfolio effect' 210 (statistical averaging) and the 'covariance effect' (whereby negative covariance in abundance 211 of competing species increases with higher diversity) (Tilman and Downing 1994; Tilman et 212 al. 2006). Ecosystem resilience (the combination of resistance and recovery) often increases 213 with diversity. This is due to the greater variety of ecological roles present within the 214 community and the number of species able to perform the same roles (functional 215 redundancy), which increases the overall capacity to tolerate perturbations (Biggs et al. 2020;

216 Yachi and Loreau 1999).



217

218 Figure 1.1: Conceptual visualisation of three common stability measures. The blue line

219 represents a community property (e.g. biomass). Dynamic stability generally describes the

temporal variability or amplitude of fluctuations in the property, resistance measures the

degree of change in the property after perturbation, and recovery represents the rate at which

222 of the property returns to baseline (dashed line) after perturbation, or the relative quantity of

223 the community property that is recovered post-disturbance.

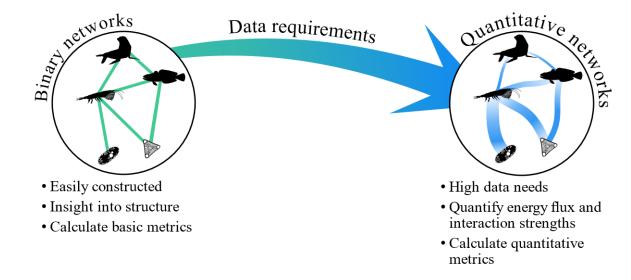
224 Much diversity-stability research has focussed on simple communities spanning single or few 225 trophic levels and may therefore fail to consider important ways in which species diversity in 226 larger, more complex ecological networks can modify ecosystem properties. As discussed by 227 Stachowicz et al. (2007), changes to predator and prey diversity in multi-trophic marine 228 communities can modify diversity-stability relationships through mechanisms such as altered 229 strength of top-down and bottom-up control, and trophic cascades. In the context of climate 230 change, there is evidence that fluctuations in species populations and overall diversity are 231 driven as much by changes in the interactions between species as by direct environmental 232 effects on organisms themselves (Ockendon et al. 2014). If we are to disentangle and predict 233 the likely ecosystem-level responses to changes in species distributions, then it is important 234 that we gain an understanding of how ecological communities are structured and what 235 implication this structure has for ecosystem stability. This approach requires the explicit 236 consideration of species interactions, which can be achieved using food web models.

# 237 *1.2 Food web models*

238 Food web models depict the feeding relationships between species and provide a useful 239 framework for relating the structure of multitrophic communities to ecosystem functioning, 240 given that they incorporate not only their constituent species but also the relevant patterning 241 of interactions (i.e. energy flow) within the system (Thompson et al. 2012; Hines et al. 2019). 242 Models are generally constructed using information on the diet of each species within the 243 focal ecosystem. This data may come from a range of sources including direct observation of 244 feeding interactions, morphological analysis of stomach or scat samples, and methods such as 245 stable isotope analysis or the DNA sequencing of tissue samples (Horswill et al. 2018). The 246 limitations to these different methods are well described. Morphological methods are biased 247 towards identifying prey species with hard structures and the resulting information only 248 represents recent dietary composition, while isotope analysis can identify more long-term diet 249 in addition to providing quantitative estimates of diet reliance but cannot resolve prey 250 taxonomy to the species-level (Nielsen et al. 2017; Horswill et al. 2018). DNA sequencing 251 sits somewhere in-between, as it can resolve recent prey taxonomy without requiring intact 252 prey specimens to be present but fails to provide information on the relative proportions of 253 species in the diet (Nielsen et al. 2017; Horswill et al. 2018). Despite their individual 254 limitations, these methods can provide invaluable contributions to the creation of 255 comprehensive food web models for a wide range of ecosystems, and recent research has 256 begun combining some of these techniques to maximise the reliability of inferred diets (e.g.

Horswill et al. 2018; Bonin et al. 2020). Data on feeding relationships are often also
supplemented with information from the primary literature to infer interactions for species
that are known to occur in the system concerned.

260 There are various approaches to modelling food webs, which exist along a spectrum of 261 capabilities and data requirements (Figure 1.2). At their most basic, food webs can be 262 described using simple unweighted binary network models, whereby species (or groups of 263 taxonomically or functionally similar organisms) are represented by nodes and their 264 interactions are described by the links between them. Such models are relatively easy to 265 construct, as they only require knowledge of which groups consume each other. Despite their simplicity, these models can provide useful insights, with a variety of metrics that can be 266 267 calculated to describe the position and role of individual nodes within the wider ecosystem or 268 provide an understanding of the overall structure of the food web, both of which are relevant 269 to determining the implications of perturbations for ecosystem functioning (Table 1.1). 270 Qualitative network models can be particularly useful in data-poor systems, as they require 271 only a general understanding of the interactions between ecosystem components, and have 272 been used to explore possible ecological consequences of climate change or management activities (Melbourne-Thomas et al. 2013; Forget et al. 2020). 273



- 274
- Figure 1.2: Conceptual diagram highlighting the requirements and capabilities of binary and
- 276 quantitative food web models.

- Table 1.1: List of common node-level and network-level food web metrics, their descriptions,
- and implications for our understanding of food web structure.

Metric	Description	Implication	References
Species richness (S)	Number of nodes within the food web.	Provides information on food web size.	Thompson et al. 2012; Kortsch et al. 2019
Link richness (L) Average food chain length	Number of feeding links between nodes. The number of nodes that energy passes through from the base to the	Provides information on food web size. Provides information about network structure	Thompson et al. 2012; Kortsch et al. 2019 Post 2002; Thompson et al. 2012
	top predator in the food chain.	and organisation of interactions.	al. 2012
Linkage density (L/S)	Average number of links per node (unweighted).	Indicates food web complexity.	Kortsch et al. 2019
Connectance (L/S <sup>2</sup> )	Proportion of total possible trophic links that actually occur in a network (unweighted).	Indicates food web complexity.	Thompson et al. 2012
Clustering coefficient	For a given node, indicates the degree to which connected nodes are also linked to each other. At the network level, indicates the average level of clustering.	Indicates food web complexity and structure.	Kortsch et al. 2019; Marina et al. 2018
Degree	Number of incoming and outgoing links to a specific node.	Indicates how connected the node is. Used to estimate generality and vulnerability.	Thompson et al. 2012; Kortsch et al. 2019; Marina et al. 2018
Degree distribution	Overall frequency distribution of the number of interactions for each node in the network	Can help identify species/groups with important roles in connecting the community.	Thompson et al. 2012; Kortsch et al. 2019; Marina et al. 2018
Generality	Number of prey consumed by a node. At network scale, estimated as the mean number of prey per consumer.	Indicates node/network sensitivity to bottom-up processes influencing prey dynamics.	Thompson et al. 2012
Vulnerability	Number of predators of a given node. At network scale, estimated as the mean number of consumers per prey.	Indicates node/network sensitivity to top-down processes influencing predator dynamics.	Thompson et al. 2012
Modularity	How densely nodes within subgroups interact with each other compared to with nodes from other subgroups.	Provides an indication of the structure and organization of links within the food web.	Grilli et al. 2016; Kortsch et al. 2019
Proportion of basal, intermediate, and top species	Fraction of species with no resources, with both consumers and resources, and with no consumers, respectively.	Provides an indication of food web structure and possible top-down or bottom-up control.	Kortsch et al. 2019; Gibert 2019
Omnivory	Variety in prey trophic level for a given consumer node.	Proportion of omnivory has implications for network stability and energy flow.	McCann and Hastings 1997; Kratina et al. 2012; Heymans et al. 2014

282 More complex models can provide additional knowledge about structure and patterns of 283 energy transfer that is not available in unweighted models. For example, these weighted 284 networks can reveal the energy flux between groups and capture the strengths of interactions 285 (Marina et al. 2024; Gauzens et al. 2019). The development of such quantitative models 286 comes with greater data requirements, such as abundances, body masses, metabolic rates and 287 relative dietary contributions, limiting their application to regions where such information is 288 available. Modelling frameworks such as Ecopath with Ecosim are used to construct models 289 of energetic fluxes that meet the assumption of mass-balance over a specific time period 290 (often a single year) (Christensen and Walters 2004). Ecopath models lie at the far end of the 291 complexity and data needs scale, with key parameters including the biomass, diet 292 composition by weight, production per unit biomass, consumption per unit biomass and 293 assimilation efficiency of each group, while additional parameters representing biomass 294 accumulation rates, fishery catches and discards, and migration rates can also be supplied 295 (Christensen and Walters 2004). Parameters are ideally location- and group-specific but can 296 also be taken from other models or literature where necessary. These form the basis of linear 297 equations underlying the production of each group, as determined both by their parameters 298 and those of their consumers. A key parameter which is often an output of Ecopath models is 299 the ecotrophic efficiency (*EE*), representing the proportion of the production or mortality of 300 each group that is explained in the model. Values range from zero (limited to top predators 301 that are not fished) to one (100% of production is consumed by other groups in the model). 302 As such, the *EE* is a key parameter representing whether or not the model is in balance, with 303 values above one indicating groups with mortality rates that cannot be sustained by 304 production rates. Extensive adjustments to group parameters must often be made before the 305 assumption of mass balance ( $EE \le 1$ ) is met for all groups. Balanced Ecopath models 306 represent a powerful tool for exploring topics including the direct and indirect impacts of 307 fisheries on trophic interactions (Coll et al. 2006; Subramaniam et al. 2020), the 308 consequences of climate change for aquaculture (Chapman et al. 2020), and the effect of 309 multiple climate and anthropogenic stressors on trophic dynamics (Stock et al. 2023). 310 Additionally, the inclusion of temporal population trends for key groups can also facilitate the 311 development of time-dynamic models using the Ecosim plugin, and spatially resolved models 312 can also be constructed (Christensen and Walters 2004).

# 313 *1.3 The drivers of food web structure*

Early research into the relationship between diversity and stability used random matrices to describe the structure of species communities (e.g. May 1973). We now know that complex networks are non-randomly structured, and that their topology may be influenced by a variety of factors.

318 A major driver of the structure of food webs is the distribution of functional traits across the 319 community (Gravel et al. 2016; Brose et al. 2019). Predator and prey traits influence both the 320 likelihood of co-occurrence of interacting species, and the ability of each species to capture 321 and consume, or escape, the other. Marine food webs are often strongly size-structured, as 322 body size determines gape limitation, prey density, handling time and energy content 323 (Petchey et al. 2008; Rall et al. 2012; Potapov et al. 2019). The relative size of predators to 324 their prey (the predator prey mass ratio, PPMR), can be used to predict food web structure 325 (Petchey et al. 2008; Morales-Castilla et al. 2015; Laigle et al. 2017). PPMR generally 326 decreases with trophic height of the consumer (Jonsson and Ebenman 1998; Tucker and 327 Rogers 2014), and may also predict the strength of trophic interactions, with a positive 328 relationship between PPMR and interaction strength identified for some consumer-resource 329 pairs (Emmerson and Raffaelli 2004; Woodward et al. 2005). Body size is also often related 330 to important life-history traits such as generation time and reproductive rate (Stearns 1983; 331 Janis and Carrano 1991; Gillooly 2000) and is therefore also strongly linked to abundance, 332 which typically declines as size increases (White et al. 2007). This relationship, in 333 combination with the fact that predator body masses are generally one to three orders of 334 magnitude greater than those of their prey (Woodward et al. 2005), often results in a 335 pyramidal structure of declining abundance with increasing trophic level (Cohen et al. 2003; 336 Jacquet et al. 2020). Further traits such as taxonomy, prey type, habitat type, and motility can 337 also be used to infer interactions (Morales-Castilla et al. 2015; Laigle et al. 2017; Brose et al. 338 2019), and phylogeny, feeding habitat and feeding method have been found to explain food 339 web structure in various marine communities (Rezende et al. 2009; Jacob et al. 2011; Cirtwill 340 and Eklöf 2018).

341 As previously discussed, environmental changes are driving extensive reorganisation of

342 marine communities, so abiotic factors can clearly also have a major influence on food web

343 structure. Gradients of seawater temperature, depth, and days of ice cover are associated with

344 significant variation in local food web metrics, and food web complexity (i.e. the number of

345 species and interactions) increases with local habitat heterogeneity (Kortsch et al. 2019).

346 Similarly, warming may drive declines in the proportion of basal species, and changes in 347 metrics such as connectance and omnivory level (Gibert 2019). Within both terrestrial and 348 aquatic food webs, warming and nutrient enrichment are expected to drive significant 349 changes in producer and consumer activity and biomass, altering the patterns of energy flow 350 within the communities (O'Connor et al. 2009; Sentis et al. 2017; O'Gorman et al. 2017; 351 2019). The greater metabolic demands associated with warming may drive declines in the 352 body size distribution of many marine communities, through changes in species composition 353 and size at age (Coghlan et al. 2024; Saunders and Tarling 2018). Larger organisms may 354 decline in size more rapidly than smaller organisms, due to reduced surface area to body mass 355 ratios and the associated challenge of maintaining a higher metabolic rate (Forster et al. 2012; 356 Petrik et al. 2020). As a result, warming may drive substantial changes to PPMR and the size 357 structure of communities, with implications for the patterning of interaction strengths and 358 energy flow. Ultimately, changes to the distribution of many marine organisms may result in 359 the gain or loss of species with key traits and disproportionately large influence on food web 360 structure and ecosystem functioning. These changes could disrupt key structural features and 361 might reduce the stability of food webs.

## *362 1.4 Food web stability*

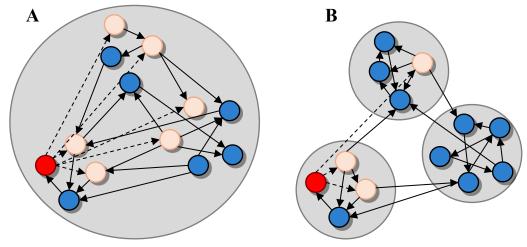
363 Several different mechanisms have been found to influence the stability of food webs. One 364 such driver is the pattern and coupling of weak and strong interactions. Interaction strength 365 quantifies the magnitude of the effect that individual species have on one another (Berlow et 366 al. 1999). Food webs are dominated by weak interactions, which provide stability by dampening population oscillations between consumers and their resources (Paine 1992; 367 368 McCann et al. 1998). As a result, removal of either strong or weak interactors can reduce the 369 temporal stability of ecosystem processes and resistance to changes in community 370 composition (O'Gorman and Emmerson 2009; 2010). In general, highly connected generalist 371 species display weaker net effects than specialist predators, which rely heavily on a small 372 number of food sources (Montoya et al. 2009; O'Gorman and Emmerson 2010; Wootton and 373 Stouffer 2016). Larger body size ratios often lead to greater interaction strengths, therefore 374 the size distribution of the different components of the food web may be an important 375 determinant of system stability (Woodward et al. 2005). The coupling of different energy 376 channels (e.g. from primary production versus detritus) can also enhance the dynamic 377 stability of higher consumers, by generating asynchrony in energy flux (Blanchard et al. 378 2011). Similarly, larger mobile consumers may also drive food web flexibility and stability

when basal resource availability fluctuates asymmetrically, either spatially or temporally, by
coupling multiple distinct sub-food webs (McCann et al. 2005; McCann and Rooney 2009;

381 Mougi 2018).

382 There has been much focus on the organisation of nodes into 'modules': subgroups of species 383 that interact often with one-another but have few connections to species outside their 384 subgroup. This structuring has been identified in a variety of social and biological networks 385 including food webs (Newman and Girvan 2004; Krause et al. 2003; Rezende et al. 2009; 386 Pérez-Matus et al. 2017; Zhao et al. 2017), although the prevalence of modules in marine 387 food webs has recently been disputed (Marina et al. 2018). Modularity has been found to increase stability in both theoretical and modelled empirical food webs by restricting the 388 389 propagation of secondary extinctions to the subgroups in which the initial species loss occurs 390 (Teng and McCann 2004; Thébault and Fontaine 2010; Stouffer and Bascompte 2011; Zhao 391 et al. 2017) (Figure 1.3). Stability in this context is generally estimated as robustness using 392 the R50 value, the proportion of species that need to be primarily removed to cause 50% of 393 the food web to become extinct (Jonsson et al. 2015). A value of 0.5 indicates a robust food 394 web, as no secondary extinctions occur (Figure 1.3). The relationship between modularity 395 and robustness to species extinctions is not necessarily clear-cut, however, as the degree to 396 which it promotes network stability may depend on spatial scale, with smaller systems 397 benefiting the most from this structure (Mougi 2018). Few papers have explicitly investigated 398 the mechanisms driving modular structure, but species' traits appear to play an important 399 role. Modules have been characterised using species' niche organisation and diets (Guimera 400 et al. 2010), and may contain separate trophic groups (clusters of species with common prey 401 and predators) (Gauzens et al. 2015). Furthermore, modularity has previously been linked 402 with body size distributions and phylogeny, and further traits such as foraging mode and 403 habitat are also suggested to contribute to this structure (Rezende et al. 2009; Kortsch et al. 404 2015).

405





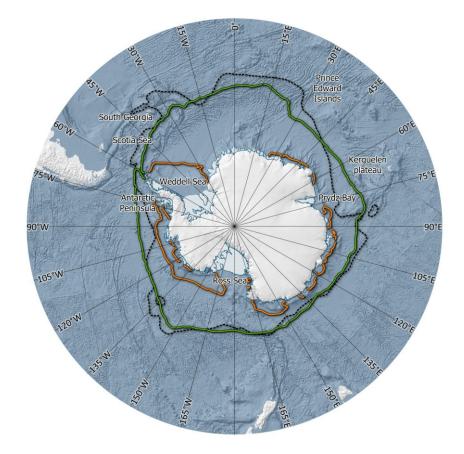
407 Figure 1.3: The theoretical stabilising effects of modularity. A species extinction (red node) 408 could potentially drive secondary extinctions in species that prey on it directly (pale orange 409 nodes connected by dashed line). A) In a non-modular food web, the high inter-410 connectedness of species means that secondary extinctions are likely to propagate widely 411 throughout the network. B) In a modular food web, the low number of links between modules 412 means that secondary extinctions are primarily restricted to the same module as the initial 413 extinction. Results of random extinction analysis confirm that the more modular network, 414 with an R<sub>50</sub> value of 0.46, exhibits greater robustness than the non-modular food web, which 415 has an  $R_{50}$  of 0.31.

# 416 *1.5 The Southern Ocean*

Given the mounting pressures of climate change and human activities, it is imperative that we establish which biotic and abiotic factors drive the structure of different marine food webs, and how this structure relates to their stability. This is particularly true in polar regions, where our understanding of the ecology of many species and the structuring of different food webs is still limited.

422 The Southern Ocean plays a key role in global oceanic circulation and nutrient distribution, 423 linking the world's major ocean basins (Carter et al. 2008). A major feature is the Antarctic 424 Circumpolar Current (ACC), a wind-driven, eastward-flowing current that surrounds the 425 continent (Carter et al. 2008) (Figure 1.4). The ACC acts as a thermal barrier, maintaining 426 stable low water temperatures in the Southern Ocean, particularly in high-latitude shelf 427 regions where temperatures range from around +2 °C down to -1.9 °C (Weiss et al. 2012; 428 Mintenbeck 2017). As a result, many Antarctic organisms are highly stenothermal (Peck et al. 429 2014). A further major physical feature of the Southern Ocean is sea ice. The Southern Ocean

- 430 can be split into broad zones based on their differing sea ice dynamics and coverage
- 431 (Deppeler and Davidson 2017). At high latitudes, sea ice coverage is nearly complete and
- 432 persists throughout the year, while areas further north experience seasonal fluctuations
- 433 between winter maxima and summer minima (Convey et al. 2009; Deppeler and Davidson
- 434 2017) (Figure 1.4). Further north still, the open ocean zone remains almost completely ice-
- 435 free throughout the year (Deppeler and Davidson 2017). Sea ice plays an important role in
- 436 structuring the water column and driving the formation of water masses (Cherkasheva et al.
- 437 2014). It also acts to limit the penetration of light into surface waters, which, in conjunction
- 438 with the strong seasonal variation in light conditions, means that light availability is a major
- 439 factor affecting Southern Ocean ecosystems (Park et al. 2017).



- 441 Figure 1.4: The Southern Ocean and Antarctic continent. Solid orange and green lines
- indicate the median February minimum and September maximum sea ice extent 1981-2010,
- 443 respectively. Dashed black line represents the general position of the Southern Antarctic
- 444 Circumpolar Current front. All map layers obtained from Quantarctica 3.0 (Matsuoka et al.
- 445 2021) and displayed in Antarctic Stereographic projection. Text labels identify locations of
- 446 particular relevance to this thesis.

447 Southern Ocean food webs have traditionally been viewed as relatively simple and centered 448 primarily around the Antarctic krill (Euphausiia superba). This species is highly abundant in 449 regions such as the south-west Atlantic Ocean, and acts as prey for many species including 450 baleen whales, penguins, seals, squid and fish (Trathan and Hill 2016). Other zooplankton are 451 also important components of regional Southern Ocean ecosystems, such as the smaller ice 452 krill (Euphausia crystallorophias) which is abundant in the permanent ice zone (Thomas and 453 Green 1988), hyperiid amphipods such as Themisto gaudichaudii, and the many copepod 454 species which can represent over 45% of pelagic filter-feeder biomass in some areas 455 (Voronina 1998; Kouwenberg et al. 2014). While some top predators may feed directly on the 456 zooplankton component, squid and fishes make up another major link in energy flow within 457 the Southern Ocean ecosystem. The Southern Ocean fish community has relatively low 458 diversity, with only 322 recognised species relative to the ~28,000 fish species known 459 globally, but their contribution to the Antarctic marine ecosystem is substantial (Eastman 460 2004). Most species are demersal, found on and around the continental shelves, and display a 461 variety of different feeding strategies and diets ranging from purely planktivorous to 462 omnivorous and in some cases primarily piscivorous (Casaux and Barrera-Oro 2013; 463 Bansode et al. 2014). Mesopelagic lanternfish (family Myctophidae) are also a key 464 component of Southern Ocean food webs, with a biomass of potentially over 200 million tonnes, and may represent a key krill-independent pathway of energy flow to higher trophic 465 466 levels (Saunders et al. 2018; 2019; McCormack et al. 2020). Clearly, while it is true that 467 Antarctic krill are a central component of many ecosystems, the complexity of Southern 468 Ocean food webs has been underestimated. If we wish to assess the impacts of environmental 469 and anthropogenic change on these ecosystems, it is important that we gain a better 470 understanding of their structure and dynamics.

471 The popular view of Antarctica and its surrounding waters is often that of a pristine, 472 undisturbed landscape. In fact, the Southern Ocean has been subject to significant human 473 activities. Exploitation of Antarctic fur seals and Southern elephant seals during the early 474 19th century resulted in the near extinction of their populations on sub-Antarctic islands like 475 South Georgia, although their populations have subsequently grown rapidly (Miller 1991; 476 Hucke-Gaete et al. 2004). The early 20th century then marked the beginning of the 477 commercial exploitation of whales within the Southern Ocean, which drove severe declines 478 in their abundance (Miller 1991). Some whale species have recently shown evidence of 479 recovery (Zerbini et al. 2019; Calderan et al. 2020), but populations of others have displayed

480 much slower recovery rates and are still at only a fraction of their pre-exploitation numbers 481 (Crespo et al. 2019; Tulloch et al. 2019). The major declines in the abundance of these top 482 predator species are likely to have had significant effects on the wider regional food webs by 483 altering levels of top-down regulation and potentially reducing the competitive pressure on 484 other predator species. The ongoing recovery of many whale species may therefore represent 485 further changes in energy flow within Southern Ocean ecosystems, as their predatory and 486 competitive influence is restored. Exploitation in the Southern Ocean has since shifted to 487 focus on finfish such as toothfish (Dissostichus mawsoni and D. eleginoides) and mackerel 488 icefish (Champsocephalus gunnari), in addition to a large krill fishery which operates 489 primarily in the South Atlantic (Kock et al. 2007). These fisheries are managed by the 490 Commission for the Conservation of Antarctic Marine Living Resources (CCAMLR), which 491 practices an ecosystem approach to management involving regular monitoring of target and 492 non-target species and abiotic conditions to minimise human impacts on ecological 493 relationships and conserve Antarctic species and ecosystem functioning (Kock et al. 2007). 494 Despite this careful approach, these fisheries still represent an important additional pressure 495 on Antarctic marine ecosystems, and their potential future expansion may have significant 496 implications for the functioning of Southern Ocean food webs, particularly when viewed in 497 combination with the effects of climate change.

498 It is becoming increasingly clear that the Antarctic is vulnerable to the effects of climate 499 change. The Southern Ocean has displayed regional variation in temperature trends, with 500 some areas remaining stable or cooling while others, such as the western Antarctic Peninsula 501 and Scotia Sea, have warmed significantly (Meredith and King 2005; Whitehouse et al. 502 2008). Similarly, there has been regional variation in sea ice trends, with rapid declines in sea 503 ice extent and concentration in some areas and increases in others (Parkinson 2019). These 504 changes have serious implications for regional ecosystems. Changes to the timing and extent 505 of seasonal sea ice expansion and contraction, and strengthened water column stratification 506 following ice melt, could alter the timing and magnitude of primary production, potentially 507 reducing the amount of food available for consumers (Quetin et al. 2007). Reductions in the 508 availability of sea ice could also directly impact the populations of key mid-trophic species 509 such as Antarctic krill and the Antarctic silverfish, *Pleuragramma antarcticum*, as they use this substrate for refuge and spawning (Massom and Stammerjohn 2010; La Mesa and 510 511 Eastman 2011). Temperature rises and sea ice declines have already driven a southward 512 contraction in the distribution of Antarctic krill, while fish species such as the Antarctic

toothfish are also expected to exhibit range shifts in the future (Cheung et al. 2008;

- 514 Kawaguchi et al. 2024). Ocean warming is also expected to drive range shifts in the benthos
- 515 due to the thermal sensitivity of many benthic species (Barnes et al. 2009). Temperature-
- 516 driven shifts in body size could alter the structure of communities, while changes in the
- 517 dominance of certain mid-trophic species (e.g. krill versus the tunicate *Salpa thompsoni*)
- 518 might reduce the efficiency of energy flow to higher trophic levels (Pauli et al. 2021; Pietzsch
- 519 et al. 2023). Warming and reductions in sea ice cover could also facilitate the influx and
- 520 successful establishment of invasive species, which will lead to changes in community
- 521 composition and novel interactions (Morley et al. 2020; Queirós et al. 2024).

522 If we are to manage Southern Ocean ecosystems effectively in the face of the varied 523 anthropogenic and environmental threats discussed above, then we must improve our 524 understanding of the drivers of food web structure and their likely responses to change. Much 525 previous research on Southern Ocean ecosystems has involved species-specific models of 526 habitat-use or population dynamics, and simple trophic models (see McCormack et al. 2021a, 527 and references therein). Various ecological network modelling studies have also been 528 conducted to investigate the structure and dynamics of regional food webs. To date, food web 529 models have been developed for various Southern Ocean regions (e.g. Ballerini et al. 2014; 530 Hill et al. 2012; López-López et al. 2021; Jacob et al. 2011; Pinkerton and Bradford-Grieve 531 2014; McCormack et al. 2020, amongst others). Such models have provided insights into 532 regional food webs, including the major energy pathways (McCormack et al. 2020; 533 McCormack et al. 2021b), the importance of environmental variables for structuring 534 networks (López-López et al. 2021; Rossi et al. 2019), and the association between species' 535 functional traits and network structure (Jacob et al. 2011). Researchers have also modelled 536 the possible ecosystem responses to scenarios of climate change and anthropogenic activities 537 including the historic exploitation of baleen whales (Surma et al. 2014) and future changes in 538 primary production (Ballerini et al. 2014), declines in the biomass of Antarctic krill (Hill et 539 al. 2012), and increased exploitation of Antarctic toothfish (Pinkerton and Bradford-Grieve 540 2014).

- 541 There remain a variety of knowledge gaps surrounding the structure of Southern Ocean food
- 542 webs and their likely responses to future change. In particular, there is still a poor
- 543 understanding of the role that body size, mobility, feeding-mode and other morphological and
- 544 behavioural traits play in driving the structure of Southern Ocean food webs. Additionally,
- 545 identifying the presence and distribution of stabilising features such as modularity, and

determining their underlying drivers, will bring our knowledge of Southern Ocean food webs more in line with wider food web theory and improve our understanding of how robust they are to perturbations. There is also a need to investigate what the consequences of changes in environmental and ecological conditions will be for ecosystem structure. Comparisons of regional food webs are still largely lacking and would provide us with a clearer understanding of the resilience of different Southern Ocean ecosystems to the above changes, which would improve our ability to implement suitable management and conservation measures.

# 553 *1.6 Thesis outline*

This thesis combines a number of approaches from theoretical and empirical ecology and ecosystem modelling at a variety of spatial scales to provide insight into the structure of Southern Ocean food webs and their possible responses to environmental and ecological change. The component data chapters can be split into two main themes, encompassing the relationships between functional traits and trophic structure and the impacts of changing ecological and environmental conditions on communities and ecosystems, although these themes overlap for some chapters, as visualised in Figure 1.5.

561 **Chapter 2** focusses on the demersal fish community around South Georgia, using stomach 562 content analysis and morphological measurements to identify how simple morphological 563 traits map onto broad feeding guilds. This is a novel approach within Southern Ocean food 564 webs which provides a baseline understanding of how morphological traits underlie the 565 ecology of demersal fish and highlights the role that krill may play in bridging ecological 566 niches imposed by morphology.

567 Chapter 3 expands upon the trait-based approach, applying it to a comparison of four highly 568 taxonomically resolved food webs across the Southern Ocean and northern hemisphere to 569 determine how functional traits (both morphological and behavioural) underlie the 570 distribution of modules. Traits such as body size, foraging habitat, feeding mode and mobility 571 are found to be good predictors of module membership. Differences in the relative 572 importance of traits, and in the structuring of modules across trophic levels, are postulated to

573 be tied to the degree of habitat heterogeneity between systems.

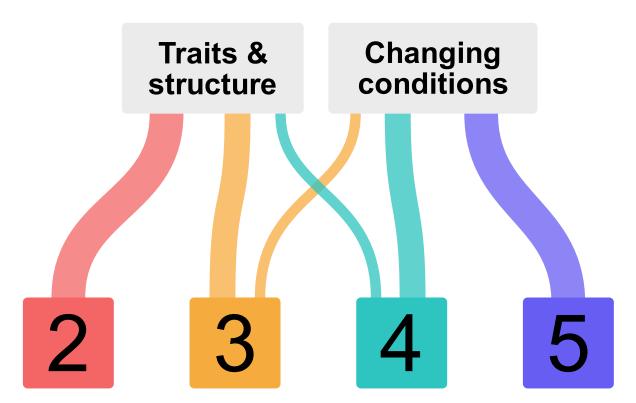
574 Chapter 4 then focuses on how warming alters body size relationships in the myctophid

575 community of the Scotia Sea, using an extensive dataset of myctophid stomach contents and

576 environmental zooplankton samples. A clear decline in predator-prey mass ratio (PPMR) with

577 warming is driven by shifts in the size distribution of both the myctophids and their prey. This

- 578 provides insight into the possible implications of environmental change for the size structure 579 and distribution of interaction strengths within Southern Ocean food webs.
- 580 **Chapter 5** explores the possible ecological consequences of the recovery of Southern Ocean
- 581 baleen whale populations and implications for management. This makes use of a suite of
- 582 published regional Ecopath models, which were then standardised and subject to a custom
- 583 balancing algorithm to generate ensembles of plausible alternative models. Clear trade-offs
- between whale recovery and the biomass of key competitors are identified, and the potential
- 585 structural features and environmental factors which could mitigate these are discussed.
- 586 Chapter 6 provides a synthesis of the insights gained in the previous chapters and puts them
- 587 into context of the wider state of knowledge regarding contemporary and future Southern
- 588 Ocean food webs.



- 590 Figure 1.5: Conceptual diagram displaying the main linkages between the primary themes of
- this thesis and each of the four data chapters. The thickness of the links indicates the extent to
- 592 which the chapters fall within each theme.

# 593 2 Morphological traits distinguish feeding guilds in a 594 Southern Ocean fish community

595 Submitted to Functional Ecology

596 Abstract

597 Morphological traits reflect an organism's ecological niche and role within ecosystems, thus 598 improving our understanding of the drivers of community structure. Here, we combined 599 morphological measurements with stomach contents analysis of nine demersal fish species 600 from the subantarctic island of South Georgia, where climate change has already affected the 601 distribution of a key prey species, Antarctic krill. Although most species include krill in their 602 diets, cluster analysis identified five distinct feeding guilds, with traits such as gape size 603 proving especially useful for determining guild membership. Individuals feeding primarily on 604 fish had larger gapes and higher caudal and pectoral fin aspect ratios, enhancing their ability 605 to capture and consume such large, fast prey. In contrast, benthic feeders had smaller gapes 606 and lower fin aspect ratios, reflecting their reliance on suction feeding and higher 607 manoeuvrability. Continued research into the relationship between morphology and diet will 608 improve understanding of the drivers of trophic dynamics in marine ecosystems and aid our 609 ability to predict the effects of environmental change on community composition and 610 structure.

## 611 2.1 Introduction

612 The field of ecology is increasingly focusing on how complex interactions between

613 individuals shape the structure and functioning of ecosystems (Åkesson et al. 2021). A key

614 component of this approach is the consideration of how functional traits including

615 physiological, morphological, behavioural and life history attributes shape how organisms

respond to each other and to their environment (Violle et al. 2007). This trait-based approach

617 to ecology seeks to identify how the functional traits of organisms combine to determine their

618 interactions and thus drive the organisation of ecological communities. By describing the

619 distribution of traits within ecosystems, it is possible to generalize the mechanisms

620 underlying complex ecological processes and predict the resilience of key ecosystem

621 functions to perturbations (Kiørboe et al. 2018).

622 Ecomorphology is a key component of trait-based ecology, whereby an individual's body

623 form is linked to its behaviour and interactions with others (Barr 2018). An organism's

- 624 physical features represent adaptation to its environment, and therefore the combination of
- 625 different morphological traits largely underlie its ecological niche. As such, morphological
- traits may be strongly tied to the distribution and functional role of organisms within
- 627 ecosystems, e.g. wing shape and beak dimensions strongly predict diet and foraging niche in
- 628 birds (Pigot et al. 2016); eye size is linked to adult habitat and activity level in amphibians
- 629 (Thomas et al. 2020); whilst diet and habitat preferences are driven by traits including body
- 630 size, eye position and head shape in ants (Gibb et al. 2015).
- 631 Marine ecosystems are often strongly size structured due to gape limitations and the interplay 632 between body size and feeding (Jennings et al. 2001; Potapov et al. 2019). In fish, gape size 633 often reflects feeding mode, with ambush piscivores generally exhibiting large gapes while 634 suction feeding planktivores tend to have small gapes (Luiz et al. 2019). Gape size also often 635 constrains the maximum size of prey that can be consumed, thus determining the structure of 636 feeding relationships (Christensen 1996). Allometric scaling relationships have been 637 identified for tropical and temperate fish species, whereby gape size, and thus also average 638 and maximum prey size, generally increase with predator body size (Bachiller and Irigoien 639 2013; Dunic and Baum 2017). In many species maximum prey size increases more rapidly 640 than minimum prey size as fish become larger, resulting in a widening of their trophic niche 641 (Scharf et al. 2000). Differences in allometric relationships for body size and gape 642 morphology could also influence levels of resource partitioning and competitive interactions 643 within the fish community (Schuckel et al. 2012; Barnes et al. 2021). Other traits may also be 644 important, such as fin morphology which is linked to habitat use and prey acquisition: e.g. 645 high aspect ratios of the caudal and pectoral fins are linked with greater swimming efficiency 646 and maximum speed (Higham 2007; Sambilay 1990) and generally found in more active 647 species that feed on pelagic or mobile prey such as zooplankton and fish (Bridge et al. 2016; 648 Hobson 1979). Lower aspect ratios provide greater manoeuvrability and thrust at low speeds 649 and may therefore be better suited to less active benthic or ambush feeding (Higham 2007; 650 Bridge et al. 2016).
- While an increasing number of marine studies use trait-based approaches, these often involve competition models focused primarily on basal groups and overlook trophic interactions between predators and their prey (Kiørboe et al. 2018). There is therefore a need to further describe the traits driving trophic relationships, particularly in remote and understudied marine ecosystems. Here we describe the relationships between morphology and diet for nine of the most abundant demersal fish species around the sub-Antarctic island of South Georgia

657 in the Atlantic sector of the Southern Ocean. Previous research on the diet and biology of 658 these species has revealed a system largely dominated by consumption of Antarctic krill, 659 *Euphausia superba*, in addition to fish and macrozooplankton such as amphipods, 660 particularly in periods of low krill availability (Kock et al. 2012). There is some evidence of 661 interspecies dietary differentiation (McKenna 1991; Targett 1981), but to date there has been 662 no comprehensive investigation of the links between morphological traits and dietary niches 663 across the wider groundfish community. Such information will improve our understanding of 664 the mechanisms underlying community structure and energy flow through this component of 665 demersal food webs. Many marine species at South Georgia are at the northern edge of their 666 distributions and may therefore be vulnerable to ocean warming, which has been particularly 667 rapid in this region (Whitehouse et al. 2008). A southward range contraction by E. superba 668 has already been observed (Kawaguchi et al. 2024) and changes in the dynamics and 669 distribution of other zooplankton groups might also be expected (Whitehouse et al. 2008), 670 ultimately driving significant changes in community composition and associated feeding 671 interactions. Identifying how morphological traits influence prey selection will provide 672 insight into the possible consequences of such shifts in prey availability for community 673 structure. The general relationships between feeding ecology and morphology identified here 674 will also be broadly applicable to other regions, furthering our ability to generalize the drivers 675 of marine ecosystem assembly.

We hypothesise that dietary differences between and within species are explained by differences in their functional traits. We expect that predators with larger gape sizes and higher fin aspect ratios primarily consume fish and krill due to their ability to capture and consume such larger, mobile prey, while those feeding on smaller, less mobile prey such as benthic invertebrates or amphipods display smaller gape sizes and lower fin aspect ratios to provide the necessary manoeuvrability for benthic foraging.

682 *2.2 Materials and methods:* 

# 683 2.2.1 Sample collection:

Sampling of the groundfish community was conducted from the FV *Robin M Lee* over the
South Georgia and Shag Rocks shelves between the 1<sup>st</sup> and 10<sup>th</sup> of February 2023 as part of
the biennial groundfish survey conducted by the British Antarctic Survey and the
Government of South Georgia and the South Sandwich Islands. The survey utilises a random
stratified design across five shelf areas and two depth strata (100-200m and 200-350m). A

- total of 47 bottom trawls were completed using an FP-120 net (Caedmon Nets, UK; Figure
- 690 2.1). See Hollyman et al. (2023) for further details on sampling methodology. For this study,
- 691 nine fish species were sampled: icefish (Champsocephalus gunnari, Chaenocephalus
- 692 aceratus, and Pseudochaenichthys georgianus); rockcods (Notothenia rossii, Trematomus
- 693 hansoni, Lepidonotothen squamifrons, L. larseni, and Gobionotothen gibberifrons); and
- 694 dragonfish (*Parachaenichthys georgianus*). Fish were opportunistically sampled from
- 695 catches, with efforts made to choose specimens representing a range of body lengths for each
- 696 species. Selected individuals were frozen at -20°C for later analysis at King Edward Point
- 697 research station, South Georgia.

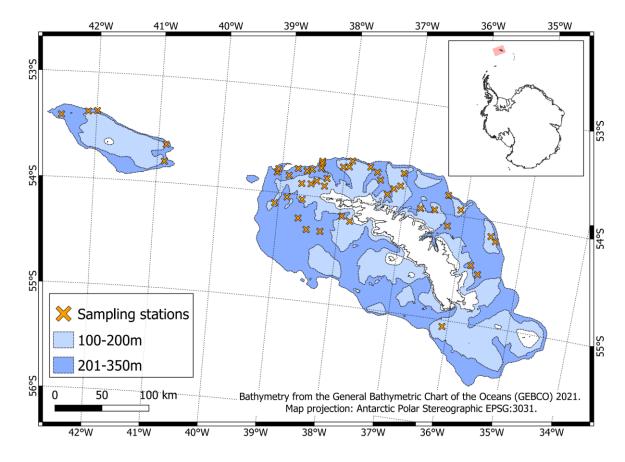


Figure 2.1: Distribution of haul locations, identifying the two depth zones sampled. Inset map
displays the sampling region (red rectangle) in the context of the wider Antarctic continent.
Map generated in QGIS 3.28.0-Firenze.

- 702 2.2.2 Morphological measurements and stomach contents dissection:
- 703 In the laboratory, each fish was thawed before being weighed using either a small (Kern,
- PCB1000-2, +/- 0.01 g) or large (M3, WPL industries, +/- 1 g) top-loading scale depending
- on the size of the fish. For large fish (>400 mm total length, *TL*), measurements of *TL* were

- recorded using a fish board and dissecting ruler. All other specimens were photographed
- vising a Sony RX100i digital camera mounted on a copy stand (Kaiser R2N), with length later
- 708 measured in ImageJ software (Schneider et al. 2012). Gape measurements to the nearest mm
- 709 were taken for each specimen using Vernier callipers for maximum vertical gape ( $G_{height}$ ) and
- a dissecting ruler for maximum horizontal gape ( $G_{width}$ ). The gape height and width of each
- 711 fish were then combined to estimate the maximum oral gape area ( $G_{area}$ ) using the following
- 712 equation (Ward-Campbell et al. 2005):

713 
$$G_{area} = \pi (0.5G_{height} * 0.5G_{width})$$

- 714 Photographs were also taken of each specimen's caudal and pectoral fins, with the latter
- excised at the fin base and laid flat. The aspect ratio (AR) of each fish's caudal and pectoral
- 716 fins was estimated using the following equation:

717 
$$AR = \frac{h^2}{a}$$

where *a* is the fin area (in mm) and *h* is either the caudal fin height or length of the leading
edge of the pectoral fin, measured in ImageJ (Pauly 1989).

Each fish stomach was dissected, and non-empty stomachs were weighed to the nearest 0.01
g. Stomach contents were grouped according to the lowest identifiable taxonomic level,
weighed, and counted, excluding fish prey displaying no evidence of digestion (likely to be
the result of net feeding). Where stomachs contained many individuals of a prey group, 30
individuals were subsampled and weighed, and the total number of individuals in the stomach
was estimated.

726 To investigate potential ontogenetic shifts in diet, each individual fish was assigned to a size 727 class, estimated by splitting the range of sampled TL across the community into four size bins 728 of 176 mm. These were numbered 1 to 4 in ascending size order. This split, whilst arbitrary, 729 provided the best balance of sample sizes across size classes for most species. We defined 730 size classes at the community level rather than at the species level to ensure that size classes 731 were comparable across species. The relative importance of each prey group in the diet of 732 each species-size class combination was estimated from three separate measures of 733 importance using the % Index of Relative Importance (%IRI), calculated as:

734 
$$\% IRI_i = \frac{(\% N_i + \% W_i) * \% FO_i}{\sum_{i=1}^n (\% N_i + \% W_i) * \% FO_i} * 100$$

- 735 Where %FO is the percentage frequency of occurrence, %N is the proportional abundance
- and %W is the proportional weight of each prey group in the diets of each species-size class
- 737 (Pinkas et al. 1970). We set a minimum sample size of five non-empty stomachs, resulting in
- the exclusion of the largest and smallest size classes of *Parachaenichthys georgianus* and
- 739 *Chaenocephalus aceratus*, respectively (n = 2 in both cases).
- 740 2.2.3 Identification of feeding guilds
- All analyses were conducted using R statistical software version 4.3.0 (R Core Team 2023;
- see Table A1 for an overview of the various packages used). Species-size classes were
- 743 grouped into feeding guilds with hierarchical cluster analysis, using Bray-Curtis
- 744 dissimilarities calculated from the prey %IRI values. Prey were grouped into eight broad
- taxonomic groups: 1) krill (all members of the Euphausiidae); 2) Themisto gaudichaudii (an
- abundant swarming amphipod); 3) other non-swarming amphipods (primarily Vibilia sp.,
- 747 *Primno macropa*, and individuals of superfamily Lysianassoidea); 4) isopods; 5) fish; 6)
- 748 mysids; 7) benthic decapods (*Notocrangon sp.* and *Chorismus sp.*); 8) miscellaneous benthos
- 749 (including polychaetes, annelids, bivalves, gastropods, and echinoderms which were
- 750 sporadically found in stomachs). Differences between assigned feeding guilds were identified
- vising the similarity percentage routine (SIMPER).

# 752 2.2.4 Predator-prey size relationships

753 We explored the relationship between predator mass and average prey mass using a linear 754 mixed effects model. This model included the count-weighted average prey body mass (log<sub>10</sub> 755 g) of each prey type within each stomach as a response, and predator body mass  $(\log_{10} g)$  and 756 feeding guild plus their interaction as predictors, to identify predator-prey size relationships 757 specific to different dietary groups. Prey type was included as a random effect to account for 758 potential differences in size relationships across prey taxa, and different covariate weighting 759 structures were investigated to account for any systematic variance in the residuals (e.g. 760 exponential, fixed, constant). Model selection by BIC comparison was used to identify the 761 best random effects, variance weighting, and fixed effects structures (in that order).

# 762 2.2.5 Morphological trait distributions

- 763 We first explored inter- and intra-specific trait variation, to provide context for later trait
- 764 analyses. This included nonparametric Kruskal-Wallis analysis of variance (due to non-
- normality of residuals), and pairwise comparisons of trait distributions from the post-hoc

Dunn's test with Bonferroni correction. We also plotted trait-body size relationships todescribe ontogenetic trends across species.

768 To identify relationships between morphological traits and feeding guild membership, the 769 distribution of trait values (gape area, caudal fin AR, and pectoral fin AR) across individuals 770 within each feeding guild were again compared using Kruskal-Wallis analysis of variance and 771 Dunn's test. We also used a principal components analysis (PCA) based on Euclidian distances for gape area and caudal and pectoral fin AR, to explore the distribution of feeding 772 773 guilds in multi-dimensional trait space. To minimise the influence of individual body size on 774 the ordination, we standardised each measurement to the TL of the individual using the 775 following equation:

$$Y_i^* = Y_i \left| \frac{TL_0}{TL_i} \right|^b$$

Where  $Y_i^*$  is the standardized predicted value of trait Y for individual *i*,  $Y_i$  is the measured 777 value of the trait for individual *i*,  $TL_i$  is the measured TL of individual *i*,  $TL_0$  is the mean TL778 779 for all individuals and the parameter b is the slope from an ordinary least-squares (OLS) 780 regression of log-transformed Y and TL (Lleonart et al. 2000). This standardization effectively 781 adjusts the trait measurements to values they would have if the individuals were of the 782 average body size for the sampled population (Lleonart et al. 2000). We conducted this 783 standardisation for all individuals of the same species that were assigned to the same feeding 784 guild, to reduce allometric effects while still reflecting situations where different size classes 785 of a given species were assigned to separate guilds.

786 We then implemented a random forest (RF) model to assess whether feeding guild membership could be predicted from the standardised morphological traits. RF modelling is a 787 788 classification tool that uses bootstraps for the prediction of group membership and provides 789 an indication of the relative importance of predictor variables for partitioning individuals into 790 clusters (Cutler et al. 2007). We implemented a cross-validation approach by randomly sub-791 sampling 70% of the data to calibrate the model and then using the remaining 30% for 792 prediction. This was repeated 100 times to investigate the variability around classification 793 accuracy and relative importance of each trait. We assessed the predictive ability of the RF 794 model through the True Skill Statistic (TSS), with values of 1 and 0 indicating perfect and 795 completely random predictions, respectively (Allouche et al. 2006).

- 796 2.3 Results
- A total of 893 individuals were sampled for this study (Table 2.1). In most species, stomach
- contents were present for the majority of individuals, although for Scotia Sea icefish
- 799 (*Chaenocephalus aceratus*) only 25% of stomachs were non-empty (Table 2.1).
- 800 Table 2.1: Species sampled in this study, including Food and Agriculture Organization 3-
- 801 alpha taxonomic identification code for each species, number of individuals sampled, range
- 802 of total lengths (TL mm) of individuals sampled, and the number of non-empty stomachs.
- 803 The number of stomachs by size group is shown in Figure 2.2.

Species	Common name	Code	N fish	TL range (mm)	N stomachs
Family Channichthyidae (icefis	sh)				
Champsocephalus gunnari	Mackerel icefish	ANI	135	154-573	88
Chaenocephalus aceratus	Scotia Sea icefish	SSI	119	164-622	30
Pseudochaenichthys georgianus	South Georgia icefish	SGI	117	187-523	75
Family Nototheniidae (rockcod	)				
Notothenia rossii	Marbled rockcod	NOR	75	336-795	66
Trematomus hansoni	Striped rockcod	TRH	69	169-383	61
Lepidonotothen squamifrons	Grey rockcod	NOS	101	100-462	86
Lepidonotothen larseni	Painted notie	NOL	81	93-216	67
Gobionotothen gibberifrons	Humped rockcod	NOG	104	150-572	95
Family Bathydraconidae (drago	onfish)				
Parachaenichthys georgianus	South Georgia dragonfish	PGE	92	123-434	58

804 2.3.1 Species trait relationships

805 The sampled fish displayed various interspecific differences in their traits. The icefish *P*.

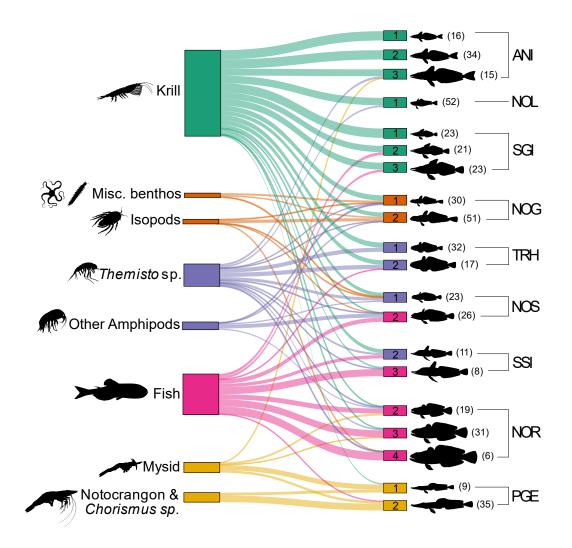
806 georgianus and C. aceratus and rockcod N. rossii had the largest gapes, while the remaining

807 rockcods, particularly L. larseni, had the smallest gapes (Figure A1a). The intercepts of the

- 808 species-specific gape-size relationships showed a similar rank order (with the icefish *P*.
- 809 georgianus highest and G. gibberifrons and L. larseni lowest) with consistent slopes across
- 810 species (Figure A1b). C. gunnari and C. aceratus displayed the highest caudal ARs and N.
- 811 *rossii* and the icefish *P. georgianus* exhibited the highest pectoral *ARs*, while the dragonfish *P.*
- 812 georgianus had the lowest caudal and pectoral ARs (Figure A1c,e). The relationships between
- 813 fin *AR* and body size varied considerably between species (Figure A1d,f).

# 814 2.3.2 Feeding guilds

- 815 The cluster analysis identified five feeding guilds at a dissimilarity level of 50% (Figure 2.2;
- 816 see A2 for pairwise similarities for each guild): (i) 'krill feeders' included all size classes of
- 817 the icefish C. gunnari and P. georgianus, and the rockcod L. larseni (which was only
- 818 represented by the smallest size class); (ii) 'benthos feeders' consumed miscellaneous
- 819 benthos and isopods, and represented all size classes of G. gibberifrons; (iii) 'Themisto and
- krill feeders' contained all size classes of *T. hansoni*, in addition to the smallest *L*.
- squamifrons and size class 2 C. aceratus, though fish were also important in their diet; (iv)
- 822 'fish feeders' contained the larger *C. aceratus* and *L. squamifrons*, and all *N. rossii*; and (v)
- 823 'benthic shrimp feeders', represented by the dragonfish *P. georgianus*, which fed primarily on
- 824 mysids and the decapods *Notocrangon sp.* and *Chorismus sp.*
- 825
- 826
- 827
- 828

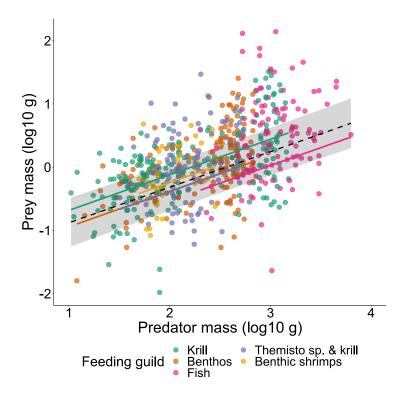


## 829

Figure 2.2: Sankey diagram depicting the trophic interactions between prey groups (left) and predators (right). Link thickness is proportional to the %IRI (links representing <1% are omitted for clarity). Node colours represent the feeding guilds determined by cluster analysis (green = 'krill feeders', orange = 'benthos feeders', purple = '*Themisto* and krill feeders', pink = 'fish feeders', yellow = 'benthic shrimp feeders'). Numbers within predator boxes indicate the size class (also represented by silhouette size), and numbers in brackets indicate sample size (number of non-empty stomachs). See Table 2.1 for a key to species codes.

The final selected linear mixed effects model of prey mass as a function of predator mass included a random intercept for prey type (reflecting different average body sizes for prey taxa), and a combination of fixed variance weighting structure for predator body mass and constant variance weighting structure for prey type (Table A3). The fixed effect structure included predator body mass and feeding guild as additive predictors, with no significant interaction identified (Table A4). Overall, the linear mixed effects model identified a significant increase in prey size with predator size, with a consistent slope but different

- intercepts of the relationship across feeding guilds ( $F_{6,599} = 134.80$ , p < 0.001; Figure 2.3;
- 845 Table A5).



846

Figure 2.3: Partial residuals plot from a linear mixed effects model of the relationship
between predator body mass and count-weighted average prey mass consumed across feeding
guilds. Each point represents one predator. Dashed line represents the overall model fit, with
shading representing 95% confidence intervals. Solid lines represent fits for each feeding

851 guild. Model coefficients are provided in Table A5.

# 852 2.3.3 Distinguishing feeding guilds with functional traits

Significant differences in trait values between feeding guilds were observed for all traits 853 854 (Figure 2.4; Table A6-A7). 'Fish feeders' had the largest gape areas, while 'krill feeders' and 855 'benthic shrimp feeders' generally had intermediate gape areas, with 'benthos feeders' and 'Themisto and krill feeders' having the smallest gape areas (Figure 2.4a). There were only 856 857 small differences in caudal fin AR across guilds, with the largest values observed in the 'fish 858 feeders' and 'krill feeders' and the smallest observed in the 'benthic shrimp feeders' (Figure 859 2.4b). Similarly, the 'fish feeders' had the highest pectoral fin ARs while the 'benthic shrimp 860 feeders' had significantly lower values compared to other groups (Figure 2.4c). These 861 differences between feeding guilds were captured in multi-dimensional space by the PCA of 862 length-standardised fish traits, which consisted of three dimensions with Dim1 and Dim2 863 together explaining 86% of the variance (Table A8). Gape area and pectoral fin AR were most

- strongly related to Dim1 (r = 0.62 and r = 0.59, respectively), while caudal fin *AR* was strongly correlated with Dim2 (r = 0.84; Figure 2.5a; Table A9). The PCA indicated substantial overlap in the trait space for each feeding guild, with the primary differentiation of the feeding guilds being between the 'fish feeders', which generally had positive Dim1 scores, and the 'benthic shrimp feeders', '*Themisto* and krill feeders', and 'benthos feeders' which generally had negative Dim1 scores and overlapped considerably with one another (Figure 2.5a). Additionally, 'benthic shrimp feeders' were separated from the '*Themisto* and
- krill feeders' guild along Dim2 (Figure 2.5a). The 'krill feeders' were the least differentiated
- by the traits, with individuals spread across most of the trait space (Figure 2.5a).
- 873 Despite the high levels of overlap in trait space for some feeding guilds, the Random Forest
- 874 model could predict feeding guild membership from the length-standardised traits relatively
- 875 well, with an average TSS score of  $0.77 \pm 0.12$  over 100 cross-validation runs. The most
- 876 important trait for predicting guild membership was gape area (84% relative importance),
- followed by caudal and pectoral fin AR (both 8% relative importance; Figure 2.5b).

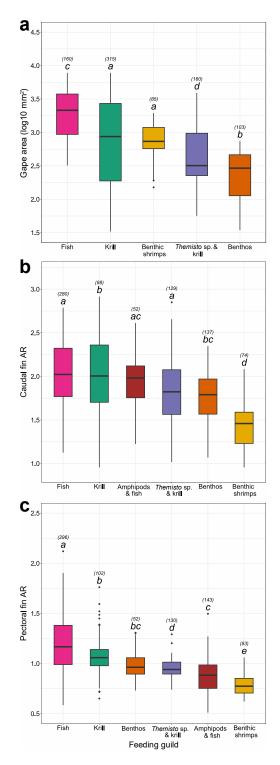


Figure 2.4: Boxplots displaying the distribution of absolute traits for each feeding guild: (a) gape area; (b) caudal fin aspect ratio (AR); (c) pectoral fin AR. Numbers in brackets represent sample sizes, letters indicate groupings assigned by a Dunn's test with Bonferroni correction (groups with a letter in common are not significantly different). Boxplots are organized in decreasing order of median trait value. Note the log scale in panel (a).

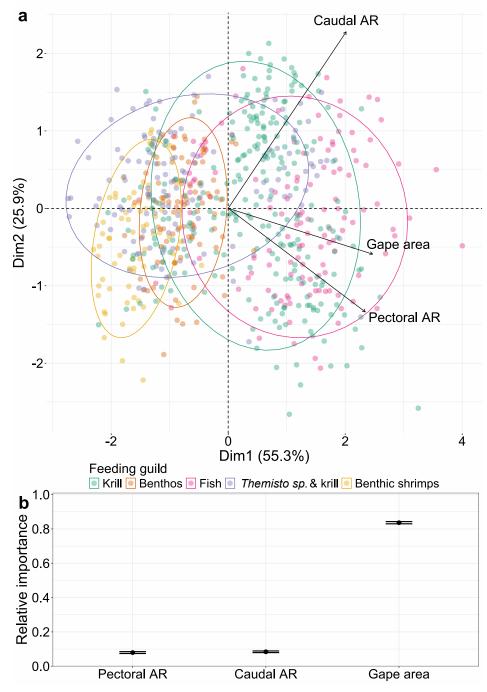




Figure 2.5: a) PCA plot of individual fish based on the length-standardised morphological
traits, coloured by feeding guild. Ellipses encompass 80% of the points from that guild; b)
relative importance of each length-standardised trait (as a proportion of the summed
importance of all traits) for classifying individuals into feeding guilds, as identified by the
Random Forest model. Error bars are the 95% confidence intervals from 100 crossvalidations of the model.

#### 891 2.4 Discussion

892 We investigated the role of morphological traits in driving prey selection at the community

893 level, providing insight into the partitioning of energy flows across species and size classes of

894 demersal fish. Such a trait-based understanding of trophic interactions can ultimately be used

- to model community structure and function (Kiørboe et al. 2018) and could thus elucidate
- 896 how future environmental change will alter the structure and stability of food webs.
- 897 Our analyses suggest that members of this community display differing levels of dietary 898 specialisation, with the diets of some groups dominated by specific taxa while others are 899 clearly more opportunistic and generalist. The feeding relationships we observed are 900 supported by previous dietary research in the region (e.g. McKenna 1991; Reid et al. 2007; 901 Targett 1981; Clarke et al. 2008; Main et al. 2009; Hollyman et al. 2021), indicating that we 902 successfully described the broad summer dietary niches of the studied fish. There can, 903 however, be interannual variability in diets (e.g. Main et al. 2009, Hollyman et al. 2021), 904 possibly driven by changes in krill availability, and it is notable that the %IRI of krill in C. 905 gunnari diets in 2023 (the collection year for this study) was the third highest in 14 years of 906 data (see Figure A2). Thus, our data may represent a situation in which krill were more 907 readily available to the demersal fish community than usual. Overall, the utilisation of krill by 908 all feeding guilds highlights the key role this group plays in maintaining energy flow within 909 Southern Ocean food webs. Demersal fish are themselves a major dietary component of 910 albatrosses, petrels, gentoo penguins, and Antarctic fur seals (Hill et al. 2005; Reid et al. 2005; Waluda et al. 2017), indicating that these fish are a key link between krill and many top 911 912 predators in the Southern Ocean.

#### 913 2.4.1 Size-based feeding

914 Average prey mass increased with predator mass, with a consistent scaling across all feeding 915 guilds regardless of prey type, suggesting strong size-structuring. Previous research has 916 shown that predator-prey size scaling relationships vary with diet type, as piscivores exhibit 917 positive allometric relationships but benthic invertivores have no significant change in prev 918 size with predator size (Dunic and Baum 2017). In contrast, our results support the 919 generalisability of predator-prey size relationships across feeding guilds, suggesting that such 920 allometric scaling could be applied more broadly to predict feeding interactions. Further work 921 will be required to determine whether the contrast with Dunic and Baum (2017) is down to 922 any differences in the available prey field and/or the behaviour of the focal fish species. The

923 variation in the intercepts of the relationship between predator size and prey size for different 924 feeding guilds may reflect differences in the levels of dietary specialisation of their 925 constituent members. Surprisingly, the 'fish feeders' had the smallest intercept, which could 926 be due to their generalist diets consisting of a broad range of prey sizes including numerous 927 very small prey items in addition to fewer large fish prey. As fish grow, their minimum prey 928 sizes often increase less steeply than their maximum prey sizes, resulting in a broadening of 929 their trophic niche, which may be the case for members of this guild (Scharf et al. 2000). In 930 contrast, the apparent high dependence of the 'krill feeders' on such a relatively large-bodied 931 prey, with minor contributions from other prey groups, may lead to a low trophic niche 932 breadth which ultimately drives the higher intercept for this feeding guild. There were also 933 some ontogenetic changes in prey selection, e.g. C. aceratus switched from a mixture of krill, 934 Themisto sp., and limited fish consumption to a fish-dominated diet as they became larger, 935 while G. gibberifrons moved from small and relatively immobile taxa like bivalves, 936 polychaetes, annelids, and gastropods to more mobile, large isopods as they grew. These 937 shifts indicate that these fish are potentially gape limited at smaller sizes or that their foraging 938 behaviour changes as they grow.

#### 939 2.4.2 Functional traits and feeding guilds

940 We found that some easily measured morphological traits can be used to distinguish feeding 941 guilds. Gape area was the best predictor of guild membership, and 'fish feeders' generally 942 had the largest gapes, reflecting the influence of gape limitation on the diets of fish. One krill-943 feeding species, the icefish P. georgianus, had absolute and standardised gape areas of similar 944 or even greater dimensions to those of 'fish feeders', which suggests that prey selection by 945 this species is not driven solely by gape limitation. Thus, the combination of multiple traits is 946 important in determining trophic niches in ecological communities. It has been proposed that 947 the elongated head, non-protractile jaw, and large gape of channichthyids including P. 948 georgianus facilitates a ram feeding mode (Bansode et al. 2014), and this might aid 949 zooplanktivores that feed on swarming prey as they can efficiently capture many prey items 950 simultaneously. The diets of larger P. georgianus also contained some fish and both P. 951 georgianus and C. aceratus ('fish feeder') are morphologically very similar, which indicates 952 that there may be further factors driving prey selection in these species. At the other end of 953 the scale, the 'benthos feeders' had the smallest absolute and relative gape areas of all the 954 feeding guilds. Possession of a relatively small mouth aperture correlates inversely with flow 955 velocity (Wainwright et al. 2007) and may benefit these fish which likely use suction feeding

to capture benthic epi- and infauna. Interestingly, *L. larseni* had an extremely small gape area despite being a member of the 'krill feeders' guild, which normally utilise large mouths to consume many prey items simultaneously. This suggests that *L. larseni* may target individual krill despite their sub-optimal trait configuration, highlighting the adaptability of the demersal fish community to incorporate such ubiquitous, high energy content prey in their diet.

961 The fin ARs measured across this community are quite low for fish in general (Sambilay 962 1990). This reflects the demersal nature of these fish, as low AR typically corresponds with 963 lower swimming efficiency but higher manoeuvrability at low speeds, suited to fish that 964 inhabit benthic environments (Bridge et al. 2016; Pauly 1989). Despite the narrow range of 965 AR values, it was possible to distinguish some species and feeding guilds based on this trait. 966 For example, C. gunnari are known to feed pelagically, which may explain their relatively 967 high caudal AR as this facilitates sustained swimming (Higham 2007). Similarly, the high 968 pectoral fin AR of the 'fish feeders' likely aids in capturing mobile prey, providing greater 969 potential for efficient, lift-based swimming (Pauly 1989; Bridge et al. 2016). The extremely 970 low fin AR observed for the 'benthic shrimp feeders' may be closely tied to the ecology of 971 their main prey (mysids and the decapods Notocrangon spp. and Chorismus spp.), which 972 spend much of their time either partially buried in substrate or perched on sponges (Gutt et al. 973 2004). Low pectoral fin AR, representing greater manoeuvrability and stability at low speeds 974 (Higham 2007), may provide this group with the mobility required to position themselves 975 rapidly and accurately in relation to these individual prey items. Additionally, malacostracan 976 crustaceans including shrimps are capable of rapid 'tail-flip' antipredator escape responses 977 (Arnott et al. 1998), therefore the high acceleration potential provided by very low caudal AR978 may allow the 'benthic shrimp feeders' to strike and capture their prey before they are able to 979 flee. The remaining guilds are more difficult to distinguish by their fin morphology alone, 980 suggesting either that similar swimming capabilities are required for feeding on krill, 981 amphipods, and benthic taxa, or that their fin morphology is not tied strongly to their diet.

#### 982 2.4.3 Further considerations

While our simple morphological traits proved useful for differentiating some feeding guilds,
we also conclude that there is a significant region of shared trait space between certain guilds.
In particular, the 'krill feeders' guild displayed a broad range of morphologies which
overlapped with all other guilds, suggesting that krill were readily available to fish regardless
of their morphology and behaviour. *Euphausia superba* is traditionally considered a pelagic
species which spends most of its time in epipelagic waters, but there is evidence that krill-

989 benthos interactions are common, with large krill swarms often observed close to the seabed 990 and krill found in the diet of strictly benthic species like the benthic skate Amblyraja 991 georgiana (Schmidt et al. 2011; Main and Collins 2011). Plasticity in krill behaviour may 992 mean they act as both a swarming prey in the water column for bentho-pelagic predators to 993 feed on and also come into contact with the epibenthos where they become available to 994 benthic feeders. The combination of such widespread accessibility and the high energetic 995 value and general abundance of krill makes them a suitable prey item for fish displaying a 996 wide variety of trait configurations. This further highlights the key role of krill within 997 Southern Ocean food webs, indicating that they effectively bridge the ecological niches 998 otherwise imposed by longer-term morphological evolution.

999 The density and availability of krill to shelf predators around South Georgia varies 1000 interannually (Fielding et al. 2014) and, as noted above, availability may have been high 1001 during sampling for the current study. Competition theory holds that niche partitioning should 1002 increase as resources become limited, with predators focusing on the prev they are best suited 1003 to exploit, thereby promoting coexistence (Schoener 1982). The link between morphology 1004 and diet might therefore become clearer in periods of krill scarcity when levels of dietary 1005 segregation within the groundfish community may increase as species match their longer-1006 term evolutionary niches. Continued monitoring of diets across the whole demersal 1007 community, including over different seasons, would provide insight into such competitive 1008 dynamics and could reveal temporal shifts in the importance of different prey taxa. For 1009 example, amphipods such as T. gaudichaudii are widely consumed by Southern Ocean fish, 1010 squid, seabirds, and marine mammals (Padovani et al. 2012; Havermans et al. 2019), and our 1011 results highlight their role in supplementing the diets of many demersal species around South 1012 Georgia. These amphipod taxa might therefore provide an alternative resource for demersal 1013 fish around South Georgia during periods of low krill availability, although the extent to 1014 which they could support the total energy requirements of the groundfish community requires 1015 further study (Kock et al. 1994).

Further studies on the links between morphological traits and diet will help elucidate the evolutionary constraints on prey selection. The traits used in this study represent broad and easily measurable morphological features expected to influence feeding, but there are likely to be further fine-scale morphological features that could be investigated in future studies.
For example, jaw length is linked to stealth and jaw closing speed and may therefore influence prey selection (Ferry et al. 2015), mouth position relates to feeding mode and

1022 habitat association (Helfman et al. 2023), and gill raker morphology determines feeding mode 1023 and minimum prey size (Macnuson and Heitz 1971). Ultimately, predator-prey interactions 1024 are determined by the combination of traits exhibited by both predator and prey individuals, 1025 including mobility, body size, physical and chemical defences, camouflage, visual acuity, 1026 feeding method, and habitat association (Spitz et al. 2014; Weigel and Bonsdorff 2018). It 1027 will therefore be important to consider the traits of prey alongside those of their predators 1028 when further investigating the drivers of feeding interactions. Detailed predator-prey trait 1029 matching could also facilitate analyses of the drivers of predation at the individual level by 1030 capturing the fine-scale variation in trait space across predator diets. By describing the 1031 distribution of traits across the available prey assemblage it is also possible to investigate how 1032 environmental change alters the suitability of the prey field for different predators (Weigel 1033 and Bonsdorff 2018), which will be a powerful tool for predicting the ecological 1034 consequences of climate change.

#### 1035 2.4.4 Conclusion

1036 Ongoing ecological changes, including shifting distributions of key prey like Antarctic krill,

1037 may result in the re-organisation of marine communities. This study provides a baseline

1038 understanding of how morphological traits underlie the ecology of Southern Ocean demersal

1039 fish. Continued investigation of the links between functional traits and prey selection will aid

1040 the production of generalisable community models to answer questions regarding trophic

1041 dynamics in marine food webs and the implications of abiotic change.

# 1042 3 Trophic structuring of modularity alters energy flow 1043 through marine food webs

1044 Published in *Frontiers in Marine Science* (https://doi.org/10.3389/fmars.2022.1046150)

1045 Abstract

1046 Food web interactions govern how ecosystems respond to climate change and biodiversity 1047 loss. Modularity, where subgroups of species interact more often with each other than with 1048 species outside their subgroup, is a key structural feature which has been linked to food web 1049 stability. We sought to address the lack of understanding of how modularity varies among 1050 ecosystems by comparing the structure of four highly resolved marine food webs and the 1051 importance of functional traits for predicting module membership. Modules in two offshore 1052 networks were partitioned largely by trophic level, creating an interdependence among them, 1053 whereas modules in two semi-enclosed bays were generally separated into energy channels 1054 with less trophic separation and containing distinct basal resources, providing greater 1055 redundancy in the flow of energy through the network. Foraging habitat and mobility 1056 predicted module membership in all networks, whilst body mass and foraging strategy also 1057 differentiated modules in the offshore and bay ecosystems, respectively. Environmental 1058 heterogeneity may be a key factor driving the differences in modularity and the relative 1059 importance of functional traits for predicting module membership. Our results indicate that, 1060 in addition to overall network modularity, the trophic structure of modules within food webs 1061 should be considered when making inferences about ecosystem stability.

#### 1062 3.1 Introduction

1063 The current global rate of species extinctions is unprecedented (Ceballos et al. 2015), and 1064 there is concern that biodiversity loss will reduce ecosystem functioning and services 1065 (Schmid et al. 2009; Tilman et al. 2014). Species interaction networks are key to 1066 understanding the ecosystem-level consequences of biodiversity loss, with certain network 1067 structures helping to limit the spread of perturbations through the ecosystem (Bruder et al. 1068 2019; Clark et al. 2020). Food webs provide tractable representations of species interactions 1069 and thereby allow us to compare the key structural features of communities that may confer 1070 stability (Rooney and McCann 2012; Ives and Carpenter 2007). One such stabilising feature 1071 is modularity, which is the presence of subgroups (modules) of species that interact often or 1072 strongly with one-another but have few or weak connections to species outside their subgroup 1073 (Krause et al. 2003). Modularity is believed to enhance food web stability by restricting the 1074 propagation of extinctions after a perturbation, thus buffering the wider network against 1075 disruption (Thébault and Fontaine 2010; Stouffer and Bascompte 2011). While common 1076 network-level properties, such as connectance or mean trophic level, are scale-dependent 1077 (Wood et al. 2015; Galiana et al. 2021), modularity is uncorrelated with species richness 1078 (Rivera-Hutinel et al. 2012; Montoya et al. 2015), facilitating structural comparisons across 1079 networks. Studies of modularity to date have generally quantified modularity in single food 1080 webs and with a variety of underlying methods, which precludes direct comparison of results 1081 across ecosystems (e.g., Rezende et al. 2009; D'Alelio et al. 2019). Assessing modularity in 1082 networks with different species assemblages would help to identify generalisable patterns in 1083 the distribution of modules, providing insight into the underlying drivers of stability.

1084 Physical and environmental variables play a key role in determining food web structure, with 1085 habitat heterogeneity shown to increase network complexity and niche availability (Tews et 1086 al. 2004; Kortsch et al. 2019). The diversity of ecological niches and refuges present in 1087 intertidal and coastal regions may therefore increase modularity compared with more uniform 1088 offshore areas. Differences in environmental factors such as temperature and depth may also 1089 drive structural contrasts between ecosystems (Gibert 2019; López-López et al. 2021). For 1090 example, the historically stable temperatures of Antarctic waters and their relative biotic 1091 isolation from other oceans (Murphy et al. 2007; Morley et al. 2020), might lead to less 1092 modular networks compared to lower latitudes.

1093 Functional traits provide a framework for describing community structure, as the match 1094 between consumer and resource traits determines the distribution of feeding interactions 1095 (Bartomeus et al. 2016). A key trait underlying trophic interactions in marine systems is body 1096 size, and the relative size of consumers to their resources has been recognised as a potentially 1097 key determinant of species organisation into modules (Rezende et al. 2009; Gravel et al. 1098 2013). The consumer-resource body mass ratio generally declines with increasing consumer 1099 size due to the higher energy demands of larger organisms, which leads to a greater reliance 1100 on proportionally larger prey (Arim et al. 2007). As larger organisms usually occur higher in 1101 the food web, the result is a negative relationship between consumer trophic level and 1102 consumer-resource body mass ratio: a macroecological pattern which is consistently found in 1103 different food webs (Jonsson et al. 2005; Tucker and Rogers 2014). This indicates that body 1104 size could also determine the distribution of modules across trophic levels. Previous research 1105 has suggested that the level of diet contiguity in the food web may determine modularity,

1106 with modules in some networks displaying trophic clustering such that they encompass a 1107 relatively limited range of trophic levels and have low overlap of trophic levels between 1108 modules (Guimera et al. 2010; Kortsch et al. 2015). Other traits may also play an important 1109 role, with foraging habitat determining the spatial distribution of species and thus their 1110 likelihood of interacting (Rezende et al. 2009; Kortsch et al. 2019). Mobility and feeding 1111 mode also contribute to the trophic role of species within networks, by determining their 1112 activity levels and the types of resources they consume (Lazzaro et al. 2009; Gilabert et al. 1113 2019). In fact, it has been proposed that modules in some networks represent semi-isolated 1114 energy channels, whereby energy flows from a distinct set of basal resources to an 1115 assemblage of higher consumers with a particular set of functional traits (Gauzens et al. 2015; 1116 Rodriguez et al. 2022). Clearly, despite the consensus that modularity acts to stabilise food 1117 webs, there are contrasting viewpoints on what the key determinants of modularity are and 1118 how modules are distributed within communities.

In this study, we compared the modular structure of the four most highly resolved marine food webs currently available. We quantified how differences in their spatial distribution and constituent taxonomic groups translate into the organisation of modules and the relative importance of functional traits for predicting module membership. Our primary research objectives were 1) to determine whether there are differences in the organisation of modules between networks; 2) to identify which functional traits can be used to predict the species that are included in each module.

#### 1126 *3.2 Materials and methods*

#### 1127 *3.2.1 Study systems*

1128 Based on a review of marine food webs in the GATEWAy (Brose et al. 2019) and ECOWeB 1129 (Cohen 2010) databases and the wider literature, we identified four systems in which the 1130 overwhelming majority of nodes were highly resolved to genus or species level (excluding a 1131 handful of cryptic taxa and basal groups such as sediment and detritus). Aggregation of taxa 1132 in the other networks could mask potential modules and introduce methodological biases 1133 (Krause et al. 2003), so they were not considered here. The four chosen food webs represent a 1134 range of locations from the high Antarctic (Weddell Sea), and (sub)Antarctic (Scotia Sea), to 1135 temperate (Lough Hyne) and Arctic (Kongsfjorden). The networks differ in their size 1136 (number of nodes and links), spatial extent (including depth), and functional groups (Table 1137 3.1). The Scotia Sea food web was obtained from the British Antarctic Survey's UK Polar

- 1138 Data Centre (López-López et al. 2021), while the remaining webs were extracted from the
- 1139 GATEWAy database (Brose et al. 2019). Each of these networks was compiled through a
- 1140 combination of direct observation and diet analysis of organisms within the focal ecosystem,
- and wider literature research to characterise the diet of organisms in other regions or for
- 1142 closely related taxa. Following Grilli et al. (2016), we removed cannibalistic links in order to
- 1143 focus on interspecific interactions.
- 1144 Table 3.1: Characteristics of the four study systems. SS = Scotia Sea, WS = Weddell Sea, LH
- 1145 = Lough Hyne, KO = Kongsfjorden.

Name	Nodes	Links	Approximate latitude (°N)	Approximate extent (km <sup>2</sup> )	Min to max bottom depth (m)	Ecosystem type	Constituent functional groups
SS	228	10,827	-57.0	1.5×10 <sup>6</sup>	>1,000 to >3,000	Offshore	Pelagic (excluding birds and mammals), benthos excluded
WS	490	15,987	-76.0	2×10 <sup>5</sup>	200 to 500	Offshore	Pelagic (including birds and mammals), benthos
LH	340	5,012	51.5	0.5	0 to 50	Coastal bay	Pelagic (including birds and mammals), benthos, intertidal
КО	260	1,590	79.0	209	0 to 400	Coastal bay	Pelagic (including birds and mammals), benthos, intertidal

1146

1147 *3.2.2 Module identification* 

1148 The modularity of each food web was calculated with a Simulated Annealing algorithm using

1149 the 'netcarto' function in the R package 'rnetcarto' (Doulcier and Stouffer 2015). This

- algorithm uses a probabilistic procedure whereby nodes are initially partitioned into arbitrary
- 1151 modules and then iteratively moved into different modules until the maximum modularity is
- 1152 obtained (Guimerà and Nunes Amaral 2005; Chen et al. 2014). Modularity ranges from -1 to
- 1153 +1, with negative and positive values indicating a less and more modular structure than
- 1154 expected at random, respectively (Newman and Girvan 2004; Newman 2006). One hundred
- simulations were conducted per network to assess the variability in outputs resulting from the
- 1156 stochastic component of the algorithm.

### 1157 *3.2.3 Functional traits*

- 1158 Eight functional traits were selected to investigate the partitioning of species into modules
- 1159 (Table 3.2). These traits were chosen because they could be easily identified and generalised
- 1160 across all species in the four ecosystems. For each species, body mass estimates were derived
- 1161 from the original food web studies, and the remaining trait values were assigned based on
- 1162 data obtained via literature review, assessment of images, and diet compositions.
- 1163 Table 3.2: Functional traits identified for each species in the four food webs. See Appendix
- 1164 B1: Supplementary methods and results for more details.

Trait	Description			
Body mass	Species averages from field measurements and literature, log <sub>10</sub> transformed			
Foraging habitat	Physical space in which organisms forage. Categories vary due to environmental differences between food webs. Scotia Sea: epipelagic, mesopelagic, bathypelagic. Weddell Sea: epipelagic, meso/benthopelagic, benthic. Lough Hyne and Kongsfjorden: pelagic, benthic, intertidal.			
Mobility	A scale of increasing mobility: sessile; passive drifter; crawler; use of swimming appendages; jet propulsion; lift-based swimming.			
Prey-capture strategy	A scale based on how actively the species captures prey: primary producer; passive capture; ambush predator; active suspension/detritus feeder; active searcher/hunter			
Prey-capture appendages	Binary, presence or absence of external appendages which could be reasonably considered to play a role in prey grasping and manipulation.			
Body robustness	A scale of body type, from fragile to robust: gelatinous; soft-tissue with no internal skeleton; soft-tissue with internal skeleton; external carapace; external hard shell			
Spines	Binary, presence or absence of defensive spines			
Translucency	Binary, used to distinguish species that are clearly see-through (e.g. most gelatinous zooplankton, some amphipods) from those which are not (e.g. crabs, fish)			

#### 1165 *3.2.4 Statistical analysis*

All analyses were conducted in R 4.1.0 (R Core Team, 2021). Differences in the modular structure of each network were investigated by comparing the distribution of node-level metrics (prey averaged trophic level, body mass, generality, vulnerability, and omnivory) across modules. The data did not conform to normality and homogeneity of residuals, and so non-parametric Kruskal-Wallis tests were performed followed by post-hoc Dunn tests with a Bonferroni correction.

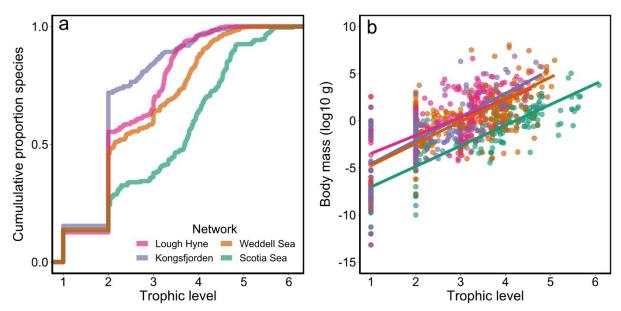
1172 We used Random Forest (RF) models to investigate the relative roles of the functional traits 1173 in explaining the modular structure of each network. The RF model is a machine learning 1174 classification tool which uses bootstraps of the data to predict observations and provide a 1175 measure of the relative importance of predictor variables (Cutler et al. 2007). A benefit of RF 1176 models is that they make no prior assumptions about the distribution of response or predictor 1177 variables and can handle datasets containing multiple data types (Cutler et al. 2007). We 1178 implemented the conditional RF algorithm using the 'cforest' function in the R package 1179 'party' (Hothorn et al. 2005), which relies on a conditional inference framework and is 1180 unbiased in cases where predictors have a highly variable number of categories or are 1181 correlated (Strobl et al. 2007; Strobl et al. 2008). For each food web, we implemented a 1182 cross-validation approach by randomly sub-sampling 70% of the data for model calibration 1183 and then making predictions from the remaining 30%. This process was repeated 20 times to 1184 give an indication of the variability in the classification accuracy and relative importance of 1185 each functional trait. The predictive ability of the models was assessed using the average True Skill Statistic (TSS), which represents the proportion of successful predictions versus false 1186 1187 predictions, with values of 0 and 1 indicating completely random and perfect predictions, 1188 respectively (Allouche et al. 2006).

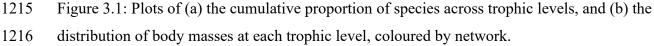
1189 To further investigate the role of body size (specifically, whether size-structured feeding is 1190 related to modularity), we used analysis of covariance (ANCOVA) to test the relationship 1191 between consumer-resource body mass ratio and consumer trophic level, while distinguishing 1192 between trophic links occurring within or between different modules. The average consumer-1193 resource body mass ratio of each consumer species was used as the dependent variable to avoid 1194 any confounding effects resulting from the fact that some consumers had many resources while 1195 others had very few. The main and interactive effects of consumer trophic level and link 1196 position (within or between modules) were the explanatory variables. Weighted Generalised 1197 Least Squares models were used, with an exponential variance structure by trophic level to1198 account for heterogeneity in the model residuals.

#### 1199 3.3 Results

1214

1200 The Scotia Sea network represents a pelagic system, dominated by phytoplankton, gelatinous 1201 zooplankton, crustaceans, and fish, while benthic taxa are excluded due to the lack of 1202 available information on benthic communities. The Weddell Sea has a similar functional 1203 group composition, though it also includes many benthic species including sessile sponges, 1204 mobile detritivores, and demersal fish, in addition to incorporating mammals and seabirds. 1205 Both Lough Hyne and Kongsfjorden include a variety of benthic, intertidal and pelagic 1206 species, including macroalgae, sponges, crustaceans, and fish, in addition to seabirds and 1207 mammals. The Weddell Sea, Lough Hyne and Kongsfjorden food webs have similar 1208 maximum trophic levels (5.1, 4.6 and 4.8, respectively), while the Scotia Sea has a maximum 1209 trophic level of 6.1. The trophic distribution of species is similar in the Weddell Sea, Lough Hyne and Kongsfjorden, with most species found between trophic levels 2 and 4, while in the 1210 1211 Scotia Sea, species are distributed quite evenly between trophic levels 2 and 5 (Figure 3.1a). 1212 The trophic distribution of body masses is similar in all networks, with larger organisms 1213 found at higher trophic levels (Figure 3.1b).





#### 1217 *3.3.1* Module identification

1218 Modularity was significantly different between all four food webs ( $X^{2}_{(8)} = 374.80, p < 0.001$ ; 1219 Dunn's test: p < 0.001). Three modules were identified in the Scotia Sea and Weddell Sea

1220 networks (Modularity =  $0.157 \pm 0.007$  and  $0.319 \pm 0.002$ , respectively), while five were found

in Lough Hyne and Kongsfjorden (Modularity =  $0.404 \pm 0.009$  and  $0.496 \pm 0.005$ , respectively)

- 1222 (Figure 3.2). Energy flow between modules in the Scotia Sea and Weddell Sea was generally
- 1223 one-sided, with the majority of links between any pair of modules flowing in the same
- 1224 direction, whereas flows were more two-sided in Lough Hyne and Kongsfjorden (Figure 3.2).
- 1225 These results were deemed to be representative of the 100 Simulated Annealing runs, with at
- 1226 least 95% of within-module interactions found to be consistent across runs in each network
- 1227 (see Appendix B1: Supplementary methods and results for details of the Simulated Annealing
- 1228 result selection process and robustness).

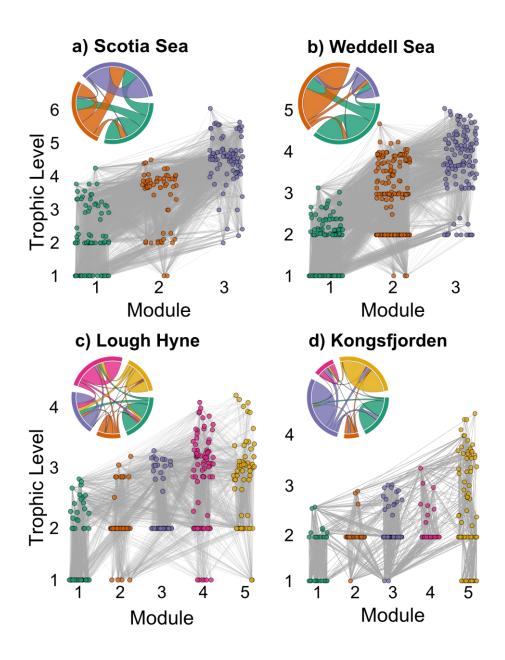
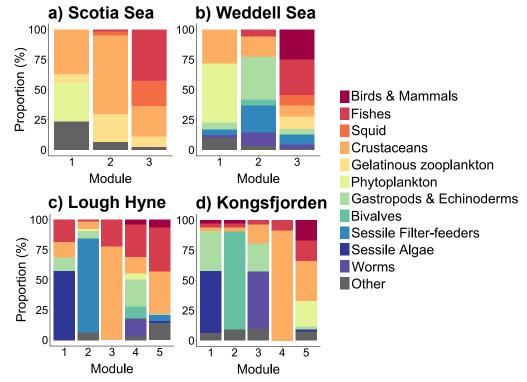




Figure 3.2: Modular structure of the four studied food webs: (a) Scotia Sea; (b) Weddell Sea; (c) Lough Hyne; (d) Kongsfjorden. Networks are plotted with nodes coloured and positioned along the x- and y-axes according to module and trophic level, respectively. Inset chord diagrams display the distribution of interactions within and between modules, with chord thickness proportional to the number of links and colour indicating the source module of the interactions.

Some of the networks displayed similarities in the distribution of species across modules. In both the Scotia Sea and Weddell Sea there was a basal module that contained most (>90%) of the basal resources, being made up largely of epipelagic phytoplankton and crustaceans, and a top predator module that was made up primarily of fish and squid (in addition to marine mammals and seabirds in the Weddell Sea) (Figure 3.3a-b). The main difference between these 1241 networks was the remaining module, which was composed primarily of gelatinous organisms 1242 and crustaceans in the Scotia Sea and benthic taxa such as echinoderms, sponges, and 1243 bryozoans in the Weddell Sea (Figure 3.3a-b). In both Lough Hyne and Kongsfjorden, basal 1244 resources were present in four out of five modules. In both networks there was a macrophyte 1245 module consisting largely of seaweeds and sessile algae and a module containing many benthic and intertidal amphipods, while fishes were distributed across all modules (Figure 3.3c-d). Both 1246 1247 food webs contained a benthic consumer module consisting mainly of gastropods, crustaceans, and worms, and a sessile filter-feeding module composed mostly of sponges and bryozoans in 1248 1249 Lough Hyne and of bivalves and barnacles in Kongsfjorden (Figure 3.3c-d). The final module 1250 in both networks was largely made up of bentho-pelagic organisms (Figure 3.3c-d).

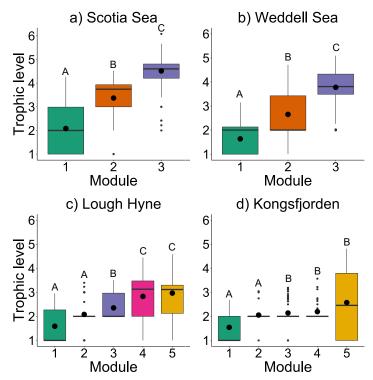


1251

Figure 3.3: The relative proportion of different taxonomic groups within each module in the four food webs: (a) Scotia Sea; (b) Weddell Sea; (c) Lough Hyne; (d) Kongsfjorden. Species were initially grouped by taxonomy and then groups with few individuals were either combined (if they had similar ecology) or were assigned to the group "Other".

#### 1256 3.3.2 Module topology

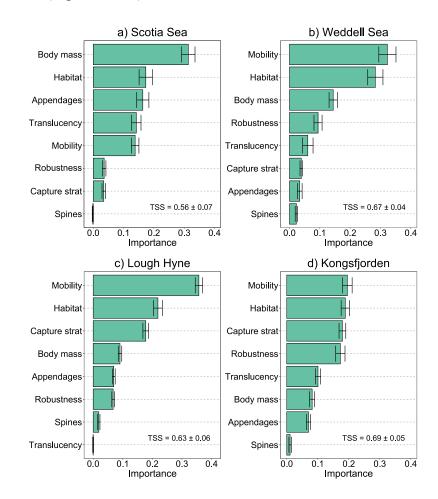
1257 There was a significant difference in trophic level between modules in each of the food webs 1258 (Scotia Sea:  $X^2_{(2)} = 147.16$ , p < 0.001; Weddell Sea:  $X^2_{(2)} = 192.56$ , p < 0.001; Lough Hyne, 1259  $X^2_{(4)} = 86.643$ , p < 0.001; Kongsfjorden,  $X^2_{(4)} = 16.57$ , p = 0.002; Figure 3.4). In the Scotia Sea 1260 and Weddell Sea, trophic level was significantly different among all three modules (Dunn's 1261 test: p < 0.001), while in Lough Hyne and Kongsfjorden significant differences were only found 1262 between certain module pairs. The mean difference in trophic level between all possible species 1263 pairs which belong to separate modules was greater in the Scotia Sea (1.62, SE = 0.009) and Weddell Sea (1.43, SE = 0.004) than in Lough Hyne (0.70, SE = 0.004) and Kongsfjorden 1264 1265 (0.44, SE = 0.005), and this pattern was retained after accounting for the effects of the differing number of modules and maximum trophic level in each network (Appendix B1). This 1266 1267 highlights the greater trophic clustering of modules that is present in the Scotia Sea and Weddell Sea compared with Lough Hyne and Kongsfjorden. A comparison of the distribution of trophic 1268 1269 levels for each module across all Simulated Annealing runs suggested that these results are 1270 robust to changes in the number of modules or distribution of nodes between modules 1271 (Appendix B1). Results for the other node-level metrics (i.e. generality, vulnerability, and 1272 omnivory) generally reflect the distribution of modules across trophic levels in these networks, 1273 with clear differences between modules for the Scotia Sea and Weddell Sea, but not for Lough 1274 Hyne and Kongsfjorden (Figure B1-B5).



1275 1276 Figure 3.4: Boxplots of prey-averaged trophic level across modules within each network: (a) 1277 Scotia Sea; (b) Weddell Sea; (c) Lough Hyne; (d) Kongsfjorden. Large black points indicate 1278 the mean, thick horizontal lines represent the median, boxes indicate the interquartile range, 1279 whiskers are  $1.5 \times$  the interquartile range, and outliers beyond this range are indicated as 1280 small black points. Boxes not sharing a common letter are significantly different from one 1281 another using a Dunn's test (p < 0.05).

#### 1282 3.3.3 Functional traits

The Random Forest models performed well at predicting module membership from the 1283 functional traits, as evidenced by their high TSS scores (>0.56; Figure 3.5). Body mass was a 1284 1285 key trait for predicting module membership in the Scotia Sea, accounting for ~30% of overall 1286 importance, followed by habitat, mobility, feeding appendages, and translucency, which all had similar values of importance (Figure 3.5a). In the Weddell Sea, mobility and habitat together 1287 accounted for almost 60% of importance, and body mass was also valuable (~13% of 1288 1289 importance, Figure 3.5b). In Lough Hyne and Kongsfjorden, mobility, capture strategy and 1290 habitat accounted for ~60% of the total importance, while body mass represented <10% of 1291 overall importance (Figure 3.5c-d).



#### 1292

1293 Figure 3.5: Relative importance of each functional trait (as a proportion of the summed

1294 importance of all traits) for classifying species into modules, as identified by Random Forest

1295 models for each network: (a) Scotia Sea; (b) Weddell Sea; (c) Lough Hyne; (d) Kongsfjorden.

- 1296 Error bars are the 95% confidence intervals resulting from 20 cross-validations of the
- 1297 importance of each trait. True Skill Statistics (TSS) values indicate the predictive
- 1298 performance of the model.

1299 The importance of body mass in the Scotia Sea and Weddell Sea RF models is reflected in the 1300 distribution of sizes across modules. There was a significant difference in body mass among all modules in the Scotia Sea ( $X^2_{(2)} = 113.75$ , p < 0.001; Figure 3.6a) and Weddell Sea ( $X^2_{(2)} = 113.75$ , p < 0.001; Figure 3.6a) 1301 191.09, p < 0.001; Figure 3.6b). While the distribution of body masses was also significantly 1302 1303 different between some modules in Lough Hyne and Kongsfjorden ( $X^{2}_{(4)} = 27.634$ , p < 0.001, and  $X^{2}_{(4)} = 32.414$ , p < 0.001, respectively), modules were not as obviously separated 1304 1305 according to body mass as in the other webs (Figure 3.6c-d), which also reflects the results of 1306 the RF models. A comparison of the distribution of body mass values in each module across 1307 all Simulated Annealing runs suggested that these results are robust to changes in the number 1308 of modules or distribution of nodes between modules (Appendix B1). See Figure B6-B12 for 1309 a description of the distribution of the remaining traits across modules in each network.

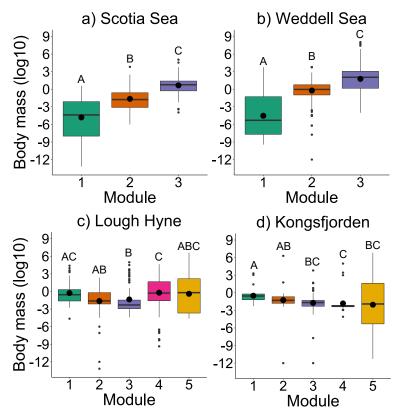


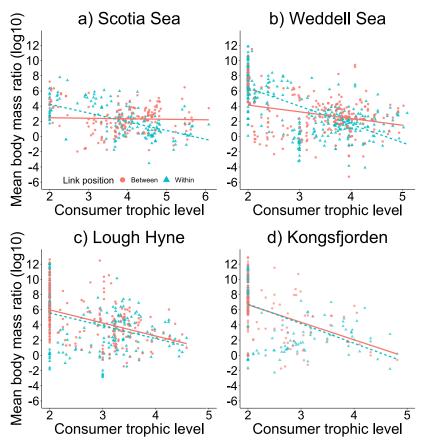
Figure 3.6: Boxplots of body mass across modules within each network: (a) Scotia Sea; (b) Weddell Sea; (c) Lough Hyne; (d) Kongsfjorden. Large black points indicate the mean, thick horizontal lines represent the median, boxes indicate the interquartile range, whiskers are 1.5 × the interquartile range, and outliers beyond this range are indicated as small black points. Boxes not sharing a common letter are significantly different from one another using a Dunn test (p < 0.05).

1318 among networks (Figure 3.7). There was a significant interaction between consumer trophic level and link position in the Scotia Sea ( $F_{1,338} = 29.76$ , p < 0.001) and Weddell Sea ( $F_{1,643} =$ 1319 44.53, p < 0.001). In both, consumer-resource body mass ratio declined more steeply with 1320 1321 consumer trophic level for interactions within modules than for interactions between modules (Scotia Sea: within, p < 0.001,  $r^2 = 0.35$ ; between: p = 0.807; Weddell Sea: within, p < 0.001, 1322  $r^2 = 0.50$ ; between, p < 0.001) (Figure 3.7a-b). In contrast, no significant interaction between 1323 1324 consumer trophic level and link position was observed in Lough Hyne ( $F_{1,447} = 0.01$ , p = 0.919) or Kongsfjorden ( $F_{1,326} = 0.39$ , p = 0.531). Instead, consumer-resource body mass ratio declined 1325 significantly with increasing consumer trophic level regardless of link position in both Lough 1326

There were some key differences in the distribution of consumer-resource body mass ratios

- 1327 Hyne ( $F_{1,449} = 106.80, p < 0.001, r^2 = 0.20$ ) and Kongsfjorden ( $F_{1,328} = 142.66, p < 0.001, r^2 = 0.20$ )
- 1328 0.27; Figure 3.7c-d).
- 1329

- 1330
- 1331



1332

1333 Figure 3.7: Average body mass ratio of every consumer to each of its resources, plotted against the trophic level of the consumer, for each network: (a) Scotia Sea; (b) Weddell Sea; 1334 1335 (c) Lough Hyne; (d) Kongsfjorden. Point shape and colour indicate whether the focal 1336 interaction occurred among species within the same module, or between modules. There was 1337 an interactive effect of consumer trophic level and link position on consumer-resource body 1338 mass ratio for the Scotia Sea (within: y = 6.62 - 1.17x, p < 0.001, r2 = 0.35; between: y = 2.481339 -0.04x, p = 0.807, r2 = 0.01) and Weddell Sea (within: y = 11.77 - 2.59x, p < 0.001, r2 = 1340 0.50; between: y = 5.27 - 0.70x, p < 0.001,  $r^2 = 0.10$ ). There was only a significant main 1341 effect of consumer trophic level on consumer-resource body mass ratio for Lough Hyne (y = 1342 9.22 - 1.71x, p< 0.001, r2 = 0. 20) and Kongsfjorden (y = 11.64 - 2.47x, p< 0.001, r2 = 1343 0.27).

#### 1344 3.4 Discussion

1345 This study provides insight into the patterns and drivers of modularity in marine food webs.

1346 We found two distinct ways in which modules were organised: (1) a strong differentiation by

- 1347 trophic level in the Weddell Sea and Scotia Sea, matching the trophic clustering of modularity
- described for other food webs (Guimera et al. 2010; Kortsch et al. 2015); and (2) multiple
- 1349 modules spanning from distinct basal resources to higher trophic levels in Lough Hyne and

1350 Kongsfjorden, resembling the description of modules as energy channels (Gauzens et al. 1351 2015; Zhao et al. 2017). Our results also confirm the importance of body mass and foraging 1352 habitat for determining modularity (Krause et al. 2003; Rezende et al. 2009), whilst 1353 highlighting the added importance of mobility and prey capture strategy. The strong size-1354 structuring of modules in the Weddell Sea and Scotia Sea leads to shallower trends in 1355 consumer-resource body mass ratios with consumer trophic level for interactions between 1356 modules than those within modules. This represents a disruption to the macroecological 1357 pattern of declining consumer-resource body mass ratios with consumer trophic level 1358 observed in many food webs (Jonsson 2014; Tucker and Rogers 2014), suggesting that 1359 feeding interactions between modules are occurring at sub-optimal size ratios. In contrast, 1360 Lough Hyne and Kongsfjorden display weak size-structuring and show a declining 1361 relationship between consumer-resource body mass ratios and consumer trophic level both 1362 for interactions occurring within and between modules. Our results suggest contrasting mechanisms underlying the structure of marine food webs in different regions, which may 1363 1364 affect their stability in the face of global change.

#### 1365 *3.4.1 Drivers of structural differences*

1366 Strong spatial and temporal variability in abiotic conditions, such as temperature, desiccation, 1367 and salinity, can drive differentiation of ecological niches and patterns of species zonation, 1368 particularly in intertidal ecosystems (Gingold et al. 2010; Kraan et al. 2013; Gallucci et al. 1369 2020). Intertidal and benthic community composition is also shaped by habitat heterogeneity, 1370 which determines the distribution of traits such as mobility and feeding mode (Pacheco et al. 2011; Buhl-Mortensen et al. 2012; Srinivas et al. 2020). In contrast, offshore ecosystems are 1371 1372 generally considered less complex with major structuring environmental gradients (e.g. light, 1373 temperature, pressure) changing predictably with depth (López-López et al. 2021), and may 1374 therefore display stronger size-structuring of trophic interactions. As the breadth of available 1375 niches increases with environmental, habitat, and resource heterogeneity, food webs may 1376 become more modular and separated into distinct energy channels. In highly heterogeneous 1377 environments, module membership may therefore be determined primarily by traits specific to the environmental niche, such as prey capture strategy and mobility, rather than by more 1378 general structuring factors such as body mass. Below, we explore this hypothesis in the 1379 1380 context of our focal food webs.

1381 The Scotia Sea network represents a pelagic oceanic ecosystem, and the Weddell Sea network 1382 represents a deep shelf system incorporating both pelagic and benthic shelf species. The 1383 offshore nature of these networks means that basal resources are limited largely to 1384 phytoplankton and detritus, with both networks excluding the pronounced heterogeneity of the 1385 intertidal zone. The Antarctic Circumpolar Current also provides these Southern Ocean 1386 ecosystems with relatively stable and predictable oceanographic conditions (Murphy et al. 1387 2007; Morley et al. 2020), which might help to drive the similarity in modular organisation. 1388 Both ecosystems experience a high degree of connectivity, with large-scale diurnal vertical 1389 migrations in the Scotia Sea and strong bentho-pelagic coupling in the Weddell Sea 1390 (Piatkowski et al. 1994; La Mesa et al. 2019; Pineda-Metz 2020). This may result in a stronger 1391 interdependence between modules in both ecosystems, as the deeper top-predator modules rely 1392 on the energy generated in the near-surface basal resource modules, linked via the diurnal 1393 migrators or bentho-pelagic couplers. This might reduce the influence of factors like habitat 1394 heterogeneity and prey capture strategy, with depth-based foraging habitat and size-based prey-1395 handling constraints becoming the primary factors structuring modularity in both networks. 1396 The additional importance of mobility in the Weddell Sea largely reflects the distinct 1397 locomotory methods used in the different modules, i.e., sessile or crawling organisms in the 1398 benthic module, drifting phytoplankton and primary consumers with appendages in the 1399 epipelagic basal module, and mobile swimmers in the top predator module.

1400

1401 In contrast, Lough Hyne and Kongsfjorden are semi-enclosed coastal ecosystems which 1402 encompass both the intertidal and subtidal zones and experience high environmental 1403 variability. Kongsfjorden is subject to significant seasonal inputs of terrestrial nutrients (Calleja 1404 et al. 2017; Retelletti Brogi et al. 2019), and experiences strong gradients in turbidity, 1405 temperature, and salinity due to glacial inputs and influxes from the West Spitsbergen Current 1406 (Hop et al. 2002; Calleja et al. 2017). As a result, there are significant differences in community 1407 composition and abundance at different locations within the fjord (Hop et al. 2002; Calleja et al. 2017). Lough Hyne experiences high terrestrial nutrient loads (Jessop et al. 2011), in 1408 1409 addition to significant pH gradients, high variability in water temperature, and seasonal 1410 hypoxia at depth (Bell 2002; Sullivan et al. 2014), which are also likely to drive spatial contrasts 1411 in community structure. The presence of the physically complex and variable intertidal zone, 1412 and the fluctuating environmental conditions may promote the differentiation of ecological 1413 niches. This could drive the diversity of energy channels centred around different types of basal 1414 resources, supporting species with a mix of foraging behaviours, mobilities, and habitat traits 1415 (Gauzens et al. 2015; Rodriguez et al. 2022).

#### 1417 *3.4.2 Implications for food web stability*

1418 Modules partially isolate sections of the food web from one another, and thereby reduce the 1419 propagation of perturbations and maintain the functioning of the wider network (Stouffer and 1420 Bascompte 2011). Previous studies have used overall network modularity to make inferences 1421 regarding their stability and functioning (e.g., Stouffer and Bascompte 2011; Grilli et al. 1422 2016; D'Alelio et al. 2019), but there has been little consideration of how the positioning of 1423 modules across trophic levels might alter stability. Our results suggest that, in networks with 1424 strong trophic clustering of modules, inter-module energy flows will be key to maintaining 1425 consumer populations and providing top-down regulation. Any perturbation affecting species 1426 in one module will have consequent effects on those in other modules, thus undermining the 1427 potential stabilising effect of modularity. The trophic clustering of modules also has 1428 implications for stabilising consumer-resource mass ratios, which are generally greatest near 1429 the base of the food web and decrease at higher trophic levels (Jonsson 2014; Tucker and 1430 Rogers 2014). This is because larger consumers need to consume larger prey to maximise 1431 energy intake and handling efficiency, such that optimal prey size gets closer to the size of the 1432 predator as its trophic level increases (Costa 2009). We found this pattern only exists for interactions within modules in networks that exhibit strong trophic (size) structuring of 1433 1434 modularity, while many interactions between modules may be allometrically sub-optimal. For 1435 example, a predator in a lower-level module may be too small to handle prey from a higher-1436 level module, while prey in lower-level modules may be too small to provide enough energy 1437 for predators in higher-level modules. This may reduce the redundancy of alternative 1438 pathways for energy flow in the food web by constraining consumers largely to within-1439 module prey choices. Furthermore, the allometric scaling of metabolism and consumption 1440 rates means that consumer-resource body mass ratios can determine the strength of trophic 1441 interactions (Emmerson and Raffaelli 2004; Vucic-Pestic et al. 2010). An environmental 1442 perturbation which results in prey loss from a given module may therefore have a strong 1443 destabilising effect as predators are forced to feed on sub-optimal prey sizes to compensate, 1444 thereby also disrupting the distribution of strong and weak interactions within the food web.

In contrast, food webs with modules that represent semi-isolated food chains may be more robust to perturbations, as species extinctions in a given module will not impact the supply of energy to species in other modules. This structure maintains the negative relationship between consumer-resource mass ratio and consumer trophic level, regardless of whether consumers interact with resources within or between modules, because such modules encompass species

1450 at a broad range of trophic levels and body masses. This means consumers are allometrically 1451 unconstrained in their ability to feed on species in different modules and may be able to adapt 1452 their feeding behaviour in response to perturbations within their own module. For example, 1453 while the loss of key basal resources from an individual module could have detrimental 1454 consequences for the specialised primary consumers in that module, higher predators may be 1455 able to maintain sufficient energy intake by feeding on species from other modules. This is 1456 analogous to fast and slow energy channels coupled by mobile predators, which promote 1457 stability by generating asynchronously fluctuating resources that dampen variation in consumer 1458 populations (Rooney et al. 2008; McCann and Rooney 2009). The stabilising effect of modular 1459 energy channels could be tested by simulating food webs with different distributions of 1460 modules and running analyses such as sequential node deletions to compare the relative effects 1461 of overall modularity and module distribution on network robustness (Dunne et al. 2004). It is 1462 important that such analyses incorporate link weighting and indirect effects such as population 1463 dynamics to avoid the underestimation of secondary extinctions (Zhao et al. 2016).

#### 1464 *3.4.3 Further considerations*

1465 We explored the potential mechanisms underlying the modular structure of marine food webs, 1466 but more highly resolved networks across a range of ecosystem types are necessary to 1467 generalise our results. While most nodes were resolved to the genus or species level, a small 1468 minority of basal and consumer groups in each network were subject to greater aggregation. 1469 It has previously been suggested that certain topological metrics such as linkage density and 1470 mean chain length are sensitive to the level of aggregation employed, though there has been 1471 no explicit investigation of the effects on modularity (Martinez 1993). However, it has also 1472 been demonstrated that there is no consistent relationship between species richness and 1473 modularity at different scales (Montoya et al. 2015); This suggests that slight 1474 underestimations of the number of species (and links) resulting from species aggregation are 1475 unlikely to have a material impact on the resulting modular structure of our focal networks.

A further unknown is the effect of different data compilation approaches on network structure. Each of the focal food webs was compiled using species- and region -specific diet information, but also broader literature sources spanning variable taxonomic and spatial resolutions, which increases the level of uncertainty over some interactions. However, modularity has been found to be relatively robust to variation in sampling effort (Rivera-Hutinel et al. 2012), and each of the focal ecosystems has been subject to extensive long-term sampling whereby the ecology of 1482 most constituent species is well understood. The underlying core species list and structure of 1483 each of our focal networks is therefore likely to be robust to minor variation in the distribution 1484 and number of interactions and nodes. We encourage researchers to provide information 1485 regarding the number of data sources used to determine the diet composition of each species 1486 and some indication of sampling completeness (e.g. yield-effort curves), and to ensure minimal 1487 and comparable levels of taxonomic aggregation, as these efforts will facilitate the assessment 1488 of the comparability of network structures (Martinez 1993; Gauzens et al. 2013). While 1489 network size alone does not drive modularity (Rivera-Hutinel et al. 2012; Montoya et al. 2015), 1490 contrasts between our study datasets may have arisen from a combination of natural and 1491 arbitrary differences in the scale at which the network is considered. In this study, the two 1492 offshore food webs had arbitrary differences in their boundaries and constituent species (e.g. 1493 omission of the sea floor, marine mammals, and seabirds in the Scotia Sea), but their modular 1494 structure was still consistent, which provides some confidence in our ability to detect 1495 overarching trends despite methodological differences. Food web modules have been found to 1496 represent distinct functional groups (i.e. groups of species with similar ecological functions 1497 such as pollination, herbivory, predation etc.; Montoya et al. 2015). Therefore, there is a risk 1498 that omitting species with certain characteristics when describing food webs means we only 1499 capture part of the processes structuring ecosystems, and that our perception of modularity or 1500 stability is influenced by the scale at which the network is considered. This is a topic which 1501 merits further investigation and should certainly be discussed when comparing networks.

#### 1502 3.4.4 Conclusion

1503 This study provides insight into the underlying drivers of modularity in marine food webs 1504 through the comparison of multiple highly resolved networks. Modules in relatively stable 1505 offshore environments appear to be structured largely by body mass, while those in more 1506 heterogeneous coastal and intertidal settings are organised according to the broader diversity 1507 of ecological niches and feeding modes. The resulting differences in modular structure (i.e. 1508 trophic clustering of modules versus differentiation into energy channels) could underpin 1509 ecosystem responses to species loss and other perturbations and suggest that traditional 1510 modularity metrics do not fully represent the stability of food webs. Further testing of the link 1511 between the distribution of modules and the degree of network robustness (e.g., using 1512 simulated networks and species extinction scenarios) will ensure that we continue to make 1513 progress towards gaining a comprehensive understanding of the underlying determinants of 1514 network stability.

# **4 Temperature alters the predator-prey size relationships**

## and size-selectivity of Southern Ocean fish.

1517 Published in *Nature Communications* (https://doi.org/10.1038/s41467-024-48279-0)

1518 Abstract

1519 A primary response of many marine ectotherms to warming is a reduction in body size, to 1520 lower the metabolic costs associated with higher temperatures. The impact of such changes 1521 on ecosystem dynamics and stability will depend on the resulting changes to community size-1522 structure, but few studies have investigated how temperature affects the relative size of 1523 predators and their prey in natural systems. We utilise >3,700 prey size measurements from ten Southern Ocean lanternfish species sampled across  $>10^{\circ}$  of latitude to investigate how 1524 1525 temperature influences predator-prey size relationships and size-selective feeding. As 1526 temperature increased, we show that predators became closer in size to their prey, which was 1527 primarily associated with a decline in predator size and an increase in the relative abundance 1528 of intermediate-sized prey. The potential implications of these changes include reduced top-1529 down control of prey populations and a reduction in the diversity of predator-prey 1530 interactions. Both factors could reduce the stability of community dynamics and ecosystem 1531 resistance to perturbations under ocean warming.

#### 1532 4.1 Introduction

1533 Global warming represents a major threat to the structure and functioning of ecosystems. One 1534 possible consequence of rising temperatures is a decrease in body size across many species 1535 and communities (Daufresne et al. 2009). At the individual level, warming alters the 1536 physiology of organisms and is likely to reduce body sizes within populations as organisms 1537 attempt to maintain metabolic functioning (Daufresne et al. 2009; Deutsch et al. 2022). At the 1538 community level, warming may alter assembly processes through environmental filtering, 1539 competition, or trophic interactions, which may result in communities dominated by smaller-1540 bodied species (Daufresne et al. 2009; Rutterford et al. 2023). The subsequent impacts on 1541 population abundances and species interactions can drive changes to structure and 1542 functioning at the ecosystem scale (Brierley and Kingsford 2009). Aquatic ectotherms such as 1543 fish are particularly susceptible to temperature-induced reductions in body size, due to the 1544 lower rates of oxygen diffusion in water and the energetic costs associated with maintaining 1545 water flow over surfaces (Forster et al. 2012). Additionally, gape limited feeding means that

1546 many fish species display ontogenetic changes in prey selection, with larger predators 1547 consuming larger, more energetically valuable prey (Scharf et al. 2000; Sánchez-Hernández 1548 et al. 2019). Declines in prey size with warming may therefore reduce the rates of energy 1549 acquisition by larger predators, resulting in reduced fish growth and smaller overall body sizes within populations (Queiros et al. 2024). Furthermore, such altered prev size 1550 1551 distributions may favour smaller-sized predator species, providing them with a competitive 1552 advantage and thereby shifting the fish community composition towards smaller body sizes 1553 (Gjoni et al. 2023). Evidence from the last interglacial period suggests that fish communities 1554 experienced declining body size in response to warmer conditions (Agiadi et al. 2022; 1555 Salvatteci et al. 2022), and the average size of contemporary fish is expected to show a 1556 similar pattern under the current rate of global warming (Cheung et al. 2013). However, there 1557 is currently little understanding of how these changes will impact the structure and stability of 1558 marine ecosystems.

1559 Body mass is a key life-history trait which determines factors such as consumption rates, 1560 handling times, and gape size (Petchey et al. 2008; Potapov et al. 2019). As such, body mass 1561 provides an important link between individual physiology and food web structure and is 1562 therefore often used to parameterise models of population dynamics and energy flow within 1563 ecosystems (Boit et al. 2012; Martinez 2020). In the marine environment, predators are 1564 generally larger than their prey, and the predator-prey mass ratio (PPMR) is a good predictor 1565 of trophic interactions. For example, allometric diet breadth models accurately predict who 1566 eats who in aquatic ecosystems (Petchey et al. 2008), whilst declines in PPMR typically result 1567 in lower per capita interaction strengths as predators are able to gain the same amount of 1568 energy by consuming fewer large prey (Brose et al. 2006). At the community level, larger 1569 ectotherms may decline in size more rapidly than smaller ectotherms with warming as a result 1570 of their reduced surface area to body mass ratio and the associated challenge of maintaining a 1571 higher metabolic rate (Forster et al. 2012; Petrik et al. 2020). This is particularly true for the 1572 marine environment, where larger fish and invertebrates display the strongest temperature-1573 size responses (Lavin et al. 2022). If warming causes a greater decline in the size of 1574 ectotherm predators relative to that of their smaller prey (i.e. changes in community size 1575 structure), the average PPMR might decrease, with consequences for interaction strengths 1576 and thus energy flow through marine ecosystems.

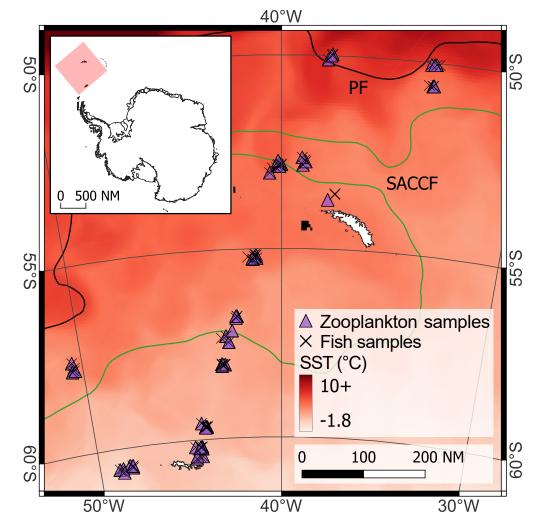
1577 The physiological basis for temperature effects on PPMR at the community level may be1578 complicated by behavioural responses to environmental change. For example, predators may

1579 select for more nutritious (larger) prey in an effort to increase per capita energy intake under 1580 energetically stressful conditions, thus reducing their PPMR (Lemoine et al. 2013; O'Gorman 1581 et al. 2016). Alternatively, predators might feed in a more density-dependent manner, 1582 consuming a greater proportion of abundant but relatively smaller prey and thereby 1583 increasing PPMR. Importantly, behavioural responses are unlikely to be uniform across 1584 predator body sizes, given the different dietary niches of small and large organisms and their 1585 differential susceptibility to warming. Previous research has identified variable size-1586 dependent relationships between PPMR and temperature, such that both systematic increases 1587 (Dobashi et al. 2018) and decreases (Gibert and Delong 2014) to per capita interaction

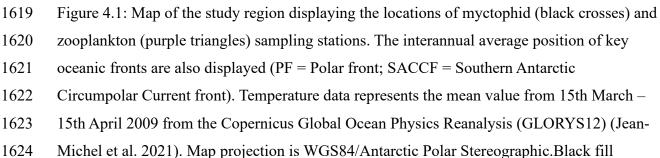
1588 strength are possible.

1589 It is clear we still have limited understanding of how temperature-driven changes in body size 1590 may alter community-level feeding relationships, and it is vital to address this knowledge gap 1591 if we are to predict ecosystem responses to warming. This is particularly true for the Southern 1592 Ocean, which is experiencing widespread environmental changes including rapid regional 1593 warming in areas such as the western Antarctic Peninsula (Meredith and King 2005) and 1594 northern Scotia Sea (Whitehouse et al. 2008). The Southern Ocean supports a diverse array of 1595 higher predator populations including seabirds, seals, penguins, and whales, with a food web 1596 largely centred around krill (particularly Euphausia superba) (Hill et al. 2006). However, it is 1597 expected that krill will shift their distribution southward in response to ocean warming 1598 (Atkinson et al. 2019), with potentially drastic consequences for many regional predator 1599 populations unless other suitable prey are available (Klein et al. 2018). Previous research has 1600 identified mesopelagic lanternfish (Family Myctophidae, hereafter myctophids) as one such 1601 potential alternative resource, due to their extremely high biomass and their role in supporting 1602 energy flow to higher predators including seals and penguins during periods of low krill 1603 availability (McCormack et al. 2021b). Additionally, myctophids themselves are major 1604 generalist consumers of prey including krill, amphipods and copepods, and therefore exert 1605 significant influence over food web dynamics (McCormack et al. 2020). Myctophids are 1606 strongly size distributed in the Southern Ocean, with smaller species and individuals found at 1607 lower (warmer) latitudes (Saunders and Tarling 2018), and they display clear size-selectivity 1608 in their feeding (Cherel et al. 2010; Saunders et al. 2019). Warming may therefore alter the 1609 size distribution of myctophids and the size relationships between these predators and their 1610 prey, and it is important that we understand what these likely changes will be in order to 1611 model ecosystem responses.

- 1612 In this study, we assessed the relationship between temperature and the relative sizes of
- 1613 myctophids and their prey using a dataset of 1,576 stomachs and 3,707 prey size
- 1614 measurements from ten myctophid species sampled across >10° of latitude in the Southern
- 1615 Ocean (Figure 4.1). We hypothesised that myctophids would exhibit a decline in PPMR with
- 1616 increasing temperature, due to (1) a greater decrease in the size of these predators versus their
- 1617 prey, and/or (2) predators selecting for larger prey as temperature increases.







1625 represents missing temperature data. Map produced using QGIS 3.28 Firenze.

#### 1626 *4.2 Materials and methods*

All data used in this study were collected following standard protocols and ethic approval
from the British Antarctic Survey and the Environmental Protocol (1991) of the Antarctic
Treaty.

#### 1630 4.2.1 Fish sampling

1631 Myctophids were collected during three research surveys conducted in austral spring (JR161, 1632 Oct-Dec 2006), summer (JR177, Jan-Feb 2008) and autumn (JR200, Mar-Apr 2009) in the 1633 Scotia Sea in the Atlantic sector of the Southern Ocean. Fish were sampled at stations across 1634 a transect spanning the entire Scotia Sea, from the Antarctic Polar Front to the sea ice zone. 1635 The exact location of these stations varied between cruises but was similar across years, with a broad latitudinal range sampled during each cruise (Figure C1-C2). Sampling was 1636 conducted using a depth-stratified 25 m<sup>2</sup> rectangular mid-water trawl net (RMT25), deployed 1637 at depth ranges of 0-200, 200-400, 400-700, and 700-1000 m (Figure 4.1). The nets had a cod 1638 1639 end mesh size of 5 mm. Hauls were conducted during both light and dark conditions in spring

and summer, but only darkness during autumn, due to a reduced daylight period.

1641 Fish were processed on-board and identified to species level where possible, with standard

1642 length (SL) measured to the nearest millimetre. A random subsample of 25 fish per species

1643 (or all individuals in the case of small catches) were set aside for stomach dissection. These

1644 stomach samples were then frozen at -20 °C for later laboratory analysis, where the stomach

1645 contents were thawed and identified to the lowest taxonomic level possible. For each

1646 stomach, the number of individuals and average weight of each prey taxon was recorded

1647 using a motion compensated balance. The resulting datasets can be accessed via the UK Polar

1648 Data Centre (Collins et al. 2020; Belcher et al. 2019).

1649 For this study, fish SL was converted to mass in grams using species-specific length-weight

1650 equations from the British Antarctic Survey's long-term records (Table C1) for those

1651 individuals which did not have empty stomachs. This was done for ten species (see Table

1652 C1), while data for a further two species were omitted due to very low sample sizes (n = 7 for

1653 *Gymnoscopelus opisthopterus*, n = 1 for *G. piabilis*). The final datasets used in this study

1654 consisted of 3,707 prey records from 1,576 fish stomachs (Table C2), in addition to a larger

- 1655 set of fish body size estimates from 6,143 individuals (the majority without stomach content
- 1656 data; Table C3) and species-specific abundance estimates for each sampling location.

#### 1657 4.2.2 Zooplankton sampling

1658 Macrozooplankton samples were collected from RMT25 nets, while mesozooplankton were 1659 sampled using paired Bongo nets (mesh size 50 µm), which were deployed to a depth of 400 1660 m during daylight hours (Ward et al. 2012; Tarling et al. 2012a; Tarling et al. 2012b). 1661 Zooplankton samples were preserved in 4% formalin with seawater and analysed in the 1662 laboratory, with taxa identified to the lowest possible taxonomic resolution. The total wet 1663 weight (g) was calculated for each macrozooplankton taxon using a motion compensated 1664 balance and divided by the number of individuals to estimate the mean body mass for each 1665 taxon. Mesozooplankton taxa were assigned an average dry mass (DM, mg) from published 1666 sources, which were converted to wet mass (WM, g) using general DM to WM conversion 1667 factors in Atkinson et al. (2012). Abundance values for macro- and mesozooplankton (standardised to individuals m<sup>-2</sup>) were calculated using the estimated area sampled by the 1668 nets. Copepods dominated the zooplankton community by abundance, constituting over 70% 1669 1670 of total density on average across hauls, followed by polychaetes and chaetognaths and, to a 1671 lesser extent, pteropods and ostracods (Table C4). The original zooplankton data are as 1672 presented in (Tarling et al. 2012a) and can be accessed from the UK Polar Data Centre (Ward 1673 et al. 2020).

#### 1674 *4.2.3 Environmental covariates*

1675 We extracted daily sea-surface temperature (SST) values for the coordinates of each station 1676 from the 1/12° gridded Copernicus Global Ocean Physics Reanalysis product GLORYS12V1 1677 (Jean-Michel et al. 2021). To investigate the consistency of results at depth, we also extracted 1678 modelled temperature data from the GLORYS12V1 ~1062m depth bin, which is the closest 1679 match to the lower depth limit of the trawls. Temperature data were averaged for the 30 days 1680 prior to and including the day of sampling. To identify the potential influence of local 1681 productivity on myctophid feeding relationships, we also extracted surface chlorophyll-a 1682 (Chl-a) values from the Copernicus-GlobColour dataset, which has a spatial resolution of 4x4 1683 km (Garnesson et al. 2019). As with the temperature data, daily Chl-a values at each station 1684 were averaged for the 30 days prior to and including the day of sampling. See Figure C3 for 1685 an overview of the relationship between temperature and latitude. The remaining methods 1686 refer to analyses involving SST but see Supplementary Information for an overview of the 1687 results of modelling with temperature at depth. We did not consider the effects of spatial 1688 heterogeneity in fishing effort as there is currently no targeted myctophid fishery in the 1689 Southern Ocean. Fish constitute the majority of bycatch by the winter krill fishery in the

- 1690 Scotia Sea but appear to consist predominantly of members of the Channichthyidae and
- 1691 Nototheniidae (Krafft et al. 2023). Overall annual average bycatch weights across all bycatch
- 1692 taxa (0.1-51.3 tonnes) are low compared to the estimated biomass of mesopelagic fish in the
- 1693 Scotia Sea (~4.5 million tonnes) and would therefore be expected to have negligible impact
- 1694 on community structure (Krafft et al. 2023).

#### 1695 4.2.4 Statistical analyses

1696 Linear mixed models (LMMs) were used to investigate the relationship between the 1697 environmental variables and multiple metrics related to myctophids and their prey, using the 1698 predator-prey body size dataset. PPMR was calculated as the body mass of each fish predator 1699 (g) divided by the abundance-weighted average prey mass (g) in its stomach. LMMs were 1700 fitted using the function 'lme' in the package 'nlme' (Pinheiro et al. 2023) with either PPMR, 1701 predator body mass, or abundance-weighted mean prey body mass as response variables (each subject to log<sub>10</sub> transformation to meet the assumptions of normality, homogeneity, and 1702 1703 independence of residuals). No strong collinearity was identified between SST and chl-a 1704 (Spearman's rho: -0.077, p = 0.002), therefore these were both entered as explanatory 1705 variables in the same model, including their interaction term. Model selection was then 1706 conducted to identify the best specification of fixed effects (SST and Chl-a) and random 1707 effects (nesting the variables 'year' and 'predator species'). The use of weighted variance 1708 structures to account for heterogeneity in residual variance by year or species was also 1709 investigated during model selection. The absence of spatial autocorrelation in model residuals 1710 was confirmed using Moran's I, therefore autocorrelation structures were not included in the 1711 models. The best model was determined by AIC comparison and visual diagnostics 1712 (heteroscedasticity and normality of residuals). All models incorporated a combined constant 1713 variance structure to account for heteroscedasticity in the errors within both year and predator 1714 species. The final selected models all included a random intercept for year and a random 1715 slope for SST by predator species. Chl-a had no significant main or interactive effects on the 1716 response variables and was therefore omitted from further analyses. See Table C5-C11 for an 1717 overview of the model selection process and Moran's I results for these models.

1718 The selectivity of predators for different prey sizes was estimated by fitting kernel density

1719 distributions to the prey body masses identified in predator stomachs (realised distribution)

and to the comparable range of prey body masses sampled from the environment

1721 (environmental distribution) (Gauzens et al. 2024). The environmental distribution represents

1722 the expected predator diet if feeding is based solely on density-dependent foraging, while the

1723 realised distribution generally represents the combination of such neutral processes and the 1724 active selection for specific prey sizes (Gauzens et al. 2024). This approach assumes that the 1725 diets of these predators are generalist and primarily size-constrained, which is supported by 1726 previous studies of Southern Ocean myctophid diets (Cherel et al. 2010; Saunders et al. 1727 2019). Using the ratio of the realised and environmental distributions, a preference 1728 distribution can be calculated, representing the selectivity of predators for different prey 1729 sizes. To link the predator diets to the distribution of potential prey sizes in the environment, 1730 we grouped predators and zooplankton samples which were collected in the same area and 1731 within a few days of one-another, resulting in a total of 24 separate sampling locations 1732 spanning the study region. Within these groups, we then aggregated predators from the same species into size-classes of  $10^{0.05}$  g to ensure that enough prey were present in the combined 1733 diets to reliably estimate a density distribution, whilst ensuring there were enough data points 1734 for later analysis (n = 164). The final size classes ranged from  $10^{-0.525} = 0.30$  g to  $10^{1.575} =$ 1735 37.58 g. For each aggregation, an average temperature was estimated from the constituent 1736 1737 stations. We used the mean value of the preference distribution for each size class to represent 1738 the average preferred prey size of predators at each temperature. We then used a LMM to 1739 investigate the relationship between preferred prey size and the interaction between 1740 temperature and predator size-class, following the same approach to model specification and 1741 selection as described above. The final model included random intercepts for year and 1742 predator species, and a combined variance structure for year and predator species (see Table 1743 C12-C13 for an overview of the model selection process).

1744 To differentiate the potential individual-level and community-level mechanisms underlying 1745 trends in body size with temperature, we also conducted analyses of predator body size and 1746 community composition using a larger dataset of individual body sizes and species 1747 abundance estimates (n = 6,143). We fitted a Generalised Least Squares (GLS) regression 1748 model of species diversity (Shannon-Wiener (log e) diversity index) as a function of SST and 1749 Chl-a to investigate whether there was any change in community structure with 1750 environmental conditions. For this analysis, densities of each species caught during each haul 1751 were estimated by multiplying counts by the product of the distance towed multiplied by the nominal net mouth area (25  $m^2$ ), and then standardised to values of individuals per 1,000  $m^{-3}$ . 1752 1753 A square-root transformation was then applied to the density estimates to reduce the 1754 weighting of dominant species. An LMM was fitted to the relationship between body mass 1755 and the interaction between SST and Chl-a at the community level, before linear models of

- body size and SST were fitted for each predator species individually, to identify whether
- 1757 community-level trends in size with temperature were present at the population level. The
- 1758 optimal model structure for each species-level analysis varied, and very low but statistically
- 1759 significant levels of spatial autocorrelation were identified for a small number of species and
- dealt with by incorporating spatial autocorrelation functions. See Table C14-C22 for model
- 1761 selection of the optimal variance weighting, random and fixed effects structures, Moran's I
- 1762 test results and implemented autocorrelation structures, and model outputs.

#### 1763 *4.3 Results and discussion*

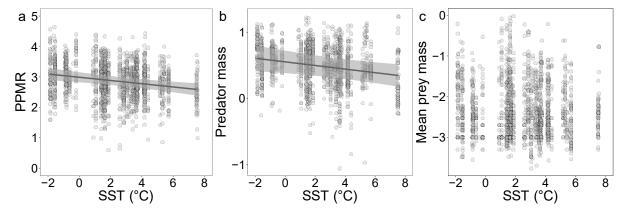
1764 PPMR declined by ~11% per °C increase in sea surface temperature (SST), associated with a 1765 significant decline in predator body size at a rate of ~6% per °C and no coherent trend in the 1766 mean body size of prey in the diet (Figure 4.2; Table 4.1). Chlorophyll *a* was initially used as 1767 a further explanatory variable but it was not significant in any model and was therefore 1768 excluded during model selection (Table C5-C11). The same general results were found when 1769 temperature at ~1000m (the estimated maximum of myctophid depth distributions) was 1770 considered instead of SST (Figure C4, Table C23Table C30). A similar decline in predator 1771 body size was also found when using a larger dataset of fish body masses (n = 6.143, the 1772 majority without stomach content data; Figure C5, Table C14-C16). In all, seven of the ten 1773 myctophid species also displayed significant declines in size with increasing temperature 1774 (Figure C6, Table C17-C19). Together, these results suggest that the decline in PPMR is 1775 associated with a greater decrease in the size of these predators relative to their prey as 1776 temperature increases.

1777 The effect of declining PPMR on interaction strengths will depend on the interactive effects 1778 of temperature and body mass on metabolism and consumption (Kratina et al. 2022), making 1779 it difficult to predict the consequences for ecosystem stability. It has previously been found 1780 that temperature alters the directionality and shape of the relationship between PPMR and 1781 predator attack rate and prey handling time, with low PPMR destabilising community 1782 dynamics under warming due to elevated predation rates at low prey density (Kratina et al. 1783 2022). Additionally, when declines in body mass under warming are restricted to isolated 1784 trophic levels, community stability is expected to be reduced (Sentis et al. 2017), possibly 1785 due to lower top-down control of prey populations (Shackell et al. 2010). However, while the 1786 reduced ingestion efficiencies and higher metabolic costs associated with higher temperatures are expected to make predator populations increasingly vulnerable to starvation, this effect is 1787 1788 exacerbated under high PPMRs (Rall et al. 2009), therefore the observed decline in predator

size with warming may in fact provide a buffer against population crashes. Ultimately, the effects of warming and PPMR on the strength of interactions will depend on factors including predator and prey identity, predator body size, and thermal tolerance. Further investigations of the combined effects of temperature and PPMR on interaction strengths will be important for determining the possible consequences of altered size-structuring of predator-prey interactions for the stability of ecological communities. This could be facilitated through the

application of ecosystem flux or dynamical population models (Gauzens et al. 2019;

1796 Sohlström et al. 2021).



1797

Figure 4.2: Effects of temperature on predator and prey body mass. (a) partial residual plot from a linear mixed model of the effect of sea-surface temperature (SST) on prey-averaged predator-prey mass ratio (PPMR); (b) partial residual plot from a linear mixed model of the effect of SST on predator body mass; (c) scatterplot of the relationship between SST and abundance-weighted average prey mass in predator stomachs. Y-axis values are in log<sub>10</sub> g. Lines represent predicted values at each SST. Shading represents 95% confidence intervals.

1804 Table 4.1: Model statistics for the effect of temperature on predator and prey body masses.

1805 Output from linear mixed models with predator-prey mass ratio (PPMR), predator body mass

1806 and abundance-weighted average prey body mass in predator stomachs as response variables

1807 (all log<sub>10</sub>). SST represents sea-surface temperature. R<sup>2</sup>m and R<sup>2</sup>c represent the Nakagawa's

1808 marginal and conditional model R<sup>2</sup> values, respectively.

Model	Coefficient	Estimate	SE	DF	t-value	p-value
PPMR	Intercept	2.988	0.085	1550	35.251	< 0.000
						1
	SST	-0.053	0.015	1550	-3.601	0.0003
$R^2m = 0.049, R^2c = 0.493$						
Predator body mass	Intercept	0.552	0.087	1550	6.346	$<\!0.000$
						1
	SST	-0.027	0.011	1550	-2.482	0.0132
$R^2m = 0.024, R^2c = 0.978$						
Mean prey body mass	Intercept	-2.371	0.069	1551	-34.249	< 0.000
-	_					1
$R^2m < 0.001, R^2c = 0.455$						

# 1809

1810 The observed decline in predator size with increasing temperature fits the wider expectation 1811 that a primary response of ectotherm vertebrates, including Southern Ocean myctophids, to 1812 warming should involve a reduction in individual body size and shifts in overall community 1813 size structure (Deutsch et al. 2022; Rubalcaba et al. 2020). Declines in size at the individual 1814 level are thought to facilitate continued persistence with warming by minimising the extent to 1815 which metabolic rate must increase to match the greater energetic demands of the 1816 environment (Riemer et al. 2018). Changes in community size structure may also be the 1817 result of a combination of physiological and competitive processes which result in species of 1818 a certain size range becoming dominant (Gjoni et al. 2023). There was a significant increase 1819 in Shannon diversity of the myctophid community with increasing SST, associated with a 1820 shift in species abundances from communities dominated by a few large-bodied species (e.g. 1821 *Electrona antarctica*) at cold high latitudes to a more even distribution of abundances in the 1822 more northerly warmer regions (Figure C7, Table C20-C22), as previously documented 1823 (Collins et al. 2012). This indicates that the link between temperature and body size at the 1824 community level may be driven in part by community assembly processes which select for 1825 species of different sizes as it becomes warmer, e.g. smaller predators are able to outcompete 1826 larger ones under the altered prey size distribution and relatively lower metabolic demands. 1827 However, our analyses of the relationship between body mass and SST at the population level also revealed significant declines in size with increasing temperature for many of the

1829 myctophid species, both for large-bodied taxa such as *E. antarctica* and for small species like

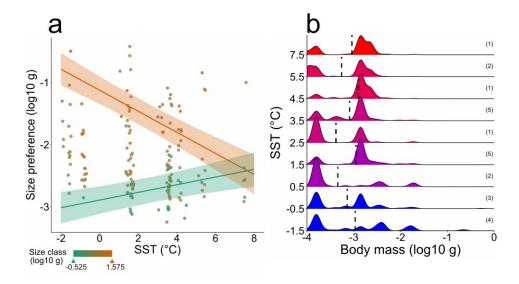
1830 *Krefftichthys. anderssoni* (Figure C6, Table C17-C19). The observed trends at the community

1831 level therefore are not explained by community assembly processes alone, but also by

1832 temperature effects on populations, likely mediated by physiological responses to warming.

1833 Under both moderate and high emissions scenarios, Antarctic waters are expected to become 1834 increasingly favourable for smaller, sub-Antarctic myctophid species, likely altering 1835 community diversity and size structure (Freer et al. 2019). Such changes may reduce their 1836 suitability as prey for predators such as penguins and seals, with knock-on effects on these higher predator populations and food web dynamics (Murphy et al. 2007). Additionally, many 1837 1838 myctophid species display size-selective feeding, with a switch from euphausiids and fish to 1839 smaller copepods as their body size decreases (Saunders et al. 2019). Thus, a reduction in the 1840 average size of myctophids may alter the diversity and size distribution of the prey 1841 community as predation rates on different species change (Rudolf 2012; Ives et al. 2004). 1842 Furthermore, smaller species are generally expected to have fewer feeding interactions across 1843 a more restricted range of trophic levels, which could alter the distribution of energy flow by reducing network complexity and trophic redundancy (Brose et al. 2017). 1844

1845 To investigate the evidence for size-selective feeding behaviour that could further underlie 1846 the decline in PPMR with temperature, we conducted an analysis of dietary size preferences 1847 for prey in the environment in relation to predator body size class and temperature (see Methods). Predator size and SST had a significant interactive effect on preferred prey size, 1848 1849 with small predators feeding on relatively larger prey and large predators feeding on 1850 relatively smaller prey in warmer regions (Figure 4.3a; Table 4.2). This partially supports 1851 hypothesis 2, that predators will select for larger prey in warmer environments, but not for the 1852 largest fish. This result may be explained by an increase in the relative abundance of 1853 intermediate prey sizes within the range of body masses commonly consumed by the fish 1854 (Figure 4.3b).



1855

Figure 4.3: Predicted interactive effect of SST and fish size class on myctophid average 1856 preferred prey size. Lines represent predicted values at each SST, for the largest and smallest 1857 1858 predator size classes. Shading represents 95% confidence intervals. Points are coloured 1859 according to size class, jittered slightly for clarity. (b) Density plots of zooplankton body mass 1860 distribution in the environment within size range commonly consumed by the myctophids, grouped into 1 °C temperature bins. Dashed lines represent abundance-weighted average body 1861 1862 mass. Y-axis indicates central temperature value for each bin. Values in brackets indicate 1863 number of hauls. Note: in panel b, large prey sizes (above approx.  $-2 \log_{10} g$ ) are present at all 1864 temperatures but extremely low abundance relative to smaller organisms prevents them from being visible. 1865

Table 4.2: Model statistics for the effect of temperature on predatory size preferences. Output
from a linear mixed effects model with mean preferred prey size (log<sub>10</sub>) as the response
variable and sea-surface temperature (SST) and predator size class (log<sub>10</sub>) as explanatory
variables. R<sup>2</sup>m and R<sup>2</sup>c represent the marginal and conditional model R<sup>2</sup> values, respectively.

Coefficient	Estimate	SE	DF	t-value	р	
Intercept	-2.460	0.194	139	-12.673	< 0.0001	
SST	0.006	0.021	139	0.295	0.7681	
Size class	0.848	0.109	139	7.795	< 0.0001	
SST*Size class	-0.110	0.031	139	-3.483	0.00071	
$R^2m = 0.298, R^2c = 0.801$						

1871 Our results suggest that temperature influences the size-structuring of feeding relationships 1872 within the Southern Ocean mid-trophic community through a combination of density-1873 dependence and active selection. Under colder conditions, large predators appear to select for 1874 relatively abundant, large, energetically valuable prey while small predators feed on small 1875 prey. Under warmer conditions, the shift in the distribution of suitable prey sizes towards 1876 intermediate body masses restricts the feeding behaviour of large predators and forces them 1877 to feed sub-optimally on smaller prey while small predators actively select for these abundant 1878 intermediate prey sizes, possibly because they provide greater per capita energy intake. This 1879 reduction in prey size diversity could constrain the foraging niches of smaller and larger 1880 predators, increasing competition and, under the general expectation that food web 1881 complexity promotes predator population stability (Petchey 2000), potentially destabilising 1882 predator-prey dynamics. Larger predators may also be forced to feed on prey that are smaller 1883 than their optimal foraging niche, thus preventing them from meeting their higher energetic 1884 demands under warmer conditions. These changes in size-selectivity may also explain the 1885 increasing prevalence of smaller myctophid species in warmer regions (Figure C7), as they 1886 can capitalise on the available prey field and outcompete their larger counterparts. The 1887 increasing dominance of smaller myctophids, which feed preferentially on larger prey in 1888 warmer regions, is likely to drive the observed decline in overall PPMR across the predator community. Thus, we suggest that the observed patterns in myctophid size and foraging with 1889 1890 temperature are likely to be the result of a combination of interacting processes acting at both 1891 the population and community levels, and we encourage further efforts to disentangle them. 1892 Overall, our results highlight the importance of considering the size structuring of biotic 1893 interactions and plasticity of size-based foraging behaviour when investigating the possible 1894 consequences of environmental change for community structure and composition.

1895 We investigated a temperature gradient across a large spatial scale (>10° of latitude) rather 1896 than directly testing the effects of temperature change over time. Such temporal changes are 1897 difficult to investigate in-situ, but mesocosm experiments could provide insight into how 1898 rapid warming affects species body sizes and biotic interactions. However, the results of such 1899 studies primarily relate to the plastic responses of individuals over the short-term, which may 1900 differ from the adaptive responses of populations to sustained gradual warming over the 1901 multi-decadal timescales that are relevant to ongoing climate change. In contrast, given the 1902 historically stable temperatures of the Southern Ocean (Morley et al. 2020), our space-for-1903 time substitution represents the long-term eco-evolutionary adaptation of predator and prey

1904 communities. One potential caveat of our approach was the use of sea-surface temperatures to 1905 represent the environmental conditions experienced by the myctophids, as temperatures at 1906 depth may differ from those at the surface. Indeed, while a positive relationship between 1907 latitude and temperature is still apparent at approximately 1,000m depth, the trend is weaker 1908 than at the surface (Figure C5). When substituting SST with the temperature at depth in our 1909 analyses, however, the results are consistent (Figure C4, Table C23-C30), suggesting that the 1910 observed relationships hold across the depth range that myctophids are thought to inhabit.

1911 As our oceans continue to warm, significant changes to the size structuring of marine 1912 communities are likely to occur in many regions, and the use of dietary preference analyses 1913 such as this will be useful for disentangling the interactive effects of behaviour and 1914 physiology on the feeding ecology of key species and functional groups. Myctophids are one 1915 of the most abundant fish families globally and a major component of many pelagic food 1916 webs, from the poles to the tropics (Morley et al. 2020; Chaudhary et al. 2021). The insights gained in this study therefore have relevance for other open ocean systems, including those 1917 1918 near the equator where warming is expected to drive strong declines in body size and changes 1919 to the distribution of many mesopelagic species (Chaudhary et al. 2021; Lefort et al. 2015). 1920 Changes in species composition with temperature may also alter community PPMR in 1921 unexpected ways, as it has previously been found that the relationship between individual 1922 body mass and PPMR varies between taxa, due to factors such as morphology and feeding 1923 strategy (Reum et al. 2019). It will therefore be important to expand these analyses to other 1924 regions and taxa to provide an overview of the generality of the observed relationships.

1925 Rising metabolic costs and oxygen limitation resulting from ocean warming are expected to 1926 drive declines in the body size distribution of many marine ectotherms (Deutsch et al. 2022; 1927 Forster et al. 2012), and we sought here to investigate the potential consequences for the size-1928 structuring of species interactions. Using an extensive dataset spanning a large latitudinal 1929 range, we have shown that increasing temperature is associated with changes in body mass 1930 and dietary size-selectivity across Southern Ocean myctophids, a key component of pelagic 1931 food webs, resulting in predator communities that are closer in size to their prey. As a result, 1932 warming might alter prey population dynamics and reduce top-down control, potentially 1933 reducing community stability. The shift in predator-prey size relationships could also drive a 1934 reduction in the diversity of predator-prey interactions and a loss of redundancy within 1935 ecological networks, which may reduce their resistance to perturbations. The trends identified 1936 in this study provide a basis for mechanistic models to investigate the potential consequences

- 1937 of warming scenarios for the structure of biotic interactions and the stability of ecosystems.
- 1938 Efforts to investigate these relationships in other regions and for other taxa will aid the search
- 1939 for macroecological patterns that can be used to predict ecosystem responses to climate
- 1940 change.
- 1941

# 1942 5 Trade-offs between the recovery of Southern Ocean 1943 baleen whales and conservation of their competitors

1944 In preparation for *PNAS* 

1945 Abstract

1946 The historical over-exploitation of Southern Ocean baleen whales is thought to have resulted 1947 in the reorganization of food webs and the expansion of competitor populations such as seals 1948 and penguins. Many whale populations are now recovering, leading to uncertainty for the 1949 impact this will have on their krill-feeding competitors, which are themselves a focus of 1950 conservation efforts. We used a circumpolar suite of standardized regional Ecopath models to 1951 explore the potential ecological trade-offs associated with increases in baleen whale 1952 populations. There was variation in the capacity of ecosystems to support increases in whale 1953 consumption while also sustaining competitor populations at close to their contemporary 1954 estimates. Under median estimates of daily krill consumption by baleen whales, only limited 1955 increases in whale biomass were possible without major reductions in competitor 1956 populations, although the impacts of whale population recovery would be mitigated under 1957 plausible future increases in primary production. We identified that the level of unexploited 1958 production by whale prey, alongside the degree of dietary overlap between whales and their 1959 competitors, are associated with the ecosystem capacity to support whale population 1960 increases and could be used to guide decision-making in relation to the implementation of 1961 regional conservation actions. Ultimately, it must be recognized that contemporary and future 1962 Southern Ocean ecosystems may have reduced capacity to sustain higher trophic levels, resulting in strong trade-offs between conservation objectives. 1963

#### 1964 5.1 Introduction

1965 Southern Ocean ecosystems are complex and diverse, supporting a variety of vital services 1966 including provisioning, biogeochemical processes and nutrient cycling (Cavanagh et al. 1967 2021). Many species there are unique and endemic, with large populations of marine 1968 mammals and seabirds sustained by a variety of mid-trophic taxa ranging from copepods to 1969 Antarctic krill (Euphausia superba) and highly abundant demersal and pelagic fish (Queirós 1970 et al. 2024). Over the past two centuries, however, Southern Ocean ecosystems have been 1971 considerably impacted by anthropogenic activities including the exploitation of seals and 1972 whales and the more recent harvesting of finfish and Antarctic krill (Miller 1991; Kock et al. 1973 2007). Climate change is also affecting the Southern Ocean, with regional warming and shifts 1974 in sea ice extent and duration, which could negatively impact the distributions and population 1975 dynamics of species such as krill and emperor penguins (Aptenodytes forsteri) (Auger et al. 1976 2021; Meredith et al. 2019; Fretwell et al. 2023). Widespread increases in productivity are 1977 also predicted for various regions, but shifts in the phytoplankton community size structure 1978 may have negative consequences for ecosystem function by driving changes in the 1979 composition of zooplankton communities, with a dominance of salps resulting in reduced 1980 efficiency of energy transfer to higher predators (Kawaguchi et al. 2024; Pinkerton et al. 1981 2021; Queirós et al. 2024). If we are to implement effective management strategies, it is 1982 imperative that we understand the implications of past and future ecological shifts and the 1983 possible trade-offs that may be required to achieve conservation goals.

The exploitation of baleen whales during the 20<sup>th</sup> century represents possibly the most 1984 extensive human impact on Southern Ocean ecosystems to date, resulting in the severe 1985 1986 depletion of many species including blue (Balaenoptera musculus), humpback (Megaptera 1987 novaeangliae), fin (B. physalus) and sei (B. borealis) whales (Christensen 2006; Mori and 1988 Butterworth 2006). These predators exert significant top-down control over Antarctic krill, 1989 zooplankton and fish populations (Tulloch et al. 2019; Bury et al. 2024), and the rapid decline 1990 of the whales will almost certainly have altered energy flow through regional food webs. For 1991 example, the release of Antarctic krill from whale predation is hypothesised to have resulted 1992 in a krill biomass 'surplus' which was then rapidly consumed by other predators including 1993 seals and penguins, increasing their populations (Laws 1977). Ecological modelling has 1994 provided some limited support for this hypothesis, although this is dependent on relatively 1995 high and stable levels of primary productivity (Surma et al. 2014). The contemporary average 1996 total Antarctic krill biomass in the Southern Ocean is estimated to be below 400 million 1997 tonnes (Atkinson et al. 2009; Kawaguchi et al. 2024), far lower than the 600-900 million 1998 tonnes estimated to be required to sustain unexploited baleen whale populations (Smetacek 1999 and Duarte 2008; Surma et al. 2014). Such high Antarctic krill biomass may historically have 2000 been sustained by elevated primary production driven by biological nutrient cycling, as 2001 whales fertilized surface waters with limiting elements such as iron and thereby promoted 2002 krill population growth (Nicol et al. 2010; Ratnarajah et al. 2016). The viability of such 2003 whale-driven surface fertilization for sustaining Antarctic krill populations is, however, 2004 unclear (Maldonado et al. 2016), and observed regional declines in the biomass of Antarctic

krill over the 20<sup>th</sup> century may have been driven more by temperature-related changes in
spawning habitat quality (Yang et al. 2020; Atkinson et al. 2022).

2007 Since the ongoing moratorium on commercial whaling began in 1985, some whale 2008 populations have begun to recover but many remain well below their estimated pre-2009 exploitation levels and there is a widespread desire to see further whale biomass recovery 2010 (Zerbini et al. 2019; Tulloch et al. 2019; Calderan et al. 2020; IWC 2024). It is unclear, 2011 however, whether contemporary Southern Ocean ecosystems can support large increases in 2012 baleen whale populations. A key uncertainty is the extent to which population recovery might 2013 come at the expense of competitor groups for krill such as seals, penguins and fish, which 2014 may have experienced competitive release as a result of whaling. Newly revised estimates of 2015 baleen whale daily consumption rates, which may in fact be up to three times greater than 2016 previously thought (Savoca et al. 2021), suggest that the extent of this potential release may 2017 also have been greater than previously assumed. Such high consumption rates mean the 2018 influence that these whales exert over key prey such as Antarctic krill is also very high, and, 2019 under limited prey availability, their population recovery may therefore require particularly 2020 large compensatory decreases in competitor populations. Two core objectives of the international convention governing conservation in the Southern Ocean are to maintain the 2021 2022 ecological relationships between species and restore depleted populations (Constable 2011). 2023 Given the likely competitive relationships between higher trophic level groups, these 2024 objectives may well be in conflict if the goal is to conserve Southern Ocean ecosystems as 2025 they are now whilst also restoring whale populations. It is therefore important that we explore 2026 the possible implications of baleen whale population recovery for the dynamics of their 2027 competitors. This will provide insight into the likely conservation outcomes that are feasible 2028 under future conditions and could help guide regional management actions.

2029 Food web models provide a tool for understanding how the structure and dynamics of 2030 regional ecosystems might respond to press perturbations such as sustained changes in the 2031 abundance of certain taxa (Montoya et al. 2009). In particular, the Ecopath framework is 2032 often used to model the structure of trophic interactions and energy flow in aquatic food 2033 webs, which can be used to test management scenarios or explore the effects of 2034 environmental changes (Christensen and Walters 2004; Heymans et al. 2016). Ecopath 2035 models have been developed for a variety of locations around the Southern Ocean, ranging 2036 from subantarctic areas such as South Georgia and the Prince Edward Islands to high latitude 2037 regions like the Antarctic Peninsula and Ross Sea (McCormack et al. 2021a). This suite of

2038 models represents a powerful resource for exploring regional similarities and contrasts in 2039 ecosystem structure and function (Hill et al. 2021). In this study, we use six regional Ecopath 2040 food web models in conjunction with a novel objective balancing approach to explore the 2041 potential ecological trade-offs resulting from the recovery of Southern Ocean baleen whale 2042 populations. Our analysis incorporates the full range of plausible whale prey consumption 2043 rates, and assesses the compensatory changes in system productivity required, which might 2044 result from a combination of climate change and whale-mediated nutrient recycling. Using 2045 the Ecopath framework allows us to explore some of the possible mechanisms underlying the 2046 capacity of different models to support increased whale populations. A key parameter in 2047 Ecopath models is the ecotrophic efficiency (EE) of each group, which ranges from zero to 2048 one and represents the extent to which their biomass production is consumed within the 2049 system, with lower values indicating greater 'spare' production which could potentially 2050 support increases in consumption. As a result, models with generally low EEs for baleen 2051 whale prey groups might be expected to have a greater capacity to support increases in baleen 2052 whale consumption without affecting other functional groups, when compared with models 2053 that have generally high *EEs*. Additionally, the degree of overlap in the consumption of prev 2054 groups by baleen whales and their competitors could also determine the capacity of the model 2055 to support increases in whale consumption. If the competitive overlap in prey consumption is 2056 high, we expect to see a greater degree of negative coupling between populations of whales 2057 and their competitor groups, as increases in consumption by whales require relatively larger 2058 compensatory changes in competitor biomasses.

2059 In this study, we implemented two scenarios to investigate the possible consequences of 2060 baleen whale population recovery. Firstly, we explored the capacity of contemporary food 2061 webs to support increasing whale populations by estimating the consequences for competitors 2062 in the absence of compensatory changes in system productivity. This also included an investigation of the influence of different whale prey consumption estimates on the rates of 2063 2064 competitor biomass change. Secondly, we explored the levels of system productivity that are 2065 necessary to support increased whale populations while maintaining contemporary 2066 competitor populations.

#### 2067 *5.2 Materials and methods*

#### 2068 *5.2.1 Modelling framework*

2069 This study makes use of the Ecopath modelling framework, which is used to construct food 2070 web models that meet the assumption of mass-balance, whereby the energy outputs of a 2071 group do not exceed their inputs (Christensen and Walters 2004). As discussed in chapter 1, 2072 Ecopath models represent a specific time period (often a year), with nodes in the food web 2073 representing single life stages, species or, more commonly, aggregated functional groups. The 2074 key parameters required are the biomass (B), diet composition by weight (DC), production 2075 per unit biomass (P/B), consumption per unit biomass (Q/B), and assimilation efficiency (AE) 2076 of each group, although parameters representing biomass accumulation rates, fishery catches 2077 and discards, and migration rates can also be supplied (Christensen and Walters 2004). These 2078 form the basis of linear equations describing the production of each group in terms of their 2079 other parameters and those of their consumers. A key parameter which is often an output of 2080 Ecopath models is the ecotrophic efficiency (EE), representing the proportion of the 2081 production of each group that is used in the system. Values range from zero (limited to top 2082 predators that are not fished) to one (100% of production is consumed by other groups in the 2083 model).

2084 The initial parameterisation often results in an unbalanced model, and *EE* values greater than 2085 one can be used to identify problematic groups with mortality rates that cannot be sustained 2086 by production rates. The most common method of balancing an Ecopath model involves an 2087 iterative process of manual adjustments to group parameters, focussed on unbalanced groups 2088 and their consumers, until balance is achieved (Heymans et al. 2016). Often, the main 2089 parameter that is adjusted is the diet composition as this is often the most uncertain, followed 2090 by biomass and energetic rates (P/B and Q/B). This process can be time-consuming with 2091 many unbalanced groups and may be subjective as the choice of groups and parameters to 2092 adjust, and the magnitude and direction of those adjustments ultimately depend on the 2093 ecological understanding and decisions of the modeller. The balancing process must therefore 2094 be extremely well documented if the results are to be reproducible, and can be aided by 2095 following thermodynamic principles such as ensuring that the growth efficiencies (GE) of 2096 groups fall within expected values (Heymans et al. 2016). To reduce the subjectivity of the 2097 balancing process and facilitate the generation of multiple versions of the same model to 2098 account for parameter uncertainties, efforts have been made to develop various automated balancing algorithms. These range from using the built-in Ecopath Monte-Carlo routine to 2099

- randomly generate alternative balanced parameter sets from a single balanced input model
  (Steenbeek et al. 2018), conducting an exhaustive random search of possible parameter
  combinations that generate a thermodynamically viable (but not necessarily balanced) model
  (Aydin et al. 2005), and making targeted adjustments to unbalanced groups to 'push' the
  model into balance (though this method was removed after Ecopath version 5.1) (Kavanagh
  et al. 2004). These methods all rely on a data 'pedigree' approach, whereby the uncertainties
  around parameter estimates are quantified and used to put bounds on the range of possible
- 2107 values (Heymans et al. 2016).
- 2108 In this study, we needed to generate a large ensemble of balanced models from a set of initial
- 2109 unbalanced input models. This meant that the approaches of Steenbeck et al. (2018) (which
- 2110 uses a balanced input model) and Aydin et al. (2005) (which generates thermodynamically
- 2111 viable, but not necessarily balanced parameters sets) were not suitable. However, the
- 2112 approach of Kavanagh et al. (2004), whereby targeted adjustments are made solely to groups
- 2113 out of balance, was also undesirable as it fails to incorporate uncertainty around the
- 2114 parameters for balanced groups. As a result, we developed a bespoke balancing algorithm and
- 2115 used this to generate 1000 versions of each regional model, to explore the effects of
- 2116 perturbation across a range of plausible alternative model parameterisations (see Model
- 2117 balancing).
- 2118 Our analytical approach in this study involved a number of sequential steps, from the
- 2119 standardisation and balancing of a set of published models to the implementation of
- 2120 perturbation scenarios. The major steps are conceptualised in Figure 5.1 and detailed below.

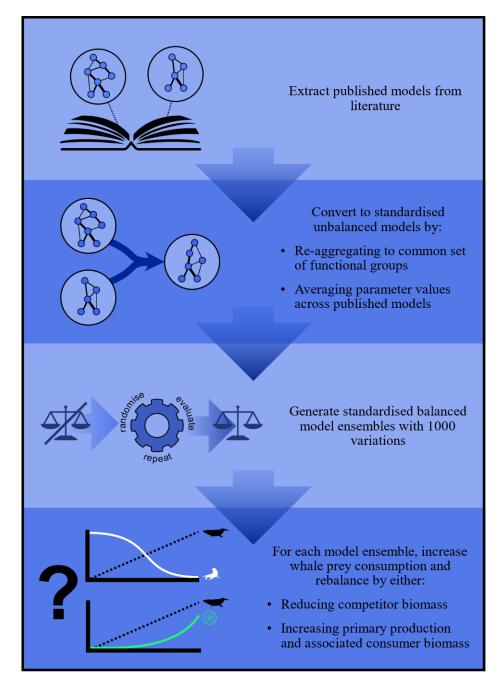


Figure 5.1: Conceptual diagram illustrating the main steps that make up the methods and analyses in this study.

- 2124 5.2.2 Regional Ecopath models
- 2125 We selected six published Ecopath-type food web models, each developed independently by a
- 2126 different group of researchers and representing a different region of the Southern Ocean: The
- 2127 South Georgia shelf (SG; Hill et al. 2012); Ross Sea (RS; Pinkerton and Bradford-Grieve
- 2128 2010); Prince Edward Islands (PE; Gurney et al. 2014); Kerguelen Plateau (KP;
- 2129 Subramaniam et al. 2019); Prydz Bay (PB; McCormack et al. 2020); and West Antarctic
- 2130 Peninsula (AP; Dahood et al. 2019). The model locations are mapped in Figure 5.2 and an

- 2131 overview of the key characteristics of each model is provided in Table 5.1. Each of these
- 2132 models represents the feeding interactions between a set of functional groups (either
- 2133 individual species or aggregates based on taxonomy, size or other aspects of their ecology),
- and provides the key parameters necessary to estimate energy flows within the system.

2135 Table 5.1: Key characteristics of each of the six published Ecopath models used in this study.

Region	South Georgia (SG)	West Antarctic Peninsula (AP)	Ross Sea (RS)	Prince Edward Islands (PE)	Kerguelen Plateau (KP)	Prydz Bay (PB)
Ocean basin	South Atlantic	South Atlantic	Pacific	Indian	Indian	Indian
Latitudinal group	Subantarctic	Antarctic	Antarctic	Subantarctic	Subantarctic	Antarctic
Ecosystem type	Island shelf	Continental shelf	Continental shelf	Island shelf	Island shelf	Continental shelf
Extent	Shelf area between coastline and 1000m depth contour. Approximately 55°S, 36°W	CCAMLR Statistical Subarea 48.1 (approximately 60-70°S and 50- 70°W)	Shelf area between 160°W and 170°E, from 3000m depth contour to permanent ice shelf	200NM radius with centre at 46°46'S, 37°51'E	Between 45- 56°S and 60- 80°E	From 60°S to the Antarctic continent, and 60-90°E
Area (km <sup>2)</sup>	45,530	630,279	637,000	431,014	1,720,348	1,433,028
Biomass units	Grams wet mass km <sup>-2</sup> y <sup>-1</sup>	Grams wet mass km <sup>-2</sup> y <sup>-1</sup>	Grams carbon m <sup>-2</sup> y <sup>-1</sup>	Grams wet mass km <sup>-2</sup> y <sup>-1</sup>	Grams wet mass km <sup>-2</sup> y <sup>-1</sup>	Grams wet mass km <sup>-2</sup> y <sup>-1</sup>
Modelling purpose	Identify data inconsistencies. Investigate the trophic roles of krill and copepods. Explore future scenarios of reduced krill abundance.	Describe dynamics of monitored and declining species. Evaluate how sea- ice cover explains variations in species biomasses.	Describe food web structure excluding current commercial fisheries.	Guide decision- making around ecosystem management. Separate models generated to represent ecosystem state in 1960s, 1908s and 2000s	Expansion of existing model (Subramania m et al. 2019) to include Heard and McDonald Islands. Provide an overview of ecosystem structure for the entire plateau including fisheries.	Identify energy pathways through mesopelagic groups (fish, krill, squid). Identify keystone species. Explore the ecosystem implications of future climate scenarios.
Modelling period	2000-2010	Nominally 1996, with biomass data from 1992-2002	1990-2000	Various, but 2000s model used here	1990s-2000s	Period around 2016
N functional groups	31	35	38	37	28	28
N baleen whale groups	1	4	2	0	1	2
Published model versions	Parameters: Balanced and unbalanced Diets: Balanced and unbalanced	Parameters: Balanced Diets: Balanced and unbalanced	Parameters: Balanced and unbalanced Diets: Balanced and unbalanced	Parameters: unbalanced Diets: Balanced and unbalanced	Parameters: Balanced and unbalanced Diets: Balanced	Parameters: Balanced and unbalanced Diets: Balanced and unbalanced
Degree of system closure	External feeding by some predators represented by additional functional groups	Closed system	Export of production represented for migratory mammals	Closed system	Closed system	Closed system

#### 2136 5.2.3 Model standardisation

2137 A key issue when using multiple models from disparate sources is that many of the

assumptions and decisions made when compiling the associated data and structuring the

- 2139 models are dependent on the objectives of the study and preferences of the authors. This
- 2140 means that models are not necessarily directly comparable, as they may include differences in
- 2141 the number of functional groups and levels of functional group aggregation and in the
- 2142 approach taken to estimate energetic parameters, which may influence their outputs (Pinnegar
- 2143 et al. 2005; Heymans et al. 2016). Standardising the energetic parameters and the number and
- 2144 identity of functional groups between models is therefore a key step in comparing different
- models (Hill et al. 2021). The following subsections describe how model standardisation wasconducted in this study.

#### 2147 Standardisation of units

Five of the selected models expressed biomass in units of wet mass (gWM m<sup>-2</sup>y<sup>-1</sup>) while one (RS) used units of organic carbon (gC m<sup>-2</sup>y<sup>-1</sup>). To standardize the model units, we converted the RS model to gWM m<sup>-2</sup>y<sup>-1</sup> using literature-derived conversion factors for the major functional groups (Table D1). The biomass of each group was scaled using the relevant conversion factor, while *P/B* is unitless and therefore does not need conversion. *Q/B* was converted following the equation:

2154 
$$\frac{Q}{B_{W,i}} = \frac{\sum_{z=1}^{n} Q_{C,i,z} / CF_z}{B_{C,i} / CF_i}$$

where  $Q/B_{W,i}$  represents the Q/B of group i in wet mass,  $Q_{C,i,z}$  represents the consumption of prey z by predator i in carbon mass,  $B_{C,i}$  represents the biomass of predator i in carbon mass, and  $CF_i$  and  $CF_z$  are the conversion factors for the predator and prey, respectively.

The diet composition of each predator was converted from proportion carbon to proportionwet weight of each prey consumed using the equation:

2160 
$$D_{W,i,z} = \frac{Q_{C,i,z}/CF_z}{\sum_{z=1}^n Q_{C,i,z}/CF_z}$$

2161 whereby  $D_{W,i,z}$  is the proportional contribution in wet weight units of prey z to the diet of

- 2162 predator *i*. The diet of each predator is expressed in terms of consumption  $(Q_{W,i,z})$  by
- 2163 multiplying the diet matrix  $(D_{W,i,z})$  by the consumption parameter Q of the predator:

2164 
$$Q_{W,i,z} = D_{W,i,z} \times \frac{Q}{B_{W,i}} \times B_{W,i}$$

#### 2165 *Re-aggregation of functional groups*

2166 Where possible, we combined existing groups in each model to generate a subset of 2167 comparable groups, but in some cases, groups were disaggregated to ensure that our new set 2168 of models all explicitly represented the same key groups (see Appendix D1 for a detailed 2169 description of the reaggregation steps applied to each functional group). Our final 2170 aggregation scheme included 21 functional groups overall, though not all of these were 2171 included in each model (Table D2). We retained a few regional contrasts representing genuine 2172 ecological differences in the groups present between models (e.g. presence of sea ice algae in 2173 high-latitude models, absence of Antarctic krill in low-latitude eastern Antarctic models). 2174 Four of the original models explicitly included bacterial groups and we retained these but did 2175 not add bacterial groups to models which did not already include them, to avoid introducing 2176 further subjectivity through our decisions surrounding the parameterisation of this group.

2177 Diets were reaggregated following Hill et al. (2021) by expressing the diet of each consumer 2178 in terms of their reaggregated prey groups, calculating a consumption-weighted average for 2179 each prey item across the constituent consumers, and then rescaling the resulting values to 2180 sum to one. We opted to use the balanced diet matrices for this as, in some cases, the 2181 unbalanced matrices were missing important prey groups from the diets of some consumers, 2182 which caused unwanted behaviour during the later balancing process. The SG model 2183 incorporated off-shelf (beyond the model boundary) feeding by some groups, which we 2184 removed to improve standardisation between models. To do so, we redistributed the diet 2185 composition of affected consumers across their relevant on-shelf prey groups. We also 2186 reduced the biomass of these consumers appropriately to reflect the reduction in available 2187 energy from solely on-shelf feeding. As a result, our model for SG represents the ecosystem 2188 structure, including top predator populations, that can be supported by on-shelf feeding alone. 2189 The published RS model incorporates export of migratory mammal production, representing 2190 emigration of individuals, but for the purposes of consistency with the other models we 2191 omitted these export parameters to model this as a closed system.

# 2192 Representation of baleen whales

We modelled baleen whales as three functional groups: humpback whales, minke whales and"other baleen whales". These groups were determined primarily based on their diets as,

2195 across models, humpbacks and minkes consumed a greater proportion of fish compared to the 2196 remaining whales which are more dependent on krill and other zooplankton. Humpback and 2197 minke whales also display some ecological differences in foraging, the former preferring 2198 open-ocean regions and the latter often feeding in the sea-ice zone (Bombosch et al. 2014), 2199 therefore these groups were kept separate. One model (PE) did not include any baleen 2200 whales, and another (KP) only included southern right whales and fin whales. In both cases, 2201 the exclusion of other whale species was due to their low contemporary occurrence in the 2202 model regions (Gurney et al. 2014; Subramaniam et al. 2020). The missing whale groups 2203 were added to these models but represented initially with negligible biomass to minimise the 2204 impact of this addition on the structure of the initial models. The addition of new whale 2205 groups to these models required the estimation of relevant diet matrices. To ensure that the 2206 initial consumptive impact of whale groups was comparable between models, we averaged 2207 the diets of each baleen whale group across models where such diet information was 2208 available and used the resulting averages as inputs across all models. The only exception to 2209 this was our treatment of krill. Antarctic krill were absent from two models (PE and KP -2210 these only included 'other krill'), which meant that simply applying an average whale diet 2211 across models was not appropriate. Instead, to apply a consistent approach to diet 2212 standardisation which accounted for fundamental differences in prey distributions, we 2213 combined Antarctic krill and 'other krill' into a single group in each model, conducted our 2214 averaging, and then split these groups apart in proportion to their relative biomass in each 2215 model. This meant that the overall proportion of krill in whale diets was the same across 2216 models, but the relative proportion of each krill group varied based on their underlying 2217 biomass estimates for the relevant model regions (Table D3).

Three of the original models included fishery takes: the KP model incorporated icefish and toothfish catches, the PE model included toothfish fishery, and the AP model incorporated krill fishery landings. The SG model does not incorporate fishery removals despite there being commercial catches of fish and krill around the shelf, as these removals are estimated to represent only 1% and 7% of input production estimates for these groups (Hill et al. 2012). For consistency, we excluded all fisheries catches and by-catch from models.

2224 Standardisation of rate parameters

To standardise the input energetic parameters (P/B and Q/B) of each aggregate group, we averaged the values for each relevant reaggregated functional group across all models for

- which an independent estimate was available. Where possible, we used the unbalanced
- 2228 parameter estimates for these calculations, to minimise the potential influence of changes to
- these input parameters made by the respective model authors during the balancing process.
- 2230 The exception to this was the AP model for which only balanced biomass and rate parameter
- 2231 estimates were available.

# 2232 5.2.4 Catch-derived estimates of plausible whale biomass

We used catch records from the International Whaling Commission (IWC) to generate region-specific upper estimates of plausible biomass for each whale group (henceforth 'limit biomass'). These data provide information on species and length of whales caught during the 20th century, at spatial resolutions ranging from the nearest degree to the nearest minute or only approximate to the nearest 5- or 10-degree grid cell, depending on the expedition.

2238 To estimate an upper limit to the total biomass of whales in each model, we aggregated 2239 catches for each whale group within a 1000km buffer around each study region (Figure D1). 2240 This assumes that all individuals within this buffer are capable of spending time in the model 2241 area, which is a reasonable assumption for such wide-ranging animals. To estimate the 2242 biomass of each whale group, we first converted length records to mass using published 2243 length-weight relationships for each species (Lockyer 1976). We assumed that each whale 2244 group spends only part of the year feeding in each region, given their migratory nature. Some 2245 models (RS, PB and KP) already provide estimates of residence time, while for the others we 2246 assumed a 90-day feeding period in the region which, according to Savoca et al. (2021), 2247 represents the lower limit of the most likely annual feeding period for individuals. By 2248 combining the total biomass, residence time and the spatial area of the model regions, we 2249 estimated the annual biomass per unit area for each whale group in each model. By adding 2250 the contemporary values from the models, we identified an upper estimate of plausible pre-2251 exploitation whale biomass for each model region. These steps are illustrated in the following 2252 equation:

2253 
$$B_{pre,i} = \frac{\sum_{i=1}^{n} B_{catch,i} \times t_i}{area} + B_{current,i}$$

2254 Where  $B_{pre,i}$  is the upper estimate of plausible pre-exploitation annual biomass density 2255 (tonnes) for baleen whale group *i*,  $B_{catch,i}$  is the biomass (tonnes) of group *i* caught in the 2256 1000km buffer,  $t_i$  is the proportion of the year that whale group *i* spends in the model region, 2257 *area* is the area of the model in  $\text{km}^2$ , and  $B_{current,i}$  is the biomass (tonnes) per unit area of 2258 whale group *i* already estimated for the published model.

# 2259 5.2.5 Estimates of baleen whale Q/B

2260 We calculated a range of Q/B estimates for each whale group using the information contained 2261 in Savoca et al. (2021). For the minimum estimates, we used the constants and metabolic 2262 exponents from previous studies, provided in Savoca et al. (2021) (Table D4), combined with 2263 average body masses for each whale group from Greenspoon et al. (2023) to calculate an 2264 average Q/B for each whale group, assuming a 90-day feeding period (applied to all model 2265 regions for consistency) (Table D5). We opted to estimate these using the 'prior' parameter 2266 values in Savoca et al. (2021) because they represent a consistent approach which establishes 2267 a lower bound on whale consumption rates. These Q/B values will henceforth be referred to

- 2268 as the 'baseline' Q/B values.
- We then used the lower, median and upper estimates of daily rations estimated by Savoca et al. (2021) combined with the average whale body masses, again assuming a 90-day feeding period, to calculate higher O/B values for later perturbation analyses (Table D5).

#### 2272 5.2.6 Model balancing

2273 To standardise the balancing process across regional models and to explore the effects of 2274 perturbation across a range of plausible alternative model parameterisations, we developed a 2275 bespoke balancing algorithm and used this to generate 1000 versions of each regional model. 2276 The algorithm employs an automated iterative stepwise approach to optimise the set of 2277 parameter values to achieve balance, and the method is explained in detail in Appendix D1, 2278 with a general overview provided here. The algorithm randomly varies B, P/B, Q/B, GE and 2279 DC to search the parameter space for parameter combinations that satisfy balance criteria (all 2280 EEs equal to or below 1), while ensuring that values remain within defined confidence 2281 intervals (Table D6). One exception is that the whale Q/B values were fixed to the baseline 2282 estimates calculated for each group, to ensure that the initial consumptive impact of whales 2283 per unit biomass was standardised across models. Assimilation efficiencies were fixed for all 2284 groups, using values obtained from Pinkerton and Bradford-Grieve (2010). At each step, the 2285 new parameter set is evaluated using an objective function (the sum of EE for all groups out 2286 of balance). A record of the 'best' model (lowest objective function) is updated throughout, 2287 until either a balanced parameter set is identified, or the algorithm has reached a specified 2288 number of steps (we set this to 2000 but more can be used, although this increases runtime).

- In the latter case, the algorithm then switches to targeted, generally small, adjustments to
- biomasses and diets to nudge the model towards balance, much like the approach of
- 2291 Kavanagh et al. (2004). These adjustments are focussed on the group most out of balance,
- and their most influential predator (i.e. the one with the greatest consumption of the prev
- group). Once a balanced model is found, the biomass values for each group are checked, and
- the model is rejected if any biomass values fall outside their specified confidence intervals.
- In some cases, the balancing process resulted in skews in the distribution of some balanced parameters across model runs, particularly for P/B (often right-skewed) and Q/B (often leftskewed) (Figure D2-D7). These skews were generally consistent between regional models, suggesting no obvious model bias. There were also some differences in the distribution of EE
- 2299 values across groups, but these were not consistent between models (Figure D8-D9).

# 2300 5.2.7 Perturbation scenarios

- 2301 Our perturbation scenarios explored the effects of increasing whale prev consumption (O) on 2302 either competitor biomasses or on system-level primary productivity demand. As Q is simply 2303 the combination of B and Q/B, it was possible to set an upper plausible limit on Q by 2304 combining the whale limit biomass with the upper Q/B estimate from Savoca et al. (2021). 2305 This range of Q values encompasses all plausible scenarios of increased biomass and revised 2306 understanding of consumption rates. This also meant that we could convert any given Q into 2307 a different B based on the relevant Q/B estimate we wanted to apply, allowing us to determine 2308 the effect of the revised daily consumption rate estimates on the capacity to support more 2309 whales.
- 2310 Scenario 1: Compensatory changes in competitor populations

2311 In this scenario, we explored the compensatory decreases in the aggregate biomass of key 2312 higher-trophic competitors (marine mammals, birds, fish, and squid) required to facilitate 2313 increases in baleen whale Q up to the plausible limit without changes to lower trophic levels. 2314 We also identified the Q that could be supported by each model at a specific threshold of total 2315 competitor biomass. For this we used a threshold of 75% of initial competitor biomass, as this 2316 has previously been identified as a suitable boundary for the ecosystem-based management of 2317 the Southern Ocean (Watters et al. 2013). To implement this scenario, we increased the whale 2318 Q in each model version in a stepwise manner for a range of values up to the maximum 2319 suggested by the relevant regional limit biomass and the upper Q/B estimates. At each step in 2320 the process, any prey groups that were pushed out of balance (EE > 1) were identified. The

biomasses of the competitor groups that fed on these prey groups were then reduced by small
amounts in proportion to their predatory impact until balance was achieved. This process
continued until the plausible limit on baleen whale Q was reached or until the biomass of all
competitor groups had been reduced to below one percent of their starting values.

2325 To investigate the possible drivers of any variation in the capacity of regional models to 2326 sustain increases in whale Q, we calculated two different metrics. The first was the 2327 production-weighted average EE across all whale prey groups, which describes the spare 2328 capacity that each model has to support further consumption by whales without requiring 2329 compensatory changes to competitor biomasses. The second was a modified version of the 2330 Schoener dietary overlap index (Schoener 1970), using the normalised summed consumption 2331 of each prey group by baleen whales compared to that of their competitors, to capture the 2332 degree of competition between baleen whales and their competitors:

2333 
$$C = 1 - \frac{1}{2} (\sum |P_{x,i} - P_{y,i}|)$$

where *C* is the Schoener index,  $P_{xi}$  is the consumption of diet item *i* as a proportion of total prey consumption by group *x* (all baleen whales), and  $P_{y,i}$  is the consumption of diet item *i* as a proportion of total prey consumption by group *y* (all whale competitors).

To identify how these metrics indicate the capacity of models to sustain increases in whale Q, we estimated the slope of the average relationship between competitor biomass proportion and whale Q across the 1000 runs within each regional model ensemble. For consistency between models and to capture the most linear part of the relationship we used the section between 80% and 20% of competitor biomass. We used a Pearson's correlation to identify the association between the slope of each model average and the average *EE* or Schoener index across each of the regional model ensembles.

# 2344 Scenario 2: Lower-trophic changes required to support baleen whale recovery and2345 competitor populations

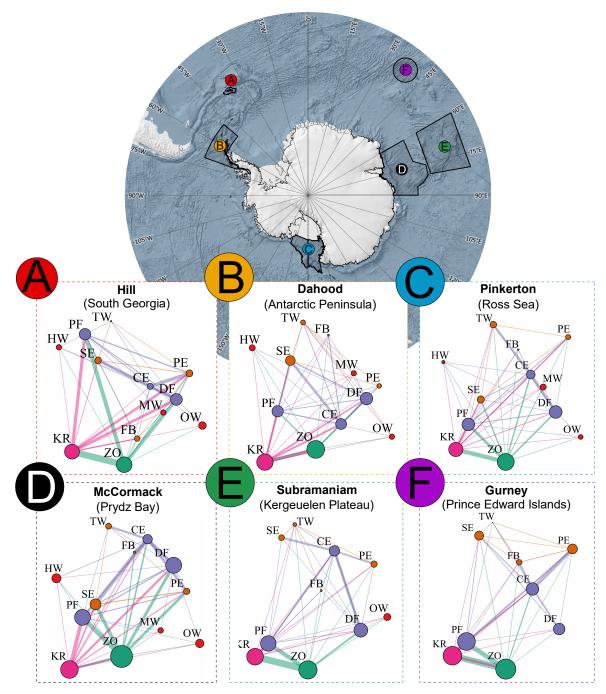
This scenario followed a similar approach to the previous one, but this time the biomasses of higher trophic level competitors (marine mammals and birds) were fixed, and the biomasses of baleen whale prey and other lower trophic level groups were increased if their EE rose above 1, to compensate for higher predation. We focussed on the relative change in primary production required, as a measure of the total system productivity needed to support these

2351 potential future ecosystems.

# 2352 *5.3 Results*

# 2353 5.3.1 Initial balanced model ensembles

- 2354 There was some regional variation in the average consumptive flows for the baleen whale
- 2355 prey and competitor groups. Krill made up a large proportion of total flows in the SG, AP, PB
- and RS models, while other zooplankton contributed more to total consumptive flows in the
- 2357 PE and KP models (Figure 5.2). The sources of consumptive flows to higher predators
- 2358 (marine mammals and seabirds) were quite varied in the SG, AP, PB and RS models, but
- were dominated by flows from squid and fish in the PE and KP models (Figure 5.2).



2360

2361 Figure 5.2: Spatial distribution of the models. Inset network diagrams display the log-2362 transformed consumptive flows between whales, their prey groups and their main 2363 competitors, averaged across model ensembles. Node and link size are proportional to 2364 biomass and consumption, respectively (not comparable between panels). Nodes are arranged 2365 evenly aong the vertical axis by rank order of their trophic level, coloured by the main 2366 groupings (red: baleen whales, orange: marine mammals and seabirds, purple: fish and squid, pink: krill, green: zooplankton). TW = toothed whales; HW = humpback whales; MW = 2367 minke whales; OW = other baleen whales; SE = seals; PE = penguins; FB = flying birds; PF 2368 = pelagic fish; DF = demersal fish; CE = cephalopods; KR = krill; ZO = zooplankton. 2369

- 2370 When comparing the models based on the combination of their production-weighted average
- EEs and Schoener index, there appeared to be three primary groupings of models: Group 1
- 2372 (PB, PE and RS) had low Schoener index and high *EE*; Group 2 (SG and KP) had high
- 2373 Schoener index and low *EE*; Group 3 (AP) had a low Schoener index and low *EE* (Figure 5.3;
- 2374 Table D5).

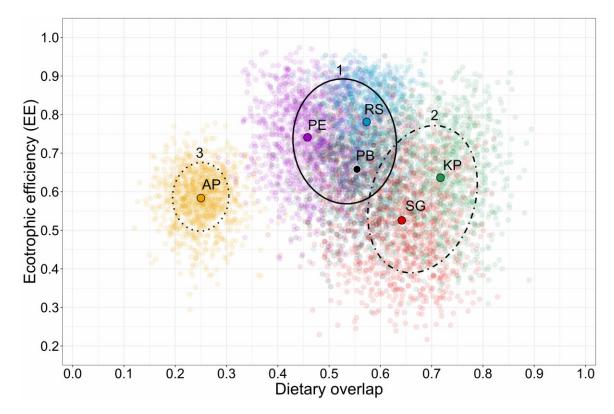




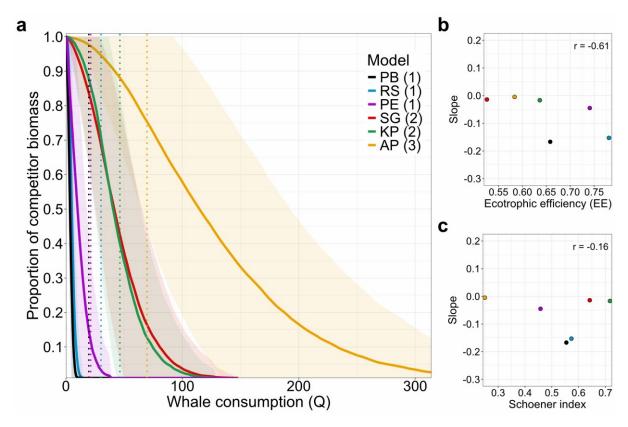
Figure 5.3: Relationship between production-weighted average ecotrophic efficiency (*EE*) of all baleen whale prey groups and the overall dietary overlap between baleen whales and their competitors. Points are coloured by model region, with small points indicating the position of individual model runs and larger points representing their average. Ellipses represent one standard deviation around the three qualitative groupings, identified by numbers on the plot. The initial balanced PE model included no whale biomass therefore the dietary overlap was calculated by adding a negligible biomass to the model.

2383 5.3.2 Catch-derived total whale biomass

There was considerable variation in the upper bound on plausible biomass for baleen whales in each model region. The SG model had the largest value (75.45 t km<sup>-2</sup> y<sup>-1</sup>), more than twenty times higher than that of the other models, which ranged between 1.04 and 3.62 t km<sup>-2</sup> y<sup>-1</sup> (Table D6). This also represented the greatest proportional increase compared to initial whale biomass amongst the six models (262 times compared to between 18 and 80 times in

- the other models) (Table D7). There were also changes in the group composition of baleen
- whale biomass between the initial unbalanced published model inputs and in the catch-
- 2391 derived limit biomass for the corresponding regions. In particular, humpback and minke
- whales made up the majority of baleen whale biomass in the initial inputs of the AP, PB and
- 2393 RS models, but 'Other baleen whales' dominated the catch in all models (Figure D10).
- 2394 *5.3.3 Perturbation scenarios*
- 2395 Scenario 1: Compensatory changes in competitor populations

2396 There were large differences in the capacity of each regional model to support increased 2397 whale consumption (Q). The model groupings identified based on the combination of their 2398 averaged EE and Schoener index (Figure 5.3) were clearly linked to differences in the 2399 average slopes of the relationship between competitor biomass and whale Q. The models with 2400 low Schoener index and high EE (PB, RS, PE) experienced the most rapid decrease in 2401 relative competitor biomass with increasing whale O, followed by those with high Schoener 2402 index and low EE (KP, SG), while the AP (low Schoener index, low EE) was able to support 2403 large whale Q increases while also retaining competitor biomass (Figure 5.4a). Of these two 2404 parameters, there was a stronger correlation between *EE* and the slope of competitor biomass 2405 (r = -0.61) than for the Schoener index (r = -0.16) (Figure 5.4b & c). It should be noted that 2406 neither of these correlations were significant (p>0.05), likely due to the small sample size (*n* 2407 = 6 models).

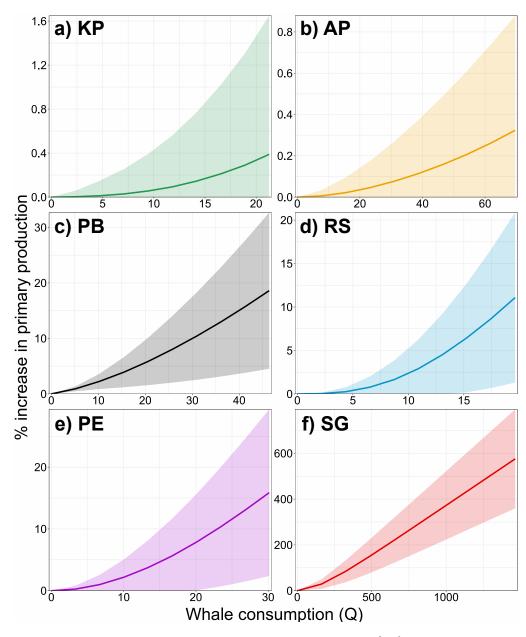


2409 Figure 5.4: a) Average relationship between competitor biomass proportion and whale consumption ( $Q t km^2 v^{-1}$ ) for each of the model ensembles. Solid lines indicate model 2410 2411 averages, shading indicates standard deviation. Vertical dashed lines identify the upper whale 2412 Q identified for each of the model regions (not shown for the SG model because this was 2413 beyond the x axis scale). Numbers in brackets represent the groupings of the models by the 2414 combination of their production-weighted ecotrophic efficiency (*EE*) and Schoener index; b) 2415 association between the slope of each model average in panel a) and the production-weighted 2416 mean EE of baleen whale prey; c) association between the slope of each model average in 2417 panel a) and the Schoener diet overlap index between baleen whales and their competitors.

2408

Across models, the use of different Q/B estimates had a large impact on the increases in 2418 2419 whale biomass that were possible. Under the baseline Q/B estimates, and while conserving 2420 the majority (99%) of competitor biomass, baleen whale biomass could be increased between 2421 1.52x (PB) and 159.35x (KP) (Figure D11, Table D8). If conserving only 75% of competitor 2422 biomass, the average increases in baleen whale biomass were substantially higher, ranging 2423 from 12.40x in the PB model to 865.62x in the KP model (Figure D11, Table D8). Using the 2424 median Q/B estimates based on Savoca et al. (2021), on average only two of the models (KP 2425 and RS) were capable of supporting any additional whale biomass while maintaining 2426 competitor biomass at above 99% of initial values (Figure D11, Table D9). At the competitor

- biomass threshold of 75% of starting values, the relative increases in baleen whale biomass
- 2428 were far more conserved than under the baseline Q/B estimates, ranging from 1.17x (PB) to
- 2429 79.92x (KP) on average (Figure D11, Table D9). Using the upper *Q/B* estimates, the
- 2430 possibility for baleen whale biomass increases was even more restricted (Table D9). Within
- 2431 each model ensemble, there was high variability in competitor responses across model runs
- 2432 (Figure D12).
- 2433 Increases in whale *Q* had different impacts on the biomasses of individual competitor groups
- 2434 in each model. Of the higher trophic competitors, seals experienced particularly rapid
- 2435 biomass declines in the AP and PB models, while penguins were especially impacted in the
- 2436 SG and RS models (Figure D12). Within the lower trophic competitors, the biomass of
- 2437 demersal fish declined particularly rapidly in the SG, PB, RS and AP models, while, in
- 2438 contrast, pelagic fish were the most strongly affected across all competitor groups in the PE
- and KP models (Figure D12).
- 2440 Scenario 2: Lower-trophic changes required to support baleen whale recovery and2441 competitor populations
- 2442 The relative increases in primary productivity required to support the limit baleen whale
- consumption were relatively modest in the AP and KP models (0.32% and 0.39%,
- respectively; Figure 5.5a & b), but considerably larger in the RS, PB and PE models (ranging
- from 11.08% to 18.61%) (Figure 5.5c-e; Table D10). The SG model required extremely large
- 2446 increases in primary production (575.97%; Figure 5.5f). Based on a linear estimation of the
- slope of the average relationship between whale Q and primary production in each model, we
- estimated that a unit increase in whale Q ( $t km^2 y^{-1}$ ) required relative increases in primary
- production ranging from 0.005% (AP) to 0.565% (RS) (Table D11).



#### 2450

Figure 5.5: Relative changes in annual primary production  $(t \ km^2 y^{-1})$  required to support increases in total whale consumption  $(Q \ t \ km^2 y^{-1})$  if lower trophic level biomasses are increased to support increases in whale Q while competitor biomasses are fixed. Solid lines indicate model averages, shading indicates standard deviation. The maximum value on the xaxis represents the upper bound on plausible baleen whale consumption (Q). Note varying axis scales.

## 2457 *5.4 Discussion*

Commercial whaling during the 19<sup>th</sup> and 20<sup>th</sup> centuries drove massive declines in baleen
whale populations within the Southern Ocean, and thus impacted wider ecosystem structure

2460 (Christensen 2006; Mori and Butterworth 2006). The recovery of these populations could

therefore have major impacts on the population trajectories of other important competitors
such as seals, penguins and fish. In this study we identified the compensatory changes to
competitor populations that would be necessary to support increases in whale consumption
rates under contemporary conditions, and the bottom-up changes necessary to simultaneously
achieve baleen whale population recovery and conservation of current competitor biomass.
Our results highlight the importance of taking a holistic approach to the management and
conservation of different competing populations.

#### 2468 Scenario 1: compensatory changes to competitor populations

2469 We found that contemporary regional Southern Ocean food webs have some capacity to 2470 support increased baleen whale populations, but this was dependent on daily prey 2471 consumption estimates and may come at the expense of competitor populations. Previous 2472 modelling has also found increases in global baleen whale populations to have strong indirect 2473 impacts on populations of other higher trophic groups including seals and seabirds (Ruzicka 2474 et al. 2013). Five-fold increases in consumption by baleen whales were found to cause 2475 declines in top predator production of up to 29%, and twenty-fold increases in baleen whale 2476 consumption required significant (up to 73%) reductions in production by competitor 2477 populations (Ruzicka et al. 2013). Under more recent estimates of higher baleen whale daily 2478 consumption rates (Savoca et al. 2021), the competitive impact of whales may be elevated. 2479 This is because values of O/B (the annual prev consumption per unit biomass) determine the 2480 relationship between O (consumption) and B (biomass), with a given O value representing 2481 ever smaller B as Q/B increases. In consequence, at high Q/B values, the whale Q thresholds 2482 at which competitor populations begin to decline will be reached at comparatively lower 2483 whale B than at low Q/B values. We found that, when using revised Q/B values based on 2484 median daily consumption estimates in Savoca et al. (2021) and conserving 75% of total 2485 competitor biomass (representing changes that are potentially reversible within two or three 2486 decades; CCAMLR 2008; Watters et al. 2013), increases in whale biomass were highly 2487 constrained (only around 9% of the values possible under baseline Q/B estimates; Table D9). 2488 In four of the six models this represented only a fraction (from 2.5% to 26%) of the plausible 2489 limit biomass, although the remaining two models (AP and KP) could still sustain the full 2490 upper bound on whale biomass. Under higher prey consumption estimates the capacity of 2491 food webs to support larger whale populations was even more constrained (Table D9). 2492 Overall, these results support the conclusions of Ruzicka et al. (2013) that whale recovery 2493 will have large impacts on competitor populations. It should be noted that there are various

other factors that might modify these relationships, such as the degree of flexibility in
competitor diets and the competitive hierarchies between competitors and whales, which are
discussed further in a later section.

2497 While our use of ensemble averages suggests a smooth and relatively consistent relationship 2498 between whale Q and competitor biomass, individual model runs displayed much more 2499 sudden inflection points along the Q scale, leading to rapid competitor biomass declines 2500 (Figure D12). Most models exhibited a phase of no change in competitor biomass, likely due 2501 to spare consumptive capacity of prey groups (i.e. low EE) which provided an initial buffer 2502 against the increases in whale Q. This was generally followed by a phase of gradual 2503 competitor biomass decline as the spare consumptive capacity of whale prey is used up and 2504 biomass adjustments are spread across multiple competitor groups based on their relative O/B2505 and degree of dietary overlap with whales. Finally, most model runs displayed a sudden 2506 changepoint after which competitor biomasses rapidly declined to zero. As the individual 2507 competitor populations are reduced, their overall consumptive impact on a given prey relative 2508 to the total consumption of that prey by other groups becomes smaller, and therefore 2509 relatively greater reductions in biomass are required to counteract the increased consumption 2510 by baleen whales. Individual competitor populations will eventually become so small they 2511 cannot be further adjusted, and the remaining competitor groups then must be subject to 2512 relatively greater biomass adjustments to account for this. This is compounded by the fact 2513 that the last remaining competitor groups are likely to be those with relatively low Q/B (as 2514 they will have had the least impact on whale prey groups and will therefore have been 2515 reduced the least in the previous phase), and therefore each unit of Q removed to balance the 2516 impact of more whales will represent a relatively greater biomass. It would be worth 2517 investigating whether the drivers of these abrupt transitions in the models actually exist in 2518 nature. It is also likely that other factors such as competitive hierarchies, dietary flexibility 2519 and population depensation would modify these relationships by determining the relative 2520 impacts of increased consumption by whales on each competitor group (Barlow et al. 2002; 2521 Abrams 2010; Liermann and Hilborn 2001), and their effects should be investigated.

# 2522 Scenario 2: Bottom-up changes to support baleen whales and competitors

2523 There was considerable variation in the total primary production required to support the limit

- whale biomass and contemporary competitor populations in each model. In particular, the SG
- 2525 model needed extremely large increases in primary production (>500%), likely because the

2526 extensive whale catches that occurred within the 1000km buffer, coupled with the small 2527 modelled spatial area, generated very high estimates of biomass per unit area (Figure D1). 2528 The shelf waters around South Georgia are highly productive and considered a biodiversity 2529 hotspot (Hogg et al. 2011), which may have driven some of the highest densities of baleen 2530 whales on the planet (Richardson et al. 2012). However, some of the models with 2531 comparatively small plausible upper whale biomass estimates (e.g. RS, PB and PE) also 2532 needed a substantial rise in primary production (up to 19%) to support whale population 2533 increases without adverse effects on competitors. These models also displayed the steepest 2534 competitor biomass declines in Scenario 1, highlighting the link between apparent sensitivity 2535 to whale recovery and the magnitude of changes in bottom-up forcing needed to mitigate negative impacts. 2536

2537 The role of baleen whales in nutrient cycling in the ocean is still poorly understood, and 2538 likely to vary by geographical location, ecosystem productivity and whale community 2539 composition (Gilbert et al. 2023). In nutrient-limited sub-tropical calving grounds, 2540 unexploited blue whale populations increased primary productivity by up to 15% compared 2541 to the average for subtropical waters (Roman et al. 2014). In the Southern Ocean, where trace 2542 metals such as iron are the primary limiting factor for phytoplankton growth the iron excreted 2543 by unexploited whale populations could represent around 12% of the average contemporary 2544 iron content in Southern Ocean surface waters (Ratnarajah et al. 2014; Nicol et al. 2010). 2545 This could have stimulated higher levels of primary productivity, possibly up to 11% greater 2546 across the Southern Ocean under high whale consumption rate estimates (Savoca et al. 2021). 2547 This value is at the lower end of the increases we estimated to be required to support the 2548 upper plausible bound on whale consumption in the three models that were most sensitive to 2549 baleen whale population recovery (PB, RS and PE), and far below the values required by the 2550 SG model. Our results therefore suggest that, across the Southern Ocean, whale-driven 2551 nutrient cycling alone is unlikely to be sufficient to fully mitigate impacts of whale recovery 2552 on other higher trophic levels.

Predicted future changes in primary production are highly uncertain, and there will likely be
considerable regional variation in trends. Overall, increases in temperature and irradiance,
combined with shallowing of mixed layer depths, are expected to drive increased primary
productivity across the Southern Ocean (Henley et al. 2020; Kaufman et al. 2017). Increases
in net primary production may be 50% or more in some high latitude regions by the end of
the century, with more modest increases (up to 30%) elsewhere (Steinacher et al. 2010; Fu et

2559 al. 2016; Fisher et al. 2024), although a study on the Ross Sea predicted more limited 2560 increases in primary productivity of 5% by 2050, and up to 14% by 2100 (Kaufman et al. 2561 2017). Overall, it seems plausible that future increases in primary production will be 2562 sufficient to mitigate at least some of the negative impacts of baleen whale population 2563 recovery identified in our models (except SG), particularly if there is an additive effect of 2564 elevated whale nutrient-cycling. This will, however, also depend on the nature of potential 2565 changes in primary producer composition as this will determine their suitability for sustaining 2566 whale prey populations (discussed further below).

#### 2567 Regional differences

2568 There were regional differences in model capacity to support increases in baleen whale Q. 2569 These distinctions could result from both arbitrary differences between models (so-called 2570 model 'personality') and true ecological differences (Hill et al. 2021). While it is beyond the 2571 scope of this study to definitively distinguish the relative influence of each of these factors, 2572 we made significant efforts to ensure the comparability of the models used. By standardising 2573 functional group composition and energetic parameters, we removed some of the possible 2574 inter-model variation that might result from decisions made during the original model 2575 construction process. Additionally, our use of a novel automated balancing algorithm 2576 facilitated the construction of model sets encompassing a range of plausible balanced 2577 parameter sets for each regional model, thus allowing us to explore uncertainty around model 2578 structure. By analysing a suite of runs from a variety of regional models, we were able to 2579 explore a broad set of ecologically viable scenarios representing the multitude of potential 2580 responses that Southern Ocean ecosystems might display in future.

2581 Models were grouped based on their combination of average whale prey *EE* and whale-2582 competitor diet overlap (Schoener's index), and these groupings matched the general order of 2583 steepness of the model average slopes of competitor biomass decline with increasing whale Q 2584 (Scenario 1). A group's *EE* inversely relates to the spare production available for additional 2585 consumption, therefore low values suggest a greater capacity for increasing consumption 2586 without pushing the prey group out of balance, and vice-versa (Christensen and Walters 2587 2004). As a consequence, the model runs with higher average prey *EE* generally had less 2588 capacity to support additional whale Q without compensation by competitors. This is because 2589 increases in Q promptly drive prey groups out of balance, resulting in compensatory declines 2590 in competitor biomass. Since a large proportion of model runs start with high prey *EE*s, the

inflection points at which competitor biomasses decline generally take place over a smallrange of whale *Q* values, leading to steeper average slopes.

2593 Whilst the *EE*s are dependent on the combination of model parameters adjusted during the 2594 balancing process, some of the differences in the distribution of *EE* values between regions 2595 could represent genuine ecological contrasts. The efficiencies of lower- and mid-trophic 2596 groups are often assumed to be close to one, suggesting that most of their production is 2597 directly utilized within the modelled ecosystem (Heymans et al. 2016). However, while 2598 predation mortality is a major driver of lower-trophic population dynamics, other sources of 2599 mortality (e.g. environmental conditions and food availability) can be considerable for 2600 organisms such as zooplankton, and advection by currents could reduce the proportion of 2601 production that is consumed by predators within the system (Tang et al. 2014; Hirst and 2602 Kiørboe 2002). It is therefore plausible for lower EE values to exist for these groups, and 2603 differences between models could reflect the influence of these local processes. Additionally, model parameters are based on data collected over multiple years and forced to meet 2604 2605 equilibrium assumptions and may encompass considerable variability in production at lower 2606 trophic levels (Plagányi and Butterworth 2004). Temporal variability in local abundances 2607 generally declines with trophic level (Siqueira et al. 2024), so populations of higher trophic 2608 consumers with relatively low-fecundity, such as marine mammals and birds, are unlikely to 2609 respond rapidly enough to changes in lower-trophic biomass to fully exploit any temporary 2610 surpluses in production. As a result, EEs for some prey groups may display inter-annual 2611 variation which could differ significantly between regions.

2612 Given the strong correlation between *EE* and the apparent capacity of models to cope with 2613 sustained press perturbations like whale population increases (Figure 4b & c), an improved 2614 understanding of the processes underlying the local sources of mortality of key prey groups 2615 would aid managers in targeting management and conservation actions. Gaining this 2616 understanding will require the integration of biological and physical data across relatively 2617 fine spatial but broad temporal scales. Identifying the processes driving natural mortality will 2618 also be key to predicting possible climate-driven changes in population dynamics and would 2619 aid our understanding of how ecosystems will change in future (Plagányi et al. 2022).

2620 The distinction between two model groupings (SG and KP vs AP) was largely due to the

2621 degree of dietary overlap between whales and their competitors. We adjusted competitor

2622 biomasses in proportion to their relative importance as consumers of each unbalanced prey

2623 group until balance was achieved and, as a result, in models where prey consumption by 2624 whales and competitors is more similar (high overlap), a unit increase in whale Q will need to 2625 be compensated for by relatively greater reductions in competitor biomass. Differences in 2626 dietary overlap may be due to genuine ecological contrasts, as the relative importance of krill, 2627 other euphausiids and pelagic fish in the diets of higher predators varies geographically 2628 (McCormack et al. 2021b), and we observed contrasts in the distribution of consumptive 2629 flows across our balanced suite of models which appear to broadly match these trends. In 2630 consequence, different competitor groups were most impacted by increases in whale Q, with 2631 seals and penguins declining most rapidly in regions with high reliance on Antarctic krill and 2632 pelagic fish strongly impacted in regions which are more heavily dependent on other groups 2633 such as Thysanoessa macrura (Wallis et al. 2019). Further studies of regional similarities and 2634 differences in diets will better resolve the degree of dietary overlap between baleen whales 2635 and other higher-trophic groups, thereby providing greater insight into the likely capacity of 2636 ecosystems to cope with future population changes.

2637 It would be valuable to further explore the relative effects of EE, dietary overlap and other 2638 potential factors (e.g. competitor group identity, biomass distribution and Q/B) on model 2639 capacity, to develop robust metrics which can be used to predict model responses to change. 2640 This could be explored further using suites of smaller (simpler) models with systematic 2641 differences in their parameters, with the shapes of competitor biomass responses to whale Q 2642 increases investigated on an individual group and model basis rather than as an aggregate. 2643 Doing so might reveal interactive effects of different parameters on the overall model 2644 capacity to support whale population increases.

2645 Further considerations:

Our study represents an initial effort to explore the potential impacts of baleen whale population recoveries on ecosystem structure. There are several additional factors which could influence the relationship between the population trajectories of whales and their competitors.

The expansion of krill fisheries and establishment of new fisheries for groups like mesopelagic fishes could reduce their availability for predators (Meyer et al. 2020; Fjeld et al. 2023). Environmental changes such as warming, sea ice loss, and changes to the production and composition of phytoplankton communities, could also alter the production of key prey (e.g. krill) and modify energy flow to higher trophic levels (Kawaguchi et al. 2024; Swadling

et al. 2023; Thomalla et al. 2023). Behavioural changes in groups such as Antarctic krill
might alter the density and distribution of swarms (Kawaguchi et al. 2024), reducing feeding
opportunities and elevating competition between whales and other groups. These
anthropogenic and environmental changes could therefore impact the capacity of regional

2659 ecosystems to sustain whale population increases with minimal effects on competitors.

Some competitor groups may have more flexible diets than others, making them better able to mitigate the impacts associated with greater competitive pressure from larger whale populations. For example, groups such as crabeater seals and chinstrap penguins are highly dependent on krill while others such as Weddell seals, fur seals and gentoo penguins have quite generalist and flexible diets according to the available prey field (Wege et al. 2021;

2665 McMahon et al. 2019; Reisinger et al. 2018).

2666 Differences in the competitive ability of whales and competitor groups may also influence the 2667 trade-offs between whale consumption and competitor biomass. Groups that are higher in the 2668 competitive hierarchy, perhaps due to greater foraging ability (Barlow et al. 2002), may be 2669 less impacted by increasing baleen whale populations because they will be able to 2670 outcompete other groups and maintain necessary energy intake, while their competitors may 2671 experience rapid declines. Additionally, while we explored the effects of increased whale 2672 biomasses on other groups, competition can also have strong effects on whales themselves 2673 (Ruzicka et al. 2013). Previous modelling suggests that increased competition for Antarctic 2674 krill (between baleen whales and other groups, and between whale species themselves) could 2675 strongly limit whale population recoveries (Tulloch et al. 2019).

2676 It is also important to consider whether changes in primary production are associated with 2677 shifts in the composition of phytoplankton communities, as these may influence the capacity for future primary productivity to mitigate the ecosystem effects of whale recovery. Sea ice 2678 2679 changes, warming, and increases in stratification and irradiation, may result in large diatoms 2680 being replaced by smaller flagellates and other groups in regions such as the sea ice zone, 2681 while more open ocean regions could experience the opposite trend due to increasing iron 2682 flux and atmospheric cloudiness (Henley et al. 2020; Krumhardt et al. 2022). In regions 2683 which experience declining diatom abundance and concurrent increases in populations of 2684 smaller phytoplankton, the efficiency of energy flow to higher trophic levels may be reduced 2685 (Krumhardt et al. 2022; Hunt et al. 2021). Additionally, some of the drivers of increased 2686 primary productivity (loss of sea ice, increases in temperature) are expected to negatively

2687 impact groups such as Antarctic krill (Flores et al. 2012). The combination of warming and 2688 shifts in phytoplankton community composition towards smaller, less energetically valuable 2689 species may also increase the abundance of salps (*Salpa thompsoni*) which can outcompete 2690 krill, resulting in fundamental changes to food web structure that could negatively impact

2691 energy supply to higher trophic levels (Pauli et al. 2021; Pietzsch et al. 2023).

Overall, the capacity of Southern Ocean food webs to support both increased whale
populations and contemporary competitor biomass will depend on a variety of interacting
factors, and it would be valuable to include these in future modelling studies to explore the
full suite of uncertainty around the potential for, and consequences of, baleen whale
population recovery.

#### 2697 *Implications for management*

2698 Anthropogenic and environmental changes have altered Southern Ocean ecosystems, creating 2699 challenges for certain management and conservation goals. Here, we have shown that the full 2700 recovery of baleen whale populations in the Southern Ocean is likely to result in strong trade-2701 offs between conservation objectives. The absolute magnitude of compensatory changes in 2702 competitor populations, and the points along the pathway of whale recovery at which they 2703 will occur remain uncertain and are probably region-specific. We identified that the levels of 2704 unexploited production by whale prey and the degree of dietary overlap between whales and 2705 their competitors may play an important role in the capacity of food webs to sustain whale 2706 population increases with minimal wider ecosystem impacts. Additionally, baleen whale 2707 consumption rates strongly influence the relative impact of whale population increases on 2708 food webs. Future changes in primary productivity due to environmental drivers and whale-2709 driven nutrient cycling, could potentially mitigate the ecosystem impacts of whale recovery. 2710 Future efforts to better resolve these factors will improve our understanding of likely regional 2711 ecosystem responses and aid management decisions. Ultimately, policymakers seeking to 2712 implement management and conservation strategies (e.g. fishery catch limits and marine 2713 protected areas) will need to integrate information regarding both the inherent capacity of 2714 ecosystems to support whale population increases (i.e. model structure) and the likelihood of 2715 beneficial environmental changes (e.g. production and composition of phytoplankton 2716 communities).

### 2718 6 General discussion

2719 Food webs are the framework upon which much of modern ecological research is built, 2720 providing insight into many aspects of ecology ranging from the drivers of individual 2721 population dynamics to the broader patterning of biodiversity and the impacts of global 2722 change on ecosystem functioning (Layman et al. 2015). The Antarctic is a particularly 2723 important focus of food web research, as the historically stable conditions and the 2724 physiological adaptations and high stenothermy of many species make Southern Ocean 2725 ecosystems especially vulnerable to changes such as warming and sea ice loss (Queirós et al. 2726 2024). Given the central role that the Southern Ocean plays within the wider earth system, it 2727 is important that we improve our understanding of the structural properties of regional food 2728 webs and how these influence their resilience to environmental and ecological change 2729 (Murphy et al. 2012). This thesis explored some key aspects of the structure of Southern 2730 Ocean food webs and their responses to change. Chapters 2 and 3 used functional traits to 2731 explain trophic structure at different scales, from the distribution of feeding links at the 2732 community level to the organisation of a key stabilising substructure, modularity, at the level 2733 of entire food webs. Chapter 4 then investigated how temperature alters the size-structuring 2734 of trophic interactions, which is a key aspect of marine food webs. Finally, chapter 5 explored 2735 the range of possible responses of Southern Ocean ecosystems to a key ecological 2736 perturbation, the recovery of baleen whales. Below, I discuss the primary contributions of 2737 each chapter to our knowledge of food webs in the Southern Ocean and beyond and explore 2738 some of the further avenues which could be taken to improve our understanding of each 2739 topic. I then finish by summarising some of the major future directions that I see for food web 2740 research in general, based on the themes covered in this thesis.

2741 6.1 Chapter contributions

# 6.1.1 Chapter 2: Morphological traits distinguish feeding guilds in a Southern Ocean fishcommunity.

Chapter 2 determined the ecomorphology of demersal fish around South Georgia by first
classifying species and size classes into feeding guilds using stomach contents data and then
using morphological traits to predict feeding guild membership. This approach is wellestablished, having been used to study a number of fish communities in freshwater and
marine systems across temperate and tropical regions, particularly reefs (e.g. Ramírez et al.
2015; Winkler et al. 2017; Podder et al. 2021; Albouy et al. 2011). Within the Southern

- Ocean, however, only one study has combined morphological analyses with direct dietary
  observations in this manner, focussing on ten species of the family Artedidraconidae in the
  Weddell Sea and identifying links between their ecological niches and their sensory
- 2753 capability and mouth morphology (Lombarte et al. 2003).

2754 My investigation of the South Georgia demersal fish community encompassed three 2755 taxonomic families (Channichthyidae, Nototheniidae and Bathydraconidae) and a large 2756 sample size, thereby providing a substantial contribution to our understanding of the 2757 associations between functional traits and ecology in Southern Ocean communities. I showed 2758 that a small number of simple, easily measurable traits can successfully capture most of the 2759 broad dietary niches present across much of the community, providing insight into the drivers 2760 of trophic interactions. This is an approach that could be applied to other Southern Ocean 2761 communities to improve our understanding of the drivers of food web structure. The ecology 2762 of demersal fish has been comprehensively studied around many regions of the Southern 2763 Ocean encompassing a diversity of bioregions (e.g. Wang et al. 2024; Cousins and Priede 2764 2012; Baena et al. 2023) but the influence of functional traits has not yet been explored for 2765 these communities. It would be interesting to investigate how spatial contrasts in factors such 2766 as prey species assemblages, benthopelagic coupling or abiotic forcing (such as between low 2767 and high latitudes or east and west-Antarctic regions) relate to differences in the association 2768 between morphological traits and trophic niches. This could be facilitated by the routine 2769 sampling of diets and associated standard morphological traits such as mouth size and shape, 2770 fin morphology, body shape and gill structure across ecological communities during scientific 2771 and fishery expeditions.

2772 Chapter 2 also highlights the possibility to identify broad dietary niches based on 2773 morphology, which could be useful for combining species into functional groups to develop 2774 better-resolved ecosystem models in data-poor environments (Ladds et al. 2018; Albouy et al. 2775 2011). A focus on traits rather than taxonomy also provides a framework for determining how 2776 environmental and biotic filtering drive functional diversity (Green et al. 2022). Within 2777 marine taxa, body size is the most commonly measured trait, likely due to its major role in 2778 structuring marine food webs and the relative ease with which it can be measured (Green et 2779 al. 2022; Potapov et al. 2019). However, as shown in this chapter, other morphological traits 2780 reflecting feeding mode and mobility can also provide insight into the drivers of niche 2781 partitioning and should therefore be included in trait-based studies. It must also be recognized 2782 that, while morphological traits provide an extremely useful basis for investigating

2783 community ecology, their use alone may not be sufficient to fully explain trophic structure. 2784 For example, the widespread distribution of krill feeders across morphological niche space 2785 could reflect a degree of trophic plasticity which is not linked obviously to morphology, and 2786 it will be important to consider further context such as general prey availability and abiotic 2787 factors. The wide range of morphologies exhibited by krill feeders could also suggest that 2788 krill possess certain traits which allow fish to feed outside their evolutionary morphological 2789 niches. These traits might include their high energy content and often widespread distribution 2790 across pelagic and benthic habitats, which make them an accessible and energetically 2791 efficient prey item for predators displaying a range of feeding modes, habitats and mobilities. 2792 This emphasizes the importance of considering both predator and prey characteristics when 2793 exploring the functional trait basis of trophic interactions (Wootton et al. 2023; Laigle et al. 2794 2017). Within the Southern Ocean, trait-based approaches are still rare (McCormack et al. 2795 2021a) and it will be important to determine the distribution of functional traits across more 2796 components of the ecosystem, as this would allow us to compare the positions of different 2797 species in multi-dimensional trait space and could help elucidate key differences which map 2798 onto their roles within food webs.

2799 It is possible that ongoing environmental change around South Georgia will drive shifts in the 2800 distribution of species and broader community composition, particularly under the loss of key 2801 groups such as Antarctic krill (Kawaguchi et al. 2024; Whitehouse et al. 2008). There are still 2802 several unanswered questions regarding the structure of the South Georgia food web, most 2803 crucial being the distribution of feeding interactions in response to interannual variation in 2804 the local abundance of krill, as this could provide insight into possible future ecosystem 2805 states. Chapter 2 therefore represents a baseline study of the position of the demersal fish 2806 community among ecological niche space which further work can build upon to describe how 2807 interannual changes in prey availability influence ecomorphological niche partitioning. In 2808 particular, it would be interesting to investigate how changes in the relative abundance of 2809 different prey and demersal fish species affect the abundance-weighted diversity of traits 2810 within multivariate space (e.g. Liu et al. 2019), as this would provide insight into the 2811 potential impacts of future ecological change for functional diversity.

2812 6.1.2 Chapter 3: Trophic structuring of modularity alters energy flow through marine food
2813 webs

The presence of modules within ecological networks is well established but investigations of this structure and links to functional traits remain quite limited, with previous studies

2816 generally using a small set of traits (e.g. body size, foraging habitat and interaction type) to 2817 explain module membership (Kortsch et al. 2015; Rezende et al. 2009; Montoya et al. 2015). 2818 Within the Southern Ocean, there has been comparatively little effort to explore the basic 2819 food web theory which has dominated research in other regions, including the presence of 2820 modules and relevance of species traits (McCormack et al. 2021a). My work in chapter 3 2821 therefore provides much needed insight into how functional traits underly the structure of 2822 Southern Ocean food webs. I conducted an extensive review of a broad range of both 2823 predator and prey characteristics for each species within multiple food webs, encompassing 2824 size, foraging behaviour, motility and defensive traits, amongst others. This dataset represents 2825 a valuable resource for further studies wishing to use these food webs to investigate how 2826 functional traits relate to network structure. Many of the traits could also be transferred to 2827 similar species in other regions, making this dataset useful beyond the focal models for 2828 investigating topics such as environmental filtering and the drivers of niche partitioning.

2829 Modules can be structured by trophic level, whereby modules encompass relatively distinct 2830 groupings of trophic levels and form a hierarchy from the base of the food web to higher 2831 predators (Kortsch et al. 2019; Rezende et al. 2009; Guimera et al. 2010), or by energy 2832 channel, whereby modules partition food webs into trophic chains running from low to high 2833 trophic levels, often encompassing discrete basal resources (Gauzens et al. 2015; Zhao et al. 2834 2017; Rodriguez et al. 2022). Previous studies have analysed individual networks exhibiting 2835 varying levels of taxonomic aggregation and have used a variety of methods to determine 2836 modularity, making it difficult to determine whether differences in module structure between 2837 studies are due to genuine ecological contrasts or methodological factors (Gauzens et al. 2838 2013). In contrast, I used four food webs selected specifically for their high taxonomic 2839 resolution and employed a consistent method for determining modularity, making direct 2840 comparisons possible. A key finding was that module structuring is not consistent, with the 2841 'trophic level' structure identified in two food webs while the remainder displayed the 2842 'energy channel' structure. The relative importance of functional traits for predicting module 2843 membership also differed, with body mass found to be key in the food webs with structuring 2844 by trophic level, and feeding strategy important for the energy channel structure, while 2845 mobility and habitat were important across all networks. I explained these differences in terms of the levels of environmental heterogeneity inherent in each of the modelled systems, 2846 2847 as this can strongly modify network structure including modularity (Kortsch et al. 2019). I 2848 proposed that networks in more homogenous ocean environments have reduced niche

2849 diversity at lower basal levels, being centered primarily on phytoplankton, and are therefore 2850 structured largely by body mass, which is thought to be a primary driver of marine food web 2851 structure in general (Petchey et al. 2008; Rall et al. 2012; Potapov et al. 2019). In systems 2852 subject to stronger environmental gradients and variability, the heterogeneity of available 2853 habitat and basal resources could drive a diversity of trophic niches resulting in more 2854 specialized modules encompassing energy channels. These results are analogous to the 2855 habitat heterogeneity hypothesis (Thompson and Townsend 2005), whereby more varied 2856 habitats provide a greater diversity of niches and resources, supporting a greater diversity of 2857 species (or in this case, energy channels).

2858 This research highlights key aspects regarding the use of functional traits and investigations 2859 of modularity that require further consideration. Firstly, underlying abiotic factors may 2860 modify the relative importance of different functional traits for determining network 2861 organisation. This means that a set of traits that provides good predictions of network structure in one region may not be so useful in another, and the environmental gradients 2862 2863 influencing trait distributions will need to be considered before one can use traits to make 2864 generalised predictions of ecosystem structure. This calls for more investigation of the links 2865 between functional trait diversity (and identity) and environmental factors. If generalisable 2866 rules can be established (e.g. greater basal resource heterogeneity results in greater 2867 differentiation in feeding behaviours and therefore a stronger effect of feeding mode on 2868 network structure), then it will be possible to tailor trait-based approaches to specific 2869 ecosystems, improving predictive accuracy.

2870 Secondly, rather than focusing solely on the absolute value of modularity, researchers should 2871 also investigate the structuring of modules in relation to trophic levels, as this could be particularly important for stability. Modularity reduces the overall connectivity within the 2872 2873 network, thereby limiting the propagation of extinctions (Stouffer and Bascompte 2011), but 2874 in cases where modules are arranged largely by trophic level, perturbations within lower 2875 modules might still cascade up to higher modules. Two networks with similar values of 2876 modularity but different modular arrangement may therefore have quite different capacity to 2877 cope with species loss. This could be evaluated with formal stability analyses using 2878 theoretical food webs with standardized characteristics such as network size and complexity. 2879 Expanding these analyses to more real food webs will require the development of a greater 2880 number of highly resolved networks across a suite of environmental gradients. A systematic 2881 effort to sample and describe the structure of food webs to the highest taxonomic resolution

2882 possible across a variety of habitats and environmental conditions would therefore be 2883 extremely valuable. This would aid in the search for generalisable network structures and 2884 could highlight regions of the Southern Ocean which may be more susceptible to 2885 environmental and ecological perturbations. It would also be useful to extend such food web 2886 analyses to quantitative networks, as the inclusion of interaction weights can greatly alter 2887 structural inferences (Banašek-Richter et al. 2009). The construction of quantitative networks 2888 is data-intensive, but recent developments make it possible to determine interaction strengths 2889 and energy fluxes based on relatively straightforward information such as body size, foraging 2890 behaviour and metabolic type (Marina et al. 2024; Gauzens et al. 2019), which should 2891 facilitate their adoption more widely within the Southern Ocean modelling community. It will 2892 still however be important to ground truth these estimates of energy flow with direct 2893 measurements of parameters such as consumer dietary preferences, metabolic rates and 2894 assimilation efficiencies (Jochum et al. 2021), as factors such as the distribution of 2895 interactions and the plasticity of metabolic rates can greatly alter the efficiency of energy 2896 flow and the magnitude of predicted energy fluxes (Kordas et al. 2022; Jochum and 2897 Eisenhauer 2022).

2898 6.1.3 Chapter 4: Temperature alters the predator-prey size relationships and size-selectivity
2899 of Southern Ocean fish

2900 Chapter 4 targeted our lack of knowledge regarding how predator-prey mass ratios (PPMR) 2901 change with temperature, by investigating how the relative sizes of mesopelagic myctophid 2902 fish and their zooplankton prey vary across a large latitudinal temperature gradient in the 2903 Southern Ocean. This is of particular interest as marine ecosystems are strongly size-2904 structured and the relative size of predators to their prey has been used to successfully predict 2905 the distribution and even the strength of trophic interactions (Petchey et al. 2008; Bideault et 2906 al. 2019; Emmerson and Raffaelli 2004). Changes in the distribution of body sizes across 2907 trophic levels could therefore alter the flows of energy within food webs, with implications 2908 for ecosystem functioning.

- 2909 By combining dietary and environmental prey size distributions it was possible to estimate
- 2910 the 'preferred PPMR', which distinguishes density-dependent and active prey selection by the
- fish (Tsai et al. 2016). This revealed some of the mechanisms underlying foraging by these
- 2912 fish under different temperatures. In particular, decreases in the average body size of fish (due
- 2913 to compositional changes and intra-specific declines in size) and shifts in the size distribution
- 2914 of zooplankton towards intermediate individuals acted together to reduce community-level

2915 PPMR. These results could be used to better inform the parameterization of predictive size2916 based models of food web dynamics, which often assume a fixed PPMR when assigning
2917 trophic interactions and may therefore fail to account for environmentally-driven shifts in
2918 prey selection (Andersen et al. 2016; Tsai et al. 2016).

2919 As illustrated by Gauzens et al. (2024), shifts in foraging behaviour and size-selectivity can 2920 be maladaptive, with important consequences for the persistence of communities under 2921 perturbations. It is therefore vital that we continue to investigate how environmental and 2922 ecological changes alter feeding preferences across taxa. It would also be interesting to 2923 further investigate the mechanisms that could be underlying the shifts in size selection by 2924 these myctophids from an energetic perspective. Previous research has shown that, in 2925 sardines, energy expenditure is influenced not only by warming but also by the size of prey 2926 available; if prey are small, sardines feed by continuous filtration which is more energy 2927 intensive than the particle feeding method employed when prey are larger, resulting in much 2928 higher energy expenditure (Queiros et al. 2024). Feeding trials of myctophids under different 2929 prey size treatments might reveal similar changes in foraging behaviour, providing further 2930 insight into the factors driving prey selection and energetics within this community.

2931 As discussed in chapter 1, the impacts of climate change on populations may be driven as 2932 much by changes to species interactions as by direct environmental effects on organisms 2933 themselves (Ockendon et al. 2014). Chapter 4 only focussed on one (albeit important) 2934 component of Southern Ocean ecosystems, and the investigation of the impact of temperature 2935 on trophic interactions should be extended to other groups including zooplankton. 2936 Experiments have been conducted on groups such as Antarctic krill and amphipods to 2937 identify the effects of temperature on metabolic rates, growth, feeding rates and mortality 2938 (Michael et al. 2021; Saba et al. 2021; Schram et al. 2016), and it would be worthwhile to 2939 extend such experiments to include investigations of how temperature influences their prev 2940 selection (both in terms of species identity and size). Gaining this understanding for a wide 2941 range of functional groups within the Southern Ocean will greatly improve our capacity to 2942 predict how climate change may reorganize ecosystems.

Given the link between PPMR and interaction strengths (Bideault et al. 2019; Emmerson and Raffaelli 2004), the community-level decline in PPMR identified in chapter 4 might represent a change in community stability. The relationship between PPMR and interaction strength in this community could be tested by measuring the densities of prey under different predator

- 2947 conditions in mesocosms (as in O'Gorman et al. 2010 and Emmerson and Raffaelli 2004),
- 2948 with different combinations of predator and prey size across multiple components of the food
- 2949 web. This information could then be used to parameterise population dynamics models (e.g.
- 2950 Gauzens et al. 2024; Bideault et al. 2019) to formally test the implications of changes in size
- structure (i.e. interaction strengths) for different facets of stability such as population
- 2952 variability and robustness.

## 2953 6.1.4 Chapter 5: Trade-offs between the recovery of Southern Ocean baleen whales and 2954 conservation of their competitors

2955 Chapter 5 provides important insights into how whale population recovery may impact 2956 Southern Ocean ecosystems, which will be a key issue for policymakers wanting to 2957 implement appropriate conservation measures under climate change. Various studies have 2958 concluded that whaling had major effects on prey and competitor abundances and overall 2959 food web dynamics (Laws 1977; Surma et al. 2014), but less attention has been drawn to the 2960 possible ecological consequences of whale population recoveries. By generating a broad suite 2961 of regional model structures, it was possible to identify some structural metrics which might 2962 be important indicators of the capacity for ecosystems to support increased whale biomass. 2963 Such indicator metrics are a key management tool for monitoring ecosystem health and 2964 resilience (Keramidas et al. 2023; Flensborg et al. 2023), and will aid the management and 2965 conservation of Southern Ocean ecosystems (Ruckelshaus et al. 2008). The analyses in this 2966 chapter focused primarily on broad responses averaged across competitor groups and model 2967 runs. There is therefore scope to further explore how the relationship between the focal 2968 metrics and model capacity varies at the individual model and functional group level and in 2969 relation to other factors (e.g. the distribution of biomass and *Q/B* values across competitor 2970 groups, or the *EE* distributions across whale prey groups). This would improve our 2971 understanding of the drivers of regional differences in ecosystem responses to whale 2972 population increases and provide greater insight into the reliability of different indicators of 2973 ecosystem resilience.

There is also scope to further investigate the factors determining regional ecosystem responses to whale recovery. Regular monitoring of the composition and diets of a range of functional groups across different regions and environmental conditions would increase our understanding of the spatial and temporal variability of trophic interactions. This would improve our ability to accurately model regional contrasts in ecosystem structure and incorporate the influence of dietary flexibility into predictions. Techniques such as DNA

2980 metabarcoding of environmental samples and stomach contents or scats could prove 2981 extremely useful for monitoring food web structure and exploring spatial differences, as they 2982 provide a relatively cost-effective way of identifying marine community composition and 2983 monitoring trophic interactions across time and space (Canals et al. 2024). This could be 2984 facilitated through the routine collection of samples from fishing vessels, which has been 2985 found to be a very effective way of reconstructing food webs in exploited areas (Cicala et al. 2986 2024). Tourist cruise vessels, which make extensive journeys across much of the Southern 2987 Ocean every year (McCarthy et al. 2022), are also a useful platform for conducting routine 2988 sampling of communities. Even the simple collection of surface water samples, such as 2989 through continuous plankton recorders (CPRs), would improve the monitoring of 2990 zooplankton and phytoplankton community composition across environmental gradients. The 2991 widespread adoption of CPR devices on tourist cruise vessels would greatly increase the 2992 coverage of existing datasets such as the SCAR Southern Ocean Continuous Plankton 2993 Recorder (SO-CPR) Survey, which is currently based primarily on samples from research and 2994 fishing vessels (Hosie et al. 2003). This would provide insight into questions such as the 2995 likelihood of regional changes in phytoplankton composition and production which mitigate 2996 baleen whale population increases in different regions.

2997 The balancing algorithm developed for this chapter helps address a primary limitation of the 2998 Ecopath with Ecosim (EwE) framework, which has been the lack of capabilities to 2999 incorporate parameter uncertainty during the modelling process (Steenbeek et al. 2018). The 3000 ability to generate plausible balanced parameter sets from an initial highly unbalanced model 3001 is valuable, as current existing EwE uncertainty plugins (e.g. 'Ecosampler') work with input 3002 models that are at, or close to, balance (Steenbeek et al. 2018), while other methods generate 3003 unbalanced models which then need testing to ensure they are thermodynamically viable 3004 (Whitehouse and Aydin 2020). The issue of model standardization, which was addressed in 3005 this chapter, applies more broadly to all comparative modeling studies across the Southern 3006 Ocean and beyond, as many network metrics are sensitive to model structure (Heymans et al. 3007 2016). Efforts to identify further indicators of ecosystem resilience must therefore ensure that 3008 models are directly comparable. The approach to model standardization and balancing taken 3009 in chapter 5 could be applied to many other comparative questions within the Southern Ocean 3010 including the effects of species loss or environmental regime shifts, aiding our ability to 3011 identify regional contrasts in food web structure and responses to change.

3012 This chapter made heavy use of *Rpath*, the R implementation of EwE (Lucey et al. 2020). 3013 The development of this package is exciting as it introduces a whole range of flexibility for 3014 modellers to ask questions that perhaps are not fully suited to the original EwE framework. 3015 For example, *Rpath* has been used to implement feedback mechanisms between an operating 3016 model and external assessment model to evaluate fishery management strategies (Lucey et al. 3017 2021). The package has also facilitated the incorporation of temperature-dependent energetic 3018 demands and metabolic costs into mass-balance models, which is a big step towards the 3019 development of robust predictions of the impacts of warming on species and ecosystem 3020 processes (Heinichen et al. 2022). These applications of *Rpath* will be extremely valuable for 3021 exploring management and conservation strategies in the Southern Ocean, where the EwE 3022 framework is relied on heavily (McCormack et al. 2021a).

#### 3023 6.2 Future Directions

This thesis touched upon a broad range of themes relating to different aspects of the organization and dynamics of Southern Ocean food webs, at a variety of spatial and ecological scales. However, there are some common threads regarding the future directions in which I see the research field heading, both within the Southern Ocean and more generally.

#### 3028 6.2.1 Using functional traits to explain and predict food web structure

3029 Functional traits clearly represent a valuable framework for explaining and predicting the 3030 current structure of food webs and will likely become increasingly popular for predicting the 3031 future effects of environmental change and shifting community compositions. Recent 3032 developments will aid the application of the trait-based approach; in particular, machine 3033 learning tools can be a powerful method for reconstructing changes in ecosystem properties 3034 from past records and predicting the structure of past and future networks from functional 3035 traits (Brown et al. 2023; Pichler et al. 2020; Fricke et al. 2022). Additionally, the 3036 development of AI-driven algorithms to extract ecological information from literature (e.g. 3037 Gougherty and Clipp 2024) will greatly aid trait-based food web research. I imagine that 3038 similar tools could also be developed to predict the occurrence of certain traits based on 3039 taxonomy by disentangling the relative effects of environmental variables and phylogeny on 3040 trait expression (Sanchez-Martinez et al. 2024), which would make it easier to compile trait 3041 information for poorly studied species and regions. It is also important to recognise that 3042 trophic interactions are highly multidimensional, made up of multiple component steps (e.g. 3043 prey identification, capture, consumption) during which the probability of success is

determined by a variety of different traits and abiotic factors (Wootton et al. 2023).

- 3045 Considering the matching between interactions and functional traits in a more fine-scale,
- 3046 modular manner will facilitate the quantification and comparison of the relative influence of
- 3047 different traits, component steps and abiotic factors, and improve the prediction of network
- 3048 structure and dynamics (Wootton et al. 2023).

3049 Overall, only a minority of studies have used functional traits to make predictions of the 3050 impacts of global change on ecological communities (Green et al. 2022). As this becomes 3051 more of a research focus, it is likely that the demand for extensive and well-resolved trait data 3052 will increase. As previously discussed, body size is a key trait which is easily measurable and 3053 has been used to successfully predict trophic interactions (Petchey et al. 2008). However, as 3054 shown in this thesis and in previous studies (e.g. Brose et al. 2019; Morales-Castilla et al. 3055 2015; Laigle et al. 2017; Rezende et al. 2009; Jacob et al. 2011), a variety of other traits 3056 including habitat association, mobility, feeding mode, and other behavioural and 3057 physiological characteristics can also play major roles in determining whether organisms 3058 interact. A key recommendation is therefore that we further develop datasets of functional 3059 traits, expanding them to encompass more species and aspects of organismal ecology, and to explicitly consider the various component steps involved in trophic interactions. Trait 3060 3061 databases already exist for many regions and taxonomic groups (for example this thesis made heavy use of resources such as Brun et al. 2017, Degen and Faulwetter 2019 and MarLIN 3062 3063 2006) but many data gaps remain, particularly for Southern Ocean taxa (Degen et al. 2018). I 3064 would therefore encourage researchers to consider what traits they can easily measure for 3065 their study organisms to further add value and support trait-based research. Doing so will 3066 allow us to build upon existing tools such as the Allometric Diet Breadth Model (Petchey et 3067 al. 2008), improving our ability to predict food web structure. A danger here is that, without 3068 some degree of standardization and coordination between researchers, the resulting suite of 3069 trait databases may not be fully comparable or compatible between studies, limiting their 3070 utility. This subject is neatly summarized in Keller et al. (2023), along with relevant 3071 guidelines to avoid these issues. I would add that it is also worth considering which traits 3072 should be prioritized (e.g. those which provide the most explanatory power for predicting 3073 trophic interactions or those which are most strongly tied to important ecosystem functions, 3074 rather than simply being the easiest to measure) as this will avoid wasting research effort and 3075 resources on identifying traits which have little practical use. Of course, the identification of 3076 priority traits will require significant effort itself, but could be initially achieved by

theoretical modelling and small-scale experimental or mesocosm studies before being appliedmore widely.

#### 3079 6.2.2 Understanding temporal and spatial variability in food web structure

3080 A common theme amongst my chapters, and indeed from much of my wider reading, has 3081 been the need to identify how trophic interactions and network structure differ over space and 3082 time. This will provide insight into the drivers of variation in community assembly, including 3083 environmental filtering and coexistence mechanisms (Pellissier et al. 2018), and will help 3084 predict the consequences of environmental and ecological change. An increasing number of 3085 studies have focused on how food web structure differs geographically (e.g. Pellissier et al. 3086 2018; Frelat et al. 2022; Kortsch et al. 2019; Gauzens et al. 2020), and temporally (e.g. 3087 Griffith et al. 2019; Olivier et al. 2019; Kortsch et al. 2021; Frelat et al. 2022), but this aspect 3088 of food web research remains understudied, particularly in the Southern Ocean where 3089 sampling is logistically limited by its overall remoteness and the inhospitableness of winter 3090 months (Van De Putte et al. 2021). The Southern Ocean has a variety of strong environmental 3091 gradients, including sea temperature, sea ice concentration and productivity (Deppeler and 3092 Davidson 2017; Morley et al. 2010; Pinkerton et al. 2021), and the impact of these factors on 3093 community structure and dynamics should be explored. Additionally, in highly seasonal 3094 environments such as polar regions, winter processes can have a strong influence on summer 3095 ecosystem dynamics, and indirect interactions between seasonally migrant and resident 3096 species can be important (Hutchison et al. 2020). Within Antarctic benthic communities in 3097 particular, seasonal sea ice break-up has been found to strongly alter food web structure, 3098 resulting in simpler, more vulnerable networks (Rossi et al. 2019; Caputi et al. 2020). This 3099 suggests that temporal changes may be a particularly fruitful topic of food web research 3100 within the Southern Ocean.

3101 I would recommend that efforts are made to construct a greater number of highly resolved 3102 and comparable food web models in a systematic manner across different global regions, and 3103 to put in place appropriate monitoring plans to facilitate the investigation of temporal changes 3104 in structure. Given the logistical constraints that apply in the Southern Ocean, it may be 3105 necessary to select a small number of sampling sites encompassing different bioregions 3106 which can be sampled regularly enough to develop time series of community composition 3107 and associated network structure and dynamics. These could target some key CCAMLR Marine Protected Area (MPA) planning domains (Teschke et al. 2021), thereby providing 3108 3109 important baselines for ongoing monitoring of the effectiveness of current and future MPAs.

3110 As shown by (Frelat et al. 2022), spatio-temporal monitoring of food web structure can be 3111 facilitated through the generation of a 'metaweb' of potential trophic interactions, which can 3112 then be resampled based on species abundances to generate spatial and temporal snapshots of 3113 network structure in a relatively straightforward manner. I would also argue that, in addition 3114 to simply describing how the distribution of trophic interactions differs across environmental 3115 gradients and over time, it is important that we move past mere correlative studies and begin 3116 to model the mechanisms linking changing network structure to its drivers. This is discussed 3117 further below.

#### 3118 6.2.3 Using bioenergetics to gain mechanistic understanding of food web dynamics

3119 To accurately forecast the impacts of environmental change for ecological communities and 3120 ecosystem functioning, we will need to understand the mechanisms linking biotic and abiotic 3121 factors to population dynamics, communities and overall ecosystem processes. This could be 3122 addressed through the bioenergetics approach, which explicitly considers the physiological 3123 and behavioural responses of organisms and accounts for the variety of indirect effects and 3124 non-linear responses which will result from changing environmental and ecological 3125 conditions (Rose et al. 2024). Energy is a relevant currency across all scales of spatial, 3126 temporal and ecological organisation, from individual cells to entire ecosystems, therefore 3127 focussing on energy flow provides a tractable basis for investigating links between different 3128 levels of biological organisation (Carlisle 2000). In particular, methods such as Dynamic 3129 Energy Budget (DEB) modelling can be used to predict both inter- and intra-specific 3130 variation in energy and mass fluxes in response to changing environments (Rose et al. 2024; Nisbet et al. 2012). DEB modelling is a highly generalisable approach which can be applied 3131 3132 to any animal to predict its intake and utilisation of energy and relate metabolic processes to 3133 physiological performance and thus wider population dynamics and ecosystem processes 3134 (Nisbet et al. 2012). This allows researchers to investigate how abiotic conditions and food 3135 availability affect organismal growth, feeding and reproduction (Pouvreau et al. 2006; Agüera 3136 et al. 2017; van der Meer et al. 2020; Teixeira et al. 2014). This approach can be scaled up to entire ecosystems (van der Meer et al. 2022), and integrating it into broader food web 3137 3138 modelling will allow us to mechanistically understand and predict how communities respond 3139 to change.

3140 The field of food web ecology is increasingly recognising the value of bioenergetic

approaches for tracking energy flux and studying the dynamics of multi-species assemblages.

3142 A number of tools based upon bioenergetics and metabolic theory have been developed for

3143 different platforms (e.g. Gauzens et al. 2019; Gauzens et al. 2023; Delmas et al. 2017), and 3144 used to ask questions such as how environmental change impacts ecosystem function and 3145 stability (Polazzo et al. 2023), how physiological plasticity influences ecosystem impacts of 3146 warming (Kordas et al. 2022), and what the interactive effects of multiple stressors are on 3147 patterns of energy flux (Wang et al. 2023). As previously discussed, the inclusion of temperature-dependent bioenergetics within the EwE framework has provided further insight 3148 3149 into the potential effects of warming on biomass production within marine ecosystems 3150 (Heinichen et al. 2022). The flux-based approach has also revealed how trophic redundancy 3151 can mitigate the impacts of warming on total energy flow within food webs (Nelson et al. 3152 2020). As these bioenergetic approaches are largely based on theory and generalisable 3153 relationships, they could be well suited to locations such as the Southern Ocean where the 3154 observational and experimental data required to parameterise more complex dynamic models 3155 are scarce (McCormack et al. 2021a; Murphy et al. 2012). It will, however, be important to 3156 consider whether some of the characteristics of many Southern Ocean taxa, e.g. stenothermy, 3157 mean they do not adhere to theoretical relationships. This emphasises the need to further 3158 investigate the fundamental rates of different Southern Ocean taxa and how they respond to 3159 environmental changes such as warming. Given that the rate of physiological process can 3160 vary between individuals and there may be intra-specific variability in physiological and 3161 behavioural responses to different stressors (Gårdmark and Huss 2020), bioenergetic 3162 approaches may be best suited to individual-based food webs which explicitly consider 3163 populations and size-classes rather than simply aggregating at the species or functional group 3164 level (Woodward et al. 2010; Gårdmark and Huss 2020).

#### 3165 *6.2.4 Summary*

3166 The results of this thesis contribute to our growing understanding of the drivers of food web 3167 structure and the impacts of environmental change. While the focus of my chapters was 3168 primarily on the Southern Ocean, many of my conclusions are also relevant to the food web 3169 modelling field more broadly. In particular, I have provided insight into some of the core 3170 aspects of food web theory, namely the relationship between functional traits and the distribution of trophic interactions, the organisation of stabilising substructures, and the 3171 3172 influence of the environment on size-based interactions. I see various avenues of research 3173 through which the food web field will advance in future. On the one hand, 'more of the same' 3174 (continued characterisation of diets and traits, and construction of networks across temporal 3175 and spatial scales and environmental gradients) will allow us to more robustly test the effects

- 3176 of abiotic and biotic factors on food web structure and dynamics. On the other hand, I foresee
- 3177 that a shift towards flux-based approaches, and a focus on individual-based food webs and
- 3178 explicit consideration of dynamic consumers with flexible diets, offers the chance to gain
- 3179 more mechanistic understanding of the processes underlying the formation and maintenance
- 3180 of natural communities. I'm sure that cutting-edge developments such as super-computing,
- 3181 machine-learning and AI will open up further possibilities for modelling ecosystems what a
- 3182 time to be a food web researcher!

### 3184 **References**

3185 Abrams, P. A. 2010. Implications of Flexible Foraging for Interspecific Interactions: 3186 Lessons from Simple Models. Functional Ecology, 24, 7-17. 3187 Agiadi, K., Quillévéré, F., Nawrot, R., Sommeville, T., Coll, M., Koskeridou, E., Fietzke, 3188 J. & Zuschin, M. 2022. Palaeontological Evidence for Community-Level Decrease in Mesopelagic Fish Size During Pleistocene Climate Warming in the Eastern 3189 3190 Mediterranean. bioRxiv, 2022.10.04.510798. 3191 Agüera, A., Ahn, I.-Y., Guillaumot, C. & Danis, B. 2017. A Dynamic Energy Budget 3192 (Deb) Model to Describe Laternula Elliptica (King, 1832) Seasonal Feeding and 3193 Metabolism. PLOS ONE, 12, e0183848. 3194 Åkesson, A., Curtsdotter, A., Eklöf, A., Ebenman, B., Norberg, J. & Barabás, G. 2021. 3195 The Importance of Species Interactions in Eco-Evolutionary Community Dynamics under 3196 Climate Change. Nature Communications, 12. Albouy, C., Guilhaumon, F., Villéger, S., Mouchet, M., Mercier, L., Culioli, J., Tomasini, 3197 3198 J., Le Loc'h, F. & Mouillot, D. 2011. Predicting Trophic Guild and Diet Overlap from 3199 Functional Traits: Statistics, Opportunities and Limitations for Marine Ecology. Marine 3200 Ecology Progress Series, 436, 17-28. 3201 Allouche, O., Tsoar, A. & Kadmon, R. 2006. Assessing the Accuracy of Species 3202 Distribution Models: Prevalence, Kappa and the True Skill Statistic (Tss). Journal of 3203 Applied Ecology, 43, 1223-1232. 3204 Andersen, K. H., Jacobsen, N. S. & Farnsworth, K. D. 2016. The Theoretical Foundations 3205 for Size Spectrum Models of Fish Communities. Canadian Journal of Fisheries and 3206 Aquatic Sciences, 73, 575-588. 3207 Arim, M., Bozinovic, F. & Marquet, P. A. 2007. On the Relationship between Trophic 3208 Position, Body Mass and Temperature: Reformulating the Energy Limitation Hypothesis. 3209 Oikos, 116, 1524-1530. 3210 Arnott, S. A., Neil, D. M. & Ansell, A. D. 1998. Tail-Flip Mechanism and Size-Dependent 3211 Kinematics of Escape Swimming in the Brown Shrimp Crangon Crangon. Journal of 3212 Experimental Biology, 201, 1771-1784. Atkinson, A., Hill, S. L., Pakhomov, E. A., Siegel, V., Reiss, C. S., Loeb, V. J., Steinberg, 3213 3214 D. K., Schmidt, K., Tarling, G. A., Gerrish, L. & Sailley, S. F. 2019. Krill (Euphausia Superba) Distribution Contracts Southward During Rapid Regional Warming. Nature 3215 Climate Change, 9, 142-147. 3216 3217 Atkinson, A., Hill, S. L., Reiss, C. S., Pakhomov, E. A., Beaugrand, G., Tarling, G. A., 3218 Yang, G., Steinberg, D. K., Schmidt, K., Edwards, M., Rombola, E. & Perry, F. A. 2022. 3219 Stepping Stones Towards Antarctica: Switch to Southern Spawning Grounds Explains an 3220 Abrupt Range Shift in Krill. Glob Chang Biol, 28, 1359-1375.

- Atkinson, A., Siegel, V., Pakhomov, E. A., Jessopp, M. J. & Loeb, V. 2009. A ReAppraisal of the Total Biomass and Annual Production of Antarctic Krill. *Deep Sea Research Part I: Oceanographic Research Papers*, 56, 727-740.
- Atkinson, A., Ward, P., Hunt, B. P. V., Pakhomov, E. A. & Hosie, G. W. 2012. An
  Overview of Southern Ocean Zooplankton Data: Abundance, Biomass, Feeding and
  Functional Relationships. *CCAMLR Science*, 19, 171 218.
- Auger, M., Morrow, R., Kestenare, E., Sallée, J.-B. & Cowley, R. 2021. Southern Ocean
   in-Situ Temperature Trends over 25 Years Emerge from Interannual Variability. *Nature Communications*, 12.
- Aydin, K. Y., Mcfarlane, G. A., King, J. R., Megrey, B. A. & Myers, K. W. 2005. Linking
  Oceanic Food Webs to Coastal Production and Growth Rates of Pacific Salmon
  (Oncorhynchus Spp.), Using Models on Three Scales. *Deep Sea Research Part II: Topical Studies in Oceanography*, 52, 757-780.
- Bachiller, E. & Irigoien, X. 2013. Allometric Relations and Consequences for Feeding in
  Small Pelagic Fish in the Bay of Biscay. *ICES Journal of Marine Science*, 70, 232-243.
- Baena, P., Santín, A., La Mesa, M., Riginella, E., Owsianowski, N., Gili, J.-M. &
  Ambroso, S. 2023. Are There Distribution Patterns and Population Structure Differences
  among Demersal Fish Species in Relation to Antarctic Benthic Communities? A Case
  Study in the Weddell Sea. *Polar Biology*, 46, 1069-1082.
- Baines, M., Jackson, J. A., Fielding, S., Warwick-Evans, V., Reichelt, M., Lacey, C.,
  Pinder, S. & Trathan, P. N. 2022. Ecological Interactions between Antarctic Krill
  (Euphausia Superba) and Baleen Whales in the South Sandwich Islands Region–
  Exploring Predator-Prey Biomass Ratios. *Deep Sea Research Part I: Oceanographic Research Papers*, 189, 103867.
- Ballerini, T., Hofmann, E. E., Ainley, D. G., Daly, K., Marrari, M., Ribic, C. A., Smith, W.
  O. & Steele, J. H. 2014. Productivity and Linkages of the Food Web of the Southern
  Region of the Western Antarctic Peninsula Continental Shelf. *Progress in Oceanography*,
  122, 10-29.
- Banašek-Richter, C., Bersier, L.-F., Cattin, M.-F., Baltensperger, R., Gabriel, J.-P., Merz,
  Y., Ulanowicz, R. E., Tavares, A. F., Williams, D. D., Ruiter, P. C., Winemiller, K. O. &
  Naisbit, R. E. 2009. Complexity in Quantitative Food Webs. *Ecology*, 90, 1470-1477.
- Bansode, M. A., Eastman, J. T. & Aronson, R. B. 2014. Feeding Biomechanics of Five
  Demersal Antarctic Fishes. *Polar Biology*, 37, 1835-1848.
- Barlow, K., Boyd, I., Croxall, J., Reid, K., Staniland, I. & Brierley, A. 2002. Are Penguins
  and Seals in Competition for Antarctic Krill at South Georgia? *Marine Biology*, 140, 205213.
- Barnes, C. L., Beaudreau, A. H. & Yamada, R. N. 2021. The Role of Size in Trophic
  Niche Separation between Two Groundfish Predators in Alaskan Waters. *Marine and Coastal Fisheries*, 13, 69-84.

3260 3261 3262	Barnes, D., Griffiths, H. & Kaiser, S. 2009. Geographic Range Shift Responses to Climate Change by Antarctic Benthos: Where We Should Look. <i>Marine Ecology Progress Series</i> , 393, 13-26.
3263	Barr, A. W. 2018. Ecomorphology. Methods in paleoecology 339-349.
3264 3265 3266	Bartomeus, I., Gravel, D., Tylianakis, J. M., Aizen, M. A., Dickie, I. A. & Bernard- Verdier, M. 2016. A Common Framework for Identifying Linkage Rules across Different Types of Interactions. <i>Functional Ecology</i> , 30, 1894-1903.
3267 3268 3269 3270	Belcher, A., Saunders, R. & Tarling, G. 2019. Length, Weight and Abundance Data of Fish Species Captured in Rmt-25 Net Surveys in the Scotia Sea, Southern Ocean in 2006, 2008, and 2009 (Version 1.0). UK Polar Data Centre, Natural Environment Research Council, UK Research & Innovation.
3271 3272	Bell, J. J. 2002. Morphological Responses of a Cup Coral to Environmental Gradients. <i>Sarsia</i> , 87, 319-330.
3273 3274	Berlow, E. L., Navarrete, S. A., Briggs, C. J., Power, M. E. & Menge, B. A. 1999. Quantifying Variation in the Strengths of Species Interactions. <i>Ecology</i> , 80, 2206-2224.
3275 3276 3277	Bideault, A., Loreau, M. & Gravel, D. 2019. Temperature Modifies Consumer-Resource Interaction Strength through Its Effects on Biological Rates and Body Mass. <i>Frontiers in</i> <i>Ecology and Evolution</i> , 7.
3278 3279 3280 3281	Biggs, C. R., Yeager, L. A., Bolser, D. G., Bonsell, C., Dichiera, A. M., Hou, Z., Keyser, S. R., Khursigara, A. J., Lu, K., Muth, A. F., Negrete, B. & Erisman, B. E. 2020. Does Functional Redundancy Affect Ecological Stability and Resilience? A Review and Meta-Analysis. <i>Ecosphere</i> , 11.
3282 3283 3284 3285	Biuw, M., Lindstrøm, U., Jackson, J. A., Baines, M., Kelly, N., Mccallum, G., Skaret, G. & Krafft, B. A. 2024. Estimated Summer Abundance and Krill Consumption of Fin Whales Throughout the Scotia Sea During the 2018/2019 Summer Season. <i>Scientific Reports</i> , 14.
3286 3287	Blanchard, J. L., Law, R., Castle, M. D. & Jennings, S. 2011. Coupled Energy Pathways and the Resilience of Size-Structured Food Webs. <i>Theoretical Ecology</i> , 4, 289-300.
3288 3289 3290 3291 3292	Blowes, S. A., Supp, S. R., Antão, L. H., Bates, A., Bruelheide, H., Chase, J. M., Moyes, F., Magurran, A., Mcgill, B., Myers-Smith, I. H., Winter, M., Bjorkman, A. D., Bowler, D. E., Byrnes, J. E. K., Gonzalez, A., Hines, J., Isbell, F., Jones, H. P., Navarro, L. M., Thompson, P. L., Vellend, M., Waldock, C. & Dornelas, M. 2019. The Geography of Biodiversity Change in Marine and Terrestrial Assemblages. <i>Science</i> , 366, 339-345.
3293 3294 3295	Boit, A., Martinez, N. D., Williams, R. J. & Gaedke, U. 2012. Mechanistic Theory and Modelling of Complex Food-Web Dynamics in Lake Constance. <i>Ecology Letters</i> , 15, 594-602.
3296 3297 3298	Bombosch, A., Zitterbart, D. P., Van Opzeeland, I., Frickenhaus, S., Burkhardt, E., Wisz, M. S. & Boebel, O. 2014. Predictive Habitat Modelling of Humpback (Megaptera Novaeangliae) and Antarctic Minke (Balaenoptera Bonaerensis) Whales in the Southern

3299 Ocean as a Planning Tool for Seismic Surveys. Deep Sea Research Part I: Oceanographic Research Papers, 91, 101-114. 3300 3301 Bonin, M., Dussault, C., Taillon, J., Lecomte, N. & Côté, S. D. 2020. Combining Stable 3302 Isotopes, Morphological, and Molecular Analyses to Reconstruct the Diet of Free-3303 Ranging Consumers. Ecology and Evolution, 10, 6664-6676. 3304 Bratbak, G. & Dundas, I. 1984. Bacterial Dry Matter Content and Biomass Estimations. Applied and Environmental Microbiology, 48, 755-757. 3305 Bridge, T. C. L., Luiz, O. J., Coleman, R. R., Kane, C. N. & Kosaki, R. K. 2016. 3306 Ecological and Morphological Traits Predict Depth-Generalist Fishes on Coral Reefs. 3307 3308 Proceedings of the Royal Society B: Biological Sciences, 283, 20152332. Brierley, A. S. & Kingsford, M. J. 2009. Impacts of Climate Change on Marine 3309 3310 Organisms and Ecosystems. Current Biology, 19, R602-R614. 3311 Brose, U., Archambault, P., Barnes, A. D., Bersier, L.-F., Boy, T., Canning-Clode, J., Conti, E., Dias, M., Digel, C., Dissanayake, A., Flores, A. a. V., Fussmann, K., Gauzens, 3312 B., Gray, C., Häussler, J., Hirt, M. R., Jacob, U., Jochum, M., Kéfi, S., Mclaughlin, O., 3313 3314 Macpherson, M. M., Latz, E., Layer-Dobra, K., Legagneux, P., Li, Y., Madeira, C., Martinez, N. D., Mendonça, V., Mulder, C., Navarrete, S. A., O'gorman, E. J., Ott, D., 3315 Paula, J., Perkins, D., Piechnik, D., Pokrovsky, I., Raffaelli, D., Rall, B. C., Rosenbaum, 3316 B., Ryser, R., Silva, A., Sohlström, E. H., Sokolova, N., Thompson, M. S. A., Thompson, 3317 R. M., Vermandele, F., Vinagre, C., Wang, S., Wefer, J. M., Williams, R. J., Wieters, E., 3318 Woodward, G. & Iles, A. C. 2019. Predator Traits Determine Food-Web Architecture 3319 3320 across Ecosystems. Nature Ecology & Evolution, 3, 919-927. 3321 Brose, U., Blanchard, J. L., Eklöf, A., Galiana, N., Hartvig, M., R. Hirt, M., Kalinkat, G., Nordström, M. C., O'gorman, E. J., Rall, B. C., Schneider, F. D., Thébault, E. & Jacob, U. 3322 3323 2017. Predicting the Consequences of Species Loss Using Size-Structured Biodiversity Approaches. Biological Reviews, 92, 684-697. 3324 3325 Brose, U., Williams, R. J. & Martinez, N. D. 2006. Allometric Scaling Enhances Stability 3326 in Complex Food Webs. Ecology Letters, 9, 1228-1236. Brown, K. A., Bunting, M. J., Carvalho, F., De Bello, F., Mander, L., Marcisz, K., Mottl, 3327 3328 O., Reitalu, T. & Svenning, J. C. 2023. Trait-Based Approaches as Ecological Time 3329 Machines: Developing Tools for Reconstructing Long-Term Variation in Ecosystems. 3330 Functional Ecology, 37, 2552-2569. 3331 Bruder, A., Frainer, A., Rota, T. & Primicerio, R. 2019. The Importance of Ecological 3332 Networks in Multiple-Stressor Research and Management. Frontiers in Environmental 3333 Science. 7. 3334 Brun, P., Payne, M. R. & Kiørboe, T. 2017. A Trait Database for Marine Copepods. Earth Syst. Sci. Data, 9, 99-113. 3335 3336 Buhl-Mortensen, L., Buhl-Mortensen, P., Dolan, M. F. J., Dannheim, J., Bellec, V. & 3337 Holte, B. 2012. Habitat Complexity and Bottom Fauna Composition at Different Scales 3338 on the Continental Shelf and Slope of Northern Norway. Hydrobiologia, 685, 191-219.

- Bury, S. J., Peters, K. J., Sabadel, A. J. M., St John Glew, K., Trueman, C., Wunder, M.
- 3340 B., Cobain, M. R. D., Schmitt, N., Donnelly, D., Magozzi, S., Owen, K., Brown, J. C. S.,
- 3341 Escobar-Flores, P., Constantine, R., O'driscoll, R. L., Double, M., Gales, N.,
- Childerhouse, S. & Pinkerton, M. H. 2024. Southern Ocean Humpback Whale Trophic
  Ecology. I. Combining Multiple Stable Isotope Methods Elucidates Diet, Trophic Position
  and Foraging Areas Marine Ecology Progress Series 724, 122, 155
- and Foraging Areas. *Marine Ecology Progress Series*, 734, 123-155.
- Calderan, S. V., Black, A., Branch, T. A., Collins, M. A., Kelly, N., Leaper, R., Lurcock,
  S., Miller, B. S., Moore, M., Olson, P. A., Širović, A., Wood, A. G. & Jackson, J. A. 2020.
  South Georgia Blue Whales Five Decades after the End of Whaling. *Endangered Species Research*, 43, 359-373.
- Calleja, M. L., Kerhervé, P., Bourgeois, S., Kędra, M., Leynaert, A., Devred, E., Babin,
  M. & Morata, N. 2017. Effects of Increase Glacier Discharge on Phytoplankton Bloom
  Dynamics and Pelagic Geochemistry in a High Arctic Fjord. *Progress in Oceanography*,
  159, 195-210.
- Canals, O., Lanzén, A., Mendibil, I., Bachiller, E., Corrales, X., Andonegi, E., Cotano, U.
  & Rodríguez-Ezpeleta, N. 2024. Increasing Marine Trophic Web Knowledge through
  DNA Analyses of Fish Stomach Content: A Step Towards an Ecosystem-Based Approach
  to Fisheries Research. *Journal of Fish Biology*, n/a.
- Caputi, S. S., Careddu, G., Calizza, E., Fiorentino, F., Maccapan, D., Rossi, L. &
  Costantini, M. L. 2020. Seasonal Food Web Dynamics in the Antarctic Benthos of Tethys
  Bay (Ross Sea): Implications for Biodiversity Persistence under Different Seasonal SeaIce Coverage. *Frontiers in Marine Science*, 7.
- Carlig, E., Di Blasi, D., Ghigliotti, L., Pisano, E., Faimali, M., O'driscoll, R., Parker, S. &
  Vacchi, M. 2018. Diversification of Feeding Structures in Three Adult Antarctic
  Nototheniid Fish. *Polar Biology*, 41, 1707-1715.
- Carlig, E., Di Blasi, D., Pisano, E., Vacchi, M., Santovito, G. & Ghigliotti, L. 2022.
  Ecomorphological Differentiation of Feeding Structures within the Antarctic Fish Species
  Flock Trematominae (Notothenioidei) from Terra Nova Bay (Ross Sea). *Journal of Marine Science and Engineering*, 10, 1876.
- Carlisle, D. M. 2000. Bioenergetic Food Webs as a Means of Linking Toxicological
  Effects across Scales of Ecological Organization. *Journal of Aquatic Ecosystem Stress and Recovery*, 7, 155-165.
- Carter, L., Mccave, I. N. & Williams, M. J. M. 2008. Chapter 4 Circulation and Water
  Masses of the Southern Ocean: A Review. *In:* Florindo, F. & Siegert, M. (eds.) *Developments in Earth and Environmental Sciences*. Elsevier.
- Casaux, R. & Barrera-Oro, E. 2013. Dietary Overlap in Inshore Notothenioid Fish from
  the Danco Coast, Western Antarctic Peninsula. *Polar Research*, 32, 21319.
- Cavanagh, R. D., Melbourne-Thomas, J., Grant, S. M., Barnes, D. K. A., Hughes, K. A.,
  Halfter, S., Meredith, M. P., Murphy, E. J., Trebilco, R. & Hill, S. L. 2021. Future Risk
  for Southern Ocean Ecosystem Services under Climate Change. *Frontiers in Marine Science*, 7.

3380 Ceballos, G., Ehrlich, P. R., Barnosky, A. D., García, A., Pringle, R. M. & Palmer, T. M. 2015. Accelerated Modern Human-Induced Species Losses: Entering the Sixth Mass 3381 Extinction. Science Advances, 1, e1400253. 3382 3383 Chapman, E. J., Byron, C. J., Lasley-Rasher, R., Lipsky, C., Stevens, J. R. & Peters, R. 3384 2020. Effects of Climate Change on Coastal Ecosystem Food Webs: Implications for 3385 Aquaculture. Marine Environmental Research, 162, 105103. 3386 Chaudhary, C., Richardson, A. J., Schoeman, D. S. & Costello, M. J. 2021. Global 3387 Warming Is Causing a More Pronounced Dip in Marine Species Richness around the Equator. Proceedings of the National Academy of Sciences, 118, e2015094118. 3388 3389 Chen, M., Kuzmin, K. & Szymanski, B. K. 2014. Community Detection Via Maximization of Modularity and Its Variants. IEEE Transactions on Computational 3390 Social Systems, 1, 46-65. 3391 3392 Cherel, Y., Fontaine, C., Richard, P. & Labatc, J.-P. 2010. Isotopic Niches and Trophic 3393 Levels of Myctophid Fishes and Their Predators in the Southern Ocean. Limnology and Oceanography, 55, 324-332. 3394 3395 Cherkasheva, A., Bracher, A., Melsheimer, C., Köberle, C., Gerdes, R., Nöthig, E. M., 3396 Bauerfeind, E. & Boetius, A. 2014. Influence of the Physical Environment on Polar Phytoplankton Blooms: A Case Study in the Fram Strait. Journal of Marine Systems, 132, 3397 3398 196-207. 3399 Cheung, W. W. L., Lam, V. W. Y. & Pauly, D. 2008. Modelling Present and Climate-3400 Shifted Distribution of Marine Fishes and Invertebrates. 3401 Cheung, W. W. L., Sarmiento, J. L., Dunne, J., Frölicher, T. L., Lam, V. W. Y., Deng 3402 Palomares, M. L., Watson, R. & Pauly, D. 2013. Shrinking of Fishes Exacerbates Impacts 3403 of Global Ocean Changes on Marine Ecosystems. Nature Climate Change, 3, 254-258. 3404 Christensen, B. 1996. Predator Foraging Capabilities and Prey Antipredator Behaviours: 3405 Pre- Versus Postcapture Constraints on Size-Dependent Predator-Prey Interactions. Oikos, 3406 76, 368-380. 3407 Christensen, L. B. 2006. Marine Mammal Populations: Reconstructing Historical 3408 Abundances at the Global Scale. Fisheries Centre research reports, 14. 3409 Christensen, V. & Walters, C. J. 2004. Ecopath with Ecosim: Methods, Capabilities and 3410 Limitations. Ecological Modelling, 172, 109-139. 3411 Cicala, D., Maiello, G., Fiorentino, F., Garofalo, G., Massi, D., Sbrana, A., Mariani, S., D'alessandro, S., Stefani, M., Perrodin, L. & Russo, T. 2024. Spatial Analysis of 3412 Demersal Food Webs through Integration of Edna Metabarcoding with Fishing Activities. 3413 Frontiers in Marine Science, 10. 3414 3415 Cirtwill, A. R. & Eklöf, A. 2018. Feeding Environment and Other Traits Shape Species' 3416 Roles in Marine Food Webs. Ecology Letters, 21, 875-884.

- Clark, J. S., Scher, C. L. & Swift, M. 2020. The Emergent Interactions That Govern
  Biodiversity Change. *Proceedings of the National Academy of Sciences*, 117, 1707417083.
- Clarke, S., Reid, W. D. K., Collins, M. A. & Belchier, M. 2008. Biology and Distribution
  of South Georgia Icefish (Pseudochaenichthys Georgianus) around South Georgia and
  Shag Rocks. *Antarctic Science*, 20, 343-353.
- Coghlan, A. R., Blanchard, J. L., Wotherspoon, S., Stuart-Smith, R. D., Edgar, G. J.,
  Barrett, N. & Audzijonyte, A. 2024. Mean Reef Fish Body Size Decreases Towards
  Warmer Waters. *Ecology Letters*, 27.
- Cohen, J. E. 2010. Ecologists' Co-Operative Web Bank. Version 1.1. Machine-Readable
  Database of Food Webs. New York: The Rockefeller University.
- Cohen, J. E., Jonsson, T. & Carpenter, S. R. 2003. Ecological Community Description
  Using the Food Web, Species Abundance, and Body Size. *Proceedings of the National Academy of Sciences*, 100, 1781-1786.
- Coll, M., Palomera, I., Tudela, S. & Sardà, F. 2006. Trophic Flows, Ecosystem Structure
  and Fishing Impacts in the South Catalan Sea, Northwestern Mediterranean. *Journal of Marine Systems*, 59, 63-96.
- Collins, M., Shreeve, R., Stowasser, G., Foster, E. & Saunders, R. 2020. Conventional
  Stomachs Contents Data for Mesopelagic Fish Collected from the Scotia Sea between
  2004-2009 (Version 1.0). UK Polar Data Centre, Natural Environment Research Council,
  UK Research & Innovation.
- Collins, M. A., Stowasser, G., Fielding, S., Shreeve, R., Xavier, J. C., Venables, H. J.,
  Enderlein, P., Cherel, Y. & Van De Putte, A. 2012. Latitudinal and Bathymetric Patterns in
  the Distribution and Abundance of Mesopelagic Fish in the Scotia Sea. *Deep Sea Research Part II: Topical Studies in Oceanography*, 59-60, 189-198.
- Constable, A. J. 2011. Lessons from Ccamlr on the Implementation of the Ecosystem
  Approach to Managing Fisheries. *Fish and Fisheries*, 12, 138-151.
- Convey, P., Bindschadler, R., Di Prisco, G., Fahrbach, E., Gutt, J., Hodgson, D. A.,
  Mayewski, P. A., Summerhayes, C. P. & Turner, J. 2009. Antarctic Climate Change and
  the Environment. *Antarctic Science*, 21, 541-563.
- Cooley, S., Schoeman, D., Bopp, L., Boyd, P., Donner, S., Ghebrehiwet, D. Y., Ito, S.-I.,
  Kiessling, W., Martinetto, P., Ojea, E., Racault, M.-F., Rost, B. & M., S.-M. 2022. Oceans
  and Coastal Ecosystems and Their Services. *Climate Change 2022 Impacts, Adaptation and Vulnerability. Contribution of Working Group Ii to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*
- Costa, G. C. 2009. Predator Size, Prey Size, and Dietary Niche Breadth Relationships in
  Marine Predators. *Ecology*, 90, 2014 2019.
- Costello, C., Cao, L., Gelcich, S., Cisneros-Mata, M. Á., Free, C. M., Froehlich, H. E.,
  Golden, C. D., Ishimura, G., Maier, J., Macadam-Somer, I., Mangin, T., Melnychuk, M.
  C., Miyahara, M., De Moor, C. L., Naylor, R., Nøstbakken, L., Ojea, E., O'reilly, E.,

3457 3458	Parma, A. M., Plantinga, A. J., Thilsted, S. H. & Lubchenco, J. 2020. The Future of Food from the Sea. <i>Nature</i> , 588, 95-100.
3459 3460 3461	Cousins, N. J. & Priede, I. G. 2012. Abyssal Demersal Fish Fauna Composition in Two Contrasting Productivity Regions of the Crozet Plateau, Southern Indian Ocean. <i>Deep Sea</i> <i>Research Part I: Oceanographic Research Papers</i> , 64, 71-77.
3462 3463 3464	Crespo, E. A., Pedraza, S. N., Dans, S. L., Svendsen, G. M., Degrati, M. & Coscarella, M. A. 2019. The Southwestern Atlantic Southern Right Whale, <i>Eubalaena Australis</i> , Population Is Growing but at a Decelerated Rate. <i>Marine Mammal Science</i> , 35, 93-107.
3465 3466	Cutler, D. R., Edwards, T. C., Beard, K. H., Cutler, A., Hess, K. T., Gibson, J. & Lawler, J. J. 2007. Random Forests for Classification in Ecology. <i>Ecology</i> , 88, 2783-2792.
3467 3468 3469	D'alelio, D., Hay Mele, B., Libralato, S., Ribera D'alcala, M. & Jordan, F. 2019. Rewiring and Indirect Effects Underpin Modularity Reshuffling in a Marine Food Web under Environmental Shifts. <i>Ecol Evol</i> , 9, 11631-11646.
3470 3471	Dahood, A., Watters, G. M. & De Mutsert, K. 2019. Using Sea-Ice to Calibrate a Dynamic Trophic Model for the Western Antarctic Peninsula. <i>PLoS One,</i> 14, e0214814.
3472 3473	Daufresne, M., Lengfellner, K. & Sommer, U. 2009. Global Warming Benefits the Small in Aquatic Ecosystems. <i>Proc Natl Acad Sci U S A</i> , 106, 12788-93.
3474 3475 3476 3477 3478	Degen, R., Aune, M., Bluhm, B. A., Cassidy, C., Kędra, M., Kraan, C., Vandepitte, L., Włodarska-Kowalczuk, M., Zhulay, I., Albano, P. G., Bremner, J., Grebmeier, J. M., Link, H., Morata, N., Nordström, M. C., Shojaei, M. G., Sutton, L. & Zuschin, M. 2018. Trait- Based Approaches in Rapidly Changing Ecosystems: A Roadmap to the Future Polar Oceans. <i>Ecological Indicators</i> , 91, 722-736.
3479 3480	Degen, R. & Faulwetter, S. 2019. The Arctic Traits Database – a Repository of Arctic Benthic Invertebrate Traits. <i>Earth Syst. Sci. Data</i> , 11, 301-322.
3481 3482 3483	Delmas, E., Brose, U., Gravel, D., Stouffer, D. B. & Poisot, T. 2017. Simulations of Biomass Dynamics in Community Food Webs. <i>Methods in Ecology and Evolution</i> , 8, 881-886.
3484 3485	Deppeler, S. L. & Davidson, A. T. 2017. Southern Ocean Phytoplankton in a Changing Climate. <i>Frontiers in Marine Science</i> , 4.
3486 3487 3488	Deutsch, C., Penn, J. L., Verberk, W. C. E. P., Inomura, K., Endress, MG. & Payne, J. L. 2022. Impact of Warming on Aquatic Body Sizes Explained by Metabolic Scaling from Microbes to Macrofauna. <i>Proceedings of the National Academy of Sciences</i> , 119.
3489 3490 3491	Dobashi, T., Iida, M. & Takemoto, K. 2018. Decomposing the Effects of Ocean Environments on Predator–Prey Body-Size Relationships in Food Webs. <i>Royal Society Open Science</i> , 5, 180707.
3492 3493	Doulcier, G. & Stouffer, D. 2015. Rnetcarto: Fast Network Modularity and Roles Computation by Simulated Annealing. <i>R package version 0.2.4</i> .

3494 Dunic, J. C. & Baum, J. K. 2017. Size Structuring and Allometric Scaling Relationships 3495 in Coral Reef Fishes. Journal of Animal Ecology, 86, 577-589. 3496 Dunne, J., Williams, R. & Martinez, N. 2004. Network Structure and Robustness of 3497 Marine Food Webs. Marine Ecology Progress Series, 273, 291-302. 3498 Eastman, J. T. 2004. The Nature of the Diversity of Antarctic Fishes. Polar Biology, 28, 3499 93-107. 3500 Emmerson, M. C. & Raffaelli, D. 2004. Predator-Prey Body Size, Interaction Strength and the Stability of a Real Food Web. Journal of Animal Ecology, 73, 399-409. 3501 3502 Eskuche-Keith, P., Hill, S. L., Hollyman, P., Taylor, M. L. & O'gorman, E. J. 2023. Trophic Structuring of Modularity Alters Energy Flow through Marine Food Webs. 3503 3504 Frontiers in Marine Science, 9. 3505 Eskuche-Keith, P., Hill, S. L., López-López, L., Rosenbaum, B., Saunders, R. A., Tarling, 3506 G. A. & O'gorman, E. J. 2024. Temperature Alters the Predator-Prey Size Relationships and Size-Selectivity of Southern Ocean Fish. Nature Communications, 15. 3507 3508 Ferry, L. A., Paig-Tran, E. M. & Gibb, A. C. 2015. Suction, Ram, and Biting: Deviations 3509 and Limitations to the Capture of Aquatic Prey. Integr Comp Biol, 55, 97-109. 3510 Fielding, S., Watkins, J. L., Trathan, P. N., Enderlein, P., Waluda, C. M., Stowasser, G., 3511 Tarling, G. A. & Murphy, E. J. 2014. Interannual Variability in Antarctic Krill (Euphausia Superba) Density at South Georgia, Southern Ocean: 1997-2013. ICES Journal of 3512 3513 Marine Science, 71, 2578-2588. Finlay, B. J. & Uhlig, G. 1981. Calorific and Carbon Values of Marine and Freshwater 3514 3515 Protozoa. Helgoländer Meeresuntersuchungen, 34, 401-412. 3516 Fisher, B. J., Poulton, A. J., Meredith, M. P., Baldry, K., Schofield, O. & Henley, S. F. 2024. Biogeochemistry of Climate Driven Shifts in Southern Ocean Primary Producers. 3517 Copernicus GmbH. 3518 3519 Fjeld, K., Tiller, R., Grimaldo, E., Grimsmo, L. & Standal, I.-B. 2023. Mesopelagics-3520 New Gold Rush or Castle in the Sky? Marine Policy, 147. 3521 Flensborg, L. C., Maureaud, A. A., Bravo, D. N. & Lindegren, M. 2023. An Indicator-3522 Based Approach for Assessing Marine Ecosystem Resilience. ICES Journal of Marine Science, 80, 1487-1499. 3523 3524 Flores, H., Atkinson, A., Kawaguchi, S., Krafft, B., Milinevsky, G., Nicol, S., Reiss, C., 3525 Tarling, G., Werner, R., Bravo Rebolledo, E., Cirelli, V., Cuzin-Roudy, J., Fielding, S., Van Franeker, J., Groeneveld, J., Haraldsson, M., Lombana, A., Marschoff, E., Meyer, B., 3526 Pakhomov, E., Van De Putte, A., Rombolá, E., Schmidt, K., Siegel, V., Teschke, M., 3527 Tonkes, H., Toullec, J., Trathan, P., Tremblay, N. & Werner, T. 2012. Impact of Climate 3528 3529 Change on Antarctic Krill. Marine Ecology Progress Series, 458, 1-19. Forget, N. L., Duplisea, D. E., Sardenne, F. & Mckindsey, C. W. 2020. Using Qualitative 3530 3531 Network Models to Assess the Influence of Mussel Culture on Ecosystem Dynamics. 3532 Ecological Modelling, 430.

- Forster, J., Hirst, A. G. & Atkinson, D. 2012. Warming-Induced Reductions in Body Size
  Are Greater in Aquatic Than Terrestrial Species. *Proceedings of the National Academy of Sciences*, 109, 19310-19314.
- Freer, J. J., Tarling, G. A., Collins, M. A., Partridge, J. C. & Genner, M. J. 2019.
  Predicting Future Distributions of Lanternfish, a Significant Ecological Resource within the Southern Ocean. *Diversity and Distributions*.
- Frelat, R., Kortsch, S., Kröncke, I., Neumann, H., Nordström, M. C., Olivier, P. E. N. &
  Sell, A. F. 2022. Food Web Structure and Community Composition: A Comparison across
  Space and Time in the North Sea. *Ecography*, 2022.
- Fretwell, P. T., Boutet, A. & Ratcliffe, N. 2023. Record Low 2022 Antarctic Sea Ice Led
  to Catastrophic Breeding Failure of Emperor Penguins. *Communications Earth & amp; Environment*, 4.
- Fricke, E. C., Hsieh, C., Middleton, O., Gorczynski, D., Cappello, C. D., Sanisidro, O.,
  Rowan, J., Svenning, J.-C. & Beaudrot, L. 2022. Collapse of Terrestrial Mammal Food
  Webs since the Late Pleistocene. *Science*, 377, 1008-1011.
- Fu, W., Randerson, J. T. & Moore, J. K. 2016. Climate Change Impacts on Net Primary
  Production (Npp) and Export Production (Ep) Regulated by Increasing Stratification and
  Phytoplankton Community Structure in the Cmip5 Models. *Biogeosciences*, 13, 51515170.
- Galiana, N., Barros, C., Braga, J., Ficetola, G. F., Maiorano, L., Thuiller, W., Montoya, J.
  M. & Lurgi, M. 2021. The Spatial Scaling of Food Web Structure across European
  Biogeographical Regions. *Ecography*, 44, 653-664.
- Gallucci, F., A. Christofoletti, R., Fonseca, G. & M. Dias, G. 2020. The Effects of Habitat
  Heterogeneity at Distinct Spatial Scales on Hard-Bottom-Associated Communities. *Diversity*, 12, 39.
- 3558Gårdmark, A. & Huss, M. 2020. Individual Variation and Interactions Explain Food Web3559Responses to Global Warming. *Philos Trans R Soc Lond B Biol Sci*, 375, 20190449.
- Garnesson, P., Mangin, A., Fanton D'andon, O., Demaria, J. & Bretagnon, M. 2019. The
  Cmems Globcolour Chlorophyll a Product Based on Satellite Observation: Multi-Sensor
  Merging and Flagging Strategies. *Ocean Science*, 15, 819-830.
- Gauzens, B., Barnes, A., Giling, D. P., Hines, J., Jochum, M., Lefcheck, J. S.,
  Rosenbaum, B., Wang, S. & Brose, U. 2019. Fluxweb : An R Package to Easily Estimate
  Energy Fluxes in Food Webs. *Methods in Ecology and Evolution*, 10, 270-279.
- Gauzens, B., Brose, U., Delmas, E. & Berti, E. 2023. Atnr: Allometric Trophic Network Models in R. *Methods in Ecology and Evolution*, 14, 2766-2773.
- Gauzens, B., Legendre, S., Lazzaro, X. & Lacroix, G. 2013. Food-Web Aggregation,
  Methodological and Functional Issues. *Oikos*, 122, 1606-1615.

- Gauzens, B., Rall, B. C., Mendonça, V., Vinagre, C. & Brose, U. 2020. Biodiversity of
  Intertidal Food Webs in Response to Warming across Latitudes. *Nature Climate Change*,
  10, 264-269.
- Gauzens, B., Rosenbaum, B., Kalinkat, G., Boy, T., Jochum, M., Kortsch, S., O'gorman,
  E. J. & Brose, U. 2024. Flexible Foraging Behaviour Increases Predator Vulnerability to
  Climate Change. *Nature Climate Change*, 14, 387-392.
- Gauzens, B., Thebault, E., Lacroix, G. & Legendre, S. 2015. Trophic Groups and
  Modules: Two Levels of Group Detection in Food Webs. *J R Soc Interface*, 12.
- Gibb, H., Stoklosa, J., Warton, D. I., Brown, A. M., Andrew, N. R. & Cunningham, S. A.
  2015. Does Morphology Predict Trophic Position and Habitat Use of Ant Species and
  Assemblages? *Oecologia*, 177, 519-531.
- Gibert, J. P. 2019. Temperature Directly and Indirectly Influences Food Web Structure.
   *Scientific Reports*, 9.
- Gibert, J. P. & Delong, J. P. 2014. Temperature Alters Food Web Body-Size Structure. *Biology Letters*, 10, 20140473.
- Gilabert, O. R., Navia, A. F., De La Cruz-Agüero, G., Molinero, J. C., Sommer, U. &
  Scotti, M. 2019. Body Size and Mobility Explain Species Centralities in the Gulf of
  California Food Web. *Community Ecology*, 20, 149-160.
- Gilbert, L., Jeanniard-Du-Dot, T., Authier, M., Chouvelon, T. & Spitz, J. 2023.
  Composition of Cetacean Communities Worldwide Shapes Their Contribution to Ocean
  Nutrient Cycling. *Nature Communications*, 14.
- Gillooly, J. F. 2000. Effect of Body Size and Temperature on Generation Time in
  Zooplankton. *Journal of Plankton Research*, 22, 241-251.
- Gingold, R., Mundo-Ocampo, M., Holovachov, O. & Rocha-Olivares, A. 2010. The Role
  of Habitat Heterogeneity in Structuring the Community of Intertidal Free-Living Marine
  Nematodes. *Marine Biology*, 157, 1741-1753.
- Gissi, E., Manea, E., Mazaris, A. D., Fraschetti, S., Almpanidou, V., Bevilacqua, S., Coll,
  M., Guarnieri, G., Lloret-Lloret, E., Pascual, M., Petza, D., Rilov, G., Schonwald, M.,
  Stelzenmüller, V. & Katsanevakis, S. 2021. A Review of the Combined Effects of Climate
  Change and Other Local Human Stressors on the Marine Environment. *Science of The Total Environment*, 755, 142564.
- Gjoni, V., Glazier, D. S., Wesner, J. S., Ibelings, B. W. & Thomas, M. K. 2023.
  Temperature, Resources and Predation Interact to Shape Phytoplankton Size–Abundance
  Relationships at a Continental Scale. *Global Ecology and Biogeography*, 32, 2006-2016.
- Gogina, M., Zettler, A. & Zettler, M. L. 2022. Weight-to-Weight Conversion Factors for
   Benthic Macrofauna: Recent Measurements from the Baltic and the North Seas. *Earth System Science Data*, 14, 1-4.

3607 Gougherty, A. V. & Clipp, H. L. 2024. Testing the Reliability of an Ai-Based Large 3608 Language Model to Extract Ecological Information from the Scientific Literature. npj 3609 Biodiversity, 3. 3610 Gravel, D., Albouy, C. & Thuiller, W. 2016. The Meaning of Functional Trait 3611 Composition of Food Webs for Ecosystem Functioning. Philosophical Transactions of the Royal Society B: Biological Sciences, 371, 20150268. 3612 Gravel, D., Poisot, T., Albouy, C., Velez, L. & Mouillot, D. 2013. Inferring Food Web 3613 3614 Structure from Predator-Prey Body Size Relationships. Methods in Ecology and 3615 Evolution, 4, 1083-1090. 3616 Green, S. J., Brookson, C. B., Hardy, N. A. & Crowder, L. B. 2022. Trait-Based Approaches to Global Change Ecology: Moving from Description to Prediction. 3617 3618 Proceedings of the Royal Society B: Biological Sciences, 289. 3619 Greenspoon, L., Krieger, E., Sender, R., Rosenberg, Y., Bar-On, Y. M., Moran, U., 3620 Antman, T., Meiri, S., Roll, U., Noor, E. & Milo, R. 2023. The Global Biomass of Wild Mammals. Proceedings of the National Academy of Sciences, 120. 3621 3622 Griffith, G. P., Hop, H., Vihtakari, M., Wold, A., Kalhagen, K. & Gabrielsen, G. W. 2019. Ecological Resilience of Arctic Marine Food Webs to Climate Change. Nature Climate 3623 Change, 9, 868-872. 3624 3625 Grilli, J., Rogers, T. & Allesina, S. 2016. Modularity and Stability in Ecological 3626 Communities. Nature Communications, 7, 12031. 3627 Guimerà, R. & Nunes Amaral, L. A. 2005. Functional Cartography of Complex Metabolic 3628 Networks. Nature, 433, 895-900. 3629 Guimera, R., Stouffer, D. B., Sales-Pardo, M., Leicht, E. A. & Newman, M. E. J. 2010. Origin of Compartmentalization in Food Webs. Ecology, 91, 2941-2951. 3630 3631 Gurney, L. J., Pakhomov, E. A. & Christensen, V. 2014. An Ecosystem Model of the 3632 Prince Edward Island Archipelago. Ecological Modelling, 294, 117-136. 3633 Gutt, J., Alvaro, M. C., Barco, A., Böhmer, A., Bracher, A., David, B., De Ridder, C., 3634 Dorschel, B., Eléaume, M., Janussen, D., Kersken, D., López-González, P. J., Martínez-3635 Baraldés, I., Schröder, M., Segelken-Voigt, A. & Teixidó, N. 2016. Macroepibenthic Communities at the Tip of the Antarctic Peninsula, an Ecological Survey at Different 3636 Spatial Scales. Polar Biology, 39, 829-849. 3637 3638 Hatton, I. A., Heneghan, R. F., Bar-On, Y. M. & Galbraith, E. D. 2021. The Global Ocean 3639 Size Spectrum from Bacteria to Whales. Science Advances, 7, eabh3732. 3640 Havermans, C., Auel, H., Hagen, W., Held, C., Ensor, N. S. & A. Tarling, G. 2019. 3641 Predatory Zooplankton on the Move: Themisto Amphipods in High-Latitude Marine 3642 Pelagic Food Webs. Elsevier. 3643 He, X., Liang, J., Zeng, G., Yuan, Y. & Li, X. 2019. The Effects of Interaction between 3644 Climate Change and Land-Use/Cover Change on Biodiversity-Related Ecosystem 3645 Services. Global Challenges, 3, 1800095.

3646 3647 3648	<ul><li>Heinichen, M., Mcmanus, M. C., Lucey, S. M., Aydin, K., Humphries, A., Innes-Gold, A.</li><li>&amp; Collie, J. 2022. Incorporating Temperature-Dependent Fish Bioenergetics into a Narragansett Bay Food Web Model. <i>Ecological Modelling</i>, 466, 109911.</li></ul>
3649 3650	Helfman, G. S., Collette, B. B., Facey, D. E. & Bowen, B. W. 2023. <i>The Diversity of Fishes: Biology, Evolution and Ecology</i> , John Wiley & Sons Ltd.
3651 3652 3653 3654	Henley, S. F., Cavan, E. L., Fawcett, S. E., Kerr, R., Monteiro, T., Sherrell, R. M., Bowie, A. R., Boyd, P. W., Barnes, D. K. A., Schloss, I. R., Marshall, T., Flynn, R. & Smith, S. 2020. Changing Biogeochemistry of the Southern Ocean and Its Ecosystem Implications. <i>Frontiers in Marine Science</i> , 7.
3655 3656 3657	Heymans, J. J., Coll, M., Libralato, S., Morissette, L. & Christensen, V. 2014. Global Patterns in Ecological Indicators of Marine Food Webs: A Modelling Approach. <i>PLoS ONE</i> , 9, e95845.
3658 3659 3660	Heymans, J. J., Coll, M., Link, J. S., Mackinson, S., Steenbeek, J., Walters, C. & Christensen, V. 2016. Best Practice in Ecopath with Ecosim Food-Web Models for Ecosystem-Based Management. <i>Ecological Modelling</i> , 331, 173-184.
3661 3662 3663	Higham, T. E. 2007. The Integration of Locomotion and Prey Capture in Vertebrates: Morphology, Behavior, and Performance. <i>Integrative and Comparative Biology</i> , 47, 82-95.
3664 3665 3666	Hill, S. L., Keeble, K., Atkinson, A. & Murphy, E. J. 2012. A Foodweb Model to Explore Uncertainties in the South Georgia Shelf Pelagic Ecosystem. <i>Deep Sea Research Part II: Topical Studies in Oceanography</i> , 59-60, 237-252.
3667 3668 3669	Hill, S. L., Murphy, E. J., Reid, K., Trathan, P. N. & Constable, A. J. 2006. Modelling Southern Ocean Ecosystems: Krill, the Food-Web, and the Impacts of Harvesting. <i>Biological Reviews</i> , 81, 581.
3670 3671 3672	Hill, S. L., Pinkerton, M. H., Ballerini, T., Cavan, E. L., Gurney, L. J., Martins, I. & Xavier, J. C. 2021. Robust Model-Based Indicators of Regional Differences in Food-Web Structure in the Southern Ocean. <i>Journal of Marine Systems</i> , 220.
3673 3674 3675 3676	Hill, S. L., Reid, K. & North, A. W. 2005. Recruitment of Mackerel Icefish (Champsocephalus Gunnari) at South Georgia Indicated by Predator Diets and Its Relationship with Sea Surface Temperature. <i>Canadian Journal of Fisheries and Aquatic</i> <i>Sciences</i> , 62, 2530-2537.
3677 3678 3679 3680 3681	Hines, J., Ebeling, A., Barnes, A. D., Brose, U., Scherber, C., Scheu, S., Tscharntke, T., Weisser, W. W., Giling, D. P., Klein, A. M. & Eisenhauer, N. 2019. Mapping Change in Biodiversity and Ecosystem Function Research: Food Webs Foster Integration of Experiments and Science Policy. <i>Mechanisms Underlying the Relationship between</i> <i>Biodiversity and Ecosystem Function</i> .
3682 3683	Hirst, A. G. & Kiørboe, T. 2002. Mortality of Marine Planktonic Copepods: Global Rates and Patterns. <i>Marine Ecology Progress Series</i> , 230, 195-209.
3684 3685	Hobson, E., S. 1979. Interactions between Piscivorous Fishes and Their Prey. <i>Predator-prey systems in fisheries management</i> , 231-242.

- Hogg, O. T., Barnes, D. K. A. & Griffiths, H. J. 2011. Highly Diverse, Poorly Studied and
  Uniquely Threatened by Climate Change: An Assessment of Marine Biodiversity on
  South Georgia's Continental Shelf. *PLoS ONE*, 6, e19795.
- Hollyman, P., Hill S.L., Gunn C., Keith P., Rodriguez, B. & Collins, M. A. 2023. Report
  of the Uk Groundfish Survey at South Georgia (Ccamlr Subarea 48.3) in February 2023.
  CCAMLR WG-FSA 2023/45.
- Hollyman, P. R., Hill, S. L., Laptikhovsky, V. V., Belchier, M., Gregory, S., Clement, A. &
  Collins, M. A. 2021. A Long Road to Recovery: Dynamics and Ecology of the Marbled
  Rockcod (Notothenia Rossii, Family: Nototheniidae) at South Georgia, 50 Years after
  Overexploitation. *ICES Journal of Marine Science*.
- Hop, H., Pearson, T., Hegseth, E. N., Kovacs, K. M., Wiencke, C., Kwasniewski, S.,
  Eiane, K., Mehlum, F., Gulliksen, B., Wlodarska-Kowalczuk, M., Lydersen, C.,
  Weslawski, J. M., Cochrane, S., Gabrielsen, G. W., Leakey, R. J. G., Lønne, O. J.,
  Zajaczkowski, M., Falk-Petersen, S., Kendall, M., Wängberg, S.-Å., Bischof, K.,
  Voronkov, A. Y., Kovaltchouk, N. A., Wiktor, J., Poltermann, M., Prisco, G. D., Papucci,
  C. & Gerland, S. 2002. The Marine Ecosystem of Kongsfjorden, Svalbard. *Polar Research*, 21, 167-208.
- Horn, S. & De La Vega, C. 2016. Relationships between Fresh Weight, Dry Weight, Ash
  Free Dry Weight, Carbon and Nitrogen Content for Selected Vertebrates. *Journal of Experimental Marine Biology and Ecology*, 481, 41-48.
- Horswill, C., Jackson, J. A., Medeiros, R., Nowell, R. W., Trathan, P. N. & O'connell, T.
  C. 2018. Minimising the Limitations of Using Dietary Analysis to Assess Foodweb
  Changes by Combining Multiple Techniques. *Ecological Indicators*, 94, 218-225.
- Hosie, G. W., Fukuchi, M. & Kawaguchi, S. 2003. Development of the Southern Ocean
  Continuous Plankton Recorder Survey. *Progress in Oceanography*, 58, 263-283.
- Hothorn, T., Bühlmann, P., Dudoit, S., Molinaro, A. & Van Der Laan, M. J. 2005.
  Survival Ensembles. *Biostatistics*, 7, 355-373.
- Hucke-Gaete, R., Osman, L. P., Moreno, C. A. & Torres, D. 2004. Examining Natural
  Population Growth from near Extinction: The Case of the Antarctic Fur Seal at the South
  Shetlands, Antarctica. *Polar Biology*, 27, 304-311.
- Hunt, B. P., Pakhomov, E. A. & Williams, R. 2011. Comparative Analysis of the 1980s
  and 2004 Macrozooplankton Composition and Distribution in the Vicinity of Kerguelen
  and Heard Islands: Seasonal Cycles and Oceanographic Forcing of Long-Term Change. *The Kerguelen plateau: marine ecosystem and fisheries*, 35, 79-92.
- Hunt, B. P. V., Espinasse, B., Pakhomov, E. A., Cherel, Y., Cotté, C., Delegrange, A. &
  Henschke, N. 2021. Pelagic Food Web Structure in High Nutrient Low Chlorophyll
  (Hnlc) and Naturally Iron Fertilized Waters in the Kerguelen Islands Region, Southern
  Ocean. *Journal of Marine Systems*, 224, 103625.
- Hutchison, C., Guichard, F., Legagneux, P., Gauthier, G., Bêty, J., Berteaux, D., Fauteux,
  D. & Gravel, D. 2020. Seasonal Food Webs with Migrations: Multi-Season Models
  Reveal Indirect Species Interactions in the Canadian Arctic Tundra. *Philosophical*

3727 Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences, 3728 378, 20190354. 3729 Ikeda, T. 2016. Routine Metabolic Rates of Pelagic Marine Fishes and Cephalopods as a 3730 Function of Body Mass, Habitat Temperature and Habitat Depth. Journal of Experimental 3731 Marine Biology and Ecology, 480, 74-86. 3732 Ives, A. R., Cardinale, B. J. & Snyder, W. E. 2004. A Synthesis of Subdisciplines: Predator-Prey Interactions, and Biodiversity and Ecosystem Functioning. Ecology Letters, 3733 3734 8, 102-116. 3735 Ives, A. R. & Carpenter, S. R. 2007. Stability and Diversity of Ecosystems. Science, 317, 3736 58-61. Iwc. 2024. Population Status [Online]. International Whaling Commission. Available: 3737 3738 https://iwc.int/about-whales/population-status [Accessed]. 3739 Jacob, U., Thierry, A., Brose, U., Arntz, W. E., Berg, S., Brey, T., Fetzer, I., Jonsson, T., Mintenbeck, K., Möllmann, C., Petchey, O. L., Riede, J. O. & Dunne, J. A. 2011. The 3740 3741 Role of Body Size in Complex Food Webs. The Role of Body Size in Multispecies 3742 Systems. 3743 Jacquet, C., Gounand, I. & Altermatt, F. 2020. How Pulse Disturbances Shape Size-3744 Abundance Pyramids. Ecology Letters, 23, 1014-1023. 3745 Janis, C. M. & Carrano, M. 1991. Scaling of Reproductive Turnover in Archosaurs and 3746 Mammals: Why Are Large Terrestrial Mammals So Rare? Annales Zoologici Fennici, 28, 3747 201-216. Jean-Michel, L., Eric, G., Romain, B.-B., Gilles, G., Angélique, M., Marie, D., Clément, 3748 3749 B., Mathieu, H., Olivier, L. G., Charly, R., Tony, C., Charles-Emmanuel, T., Florent, G., Giovanni, R., Mounir, B., Yann, D. & Pierre-Yves, L. T. 2021. The Copernicus Global 3750 3751 1/12° Oceanic and Sea Ice Glorys12 Reanalysis. Frontiers in Earth Science, 9. Jennings, S., Pinnegar, J. K., Polunin, N. V. C. & Boon, T. W. 2001. Weak Cross-Species 3752 Relationships between Body Size and Trophic Level Belie Powerful Size-Based Trophic 3753 Structuring in Fish Communities. Journal of Animal Ecology, 70, 934-944. 3754 3755 Jessop, M., Mcallen, R., O'halloran, J. & Kelly, T. 2011. Nutrient and Ecosystem Dynamics in Ireland's Only Marine Nature Reserve (Neidin). EPA STRIVE Programme 3756 2007–2013. Environmental Protection Agency. 3757 3758 Jochum, M., Barnes, A. D., Brose, U., Gauzens, B., Sünnemann, M., Amyntas, A. & Eisenhauer, N. 2021. For Flux's Sake: General Considerations for Energy-Flux 3759 3760 Calculations in Ecological Communities. Ecology and Evolution, 11, 12948-12969. Jochum, M. & Eisenhauer, N. 2022. Out of the Dark: Using Energy Flux to Connect 3761 3762 above- and Belowground Communities and Ecosystem Functioning. European Journal of 3763 Soil Science, 73. Jonsson, T. 2014. Trophic Links and the Relationship between Predator and Prey Body 3764 3765 Sizes in Food Webs. Community Ecology, 15, 54-64.

- Jonsson, T., Berg, S., Pimenov, A., Palmer, C. & Emmerson, M. 2015. The Reliability of *R*<sub>50</sub> as a Measure of Vulnerability of Food Webs to Sequential Species Deletions. *Oikos*,
  124, 446-457.
- Jonsson, T., Cohen, J. E. & Carpenter, S. R. 2005. Food Webs, Body Size, and Species
  Abundance in Ecological Community Description. *Advances in Ecological Research*, 36,
  1-84.
- Jonsson, T. & Ebenman, B. 1998. Effects of Predator–Prey Body Size Ratios on the
  Stability of Food Chains. *Journal of Theoretical Biology*, 193, 407-417.
- Kaufman, D. E., Friedrichs, M. a. M., Smith, W. O., Hofmann, E. E., Dinniman, M. S. &
  Hemmings, J. C. P. 2017. Climate Change Impacts on Southern Ross Sea Phytoplankton
  Composition, Productivity, and Export. *Journal of Geophysical Research: Oceans*, 122,
  2339-2359.
- Kavanagh, P., Newlands, N., Christensen, V. & Pauly, D. 2004. Automated Parameter
  Optimization for Ecopath Ecosystem Models. *Ecological Modelling*, 172, 141-149.
- Kawaguchi, S., Atkinson, A., Bahlburg, D., Bernard, K. S., Cavan, E. L., Cox, M. J., Hill,
  S. L., Meyer, B. & Veytia, D. 2024. Climate Change Impacts on Antarctic Krill Behaviour
  and Population Dynamics. *Nature Reviews Earth & Environment*, 5, 43-58.
- Keller, A., Ankenbrand, M. J., Bruelheide, H., Dekeyzer, S., Enquist, B. J., Erfanian, M.
  B., Falster, D. S., Gallagher, R. V., Hammock, J., Kattge, J., Leonhardt, S. D., Madin, J.
  S., Maitner, B., Neyret, M., Onstein, R. E., Pearse, W. D., Poelen, J. H., Salguero-Gomez,
  R., Schneider, F. D., Tóth, A. B. & Penone, C. 2023. Ten (Mostly) Simple Rules to
  Future-Proof Trait Data in Ecological and Evolutionary Sciences. *Methods in Ecology and Evolution*, 14, 444-458.
- Keramidas, I., Dimarchopoulou, D., Ofir, E., Scotti, M., Tsikliras, A. C. & Gal, G. 2023.
  Ecotrophic Perspective in Fisheries Management: A Review of Ecopath with Ecosim
  Models in European Marine Ecosystems. *Frontiers in Marine Science*, 10.
- Kiørboe, T. 2013. Zooplankton Body Composition. *Limnology and Oceanography*, 58, 1843-1850.
- Kiørboe, T., Visser, A., Andersen, K. H. & Browman, H. 2018. A Trait-Based Approach to
  Ocean Ecology. *ICES Journal of Marine Science*, 75, 1849-1863.
- Klein, E. S., Hill, S. L., Hinke, J. T., Phillips, T. & Watters, G. M. 2018. Impacts of Rising
  Sea Temperature on Krill Increase Risks for Predators in the Scotia Sea. *PLOS ONE*, 13,
  e0191011.
- Kock, K.-H., Reid, K., Croxall, J. & Nicol, S. 2007. Fisheries in the Southern Ocean: An
  Ecosystem Approach. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 362, 2333-2349.
- Kock, K., H.,, Barrera-Oro, E., Belchier, M., Collins, M. A., Duhamel, G., Hanchet, S.,
  Pshenichnov, L., Welsford, D. & Williams, R. 2012. The Role of Fish as Predators of
  Krill (Euphausia Superba) and Other Pelagic Resources in the Southern Ocean. *CCAMLR Science*, 19, 115-169.

- Kock, K. H., Wilhelms, S., Everson, I. & Gröger, J. 1994. Variations in the Diet
  Composition and Feeding Intensity of Mackerel Icefish Champsocephalus Gunnari at
  South Georgia (Antarctic). *Marine Ecology Progress Series*, 108, 43-57.
- Kordas, R. L., Pawar, S., Kontopoulos, D.-G., Woodward, G. & O'gorman, E. J. 2022.
  Metabolic Plasticity Can Amplify Ecosystem Responses to Global Warming. *Nature Communications*, 13.
- Kortsch, S., Frelat, R., Pecuchet, L., Olivier, P., Putnis, I., Bonsdorff, E., Ojaveer, H.,
  Jurgensone, I., Strāķe, S., Rubene, G., Krūze, Ē. & Nordström, M. C. 2021. Disentangling
  Temporal Food Web Dynamics Facilitates Understanding of Ecosystem Functioning. *Journal of Animal Ecology*, 90, 1205-1216.
- Kortsch, S., Primicerio, R., Aschan, M., Lind, S., Dolgov, A. V. & Planque, B. 2019.
  Food-Web Structure Varies Along Environmental Gradients in a High-Latitude Marine
  Ecosystem. *Ecography*, 42, 295-308.
- Kortsch, S., Primicerio, R., Fossheim, M., Dolgov, A. V. & Aschan, M. 2015. Climate
  Change Alters the Structure of Arctic Marine Food Webs Due to Poleward Shifts of
  Boreal Generalists. *Proceedings of the Royal Society B: Biological Sciences*, 282,
  20151546.
- Kouwenberg, J. H. M., Razouls, C. & Desreumaux, N. 2014. Chapter 6.6. Southern
  Ocean Pelagic Copepods. *In:* De Broyer C., Koubbi P., Griffiths H.J., Raymond B.,
  Udekem D'acoz C. D', Van De Putte A.P., Danis B., David B., Grant S., Gutt J., Held C.,
  Hosie G., Huettmann F., Post A. & Y., R.-C. (eds.) *Biogeographic Atlas of the Southern Ocean.* Cambridge: Scientific Committee on Antarctic Research.
- Kraan, C., Aarts, G., Piersma, T. & Dormann, C. F. 2013. Temporal Variability of
  Ecological Niches: A Study on Intertidal Macrobenthic Fauna. *Oikos*, 122, 754-760.
- Krafft, B. A., Lowther, A. & Krag, L. A. 2023. Bycatch in the Antarctic Krill
  (<I>Euphausia Superba</I>) Trawl Fishery. *Fisheries Management and Ecology*, 30, 154160.
- Kratina, P., Lecraw, R. M., Ingram, T. & Anholt, B. R. 2012. Stability and Persistence of
  Food Webs with Omnivory: Is There a General Pattern? *Ecosphere*, 3, 1-18.
- Kratina, P., Rosenbaum, B., Gallo, B., Horas, E. L. & O'gorman, E. J. 2022. The
  Combined Effects of Warming and Body Size on the Stability of Predator-Prey
  Interactions. *Frontiers in Ecology and Evolution*, 9.
- Krause, A. E., Frank, K. A., Mason, D. M., Ulanowicz, R. E. & Taylor, W. W. 2003.
  Compartments Revealed in Food-Web Structure. *Nature*, 426, 282-285.
- Krumhardt, K. M., Long, M. C., Sylvester, Z. T. & Petrik, C. M. 2022. Climate Drivers of
  Southern Ocean Phytoplankton Community Composition and Potential Impacts on
  Higher Trophic Levels. *Frontiers in Marine Science*, 9.
- La Mesa, M. & Eastman, J. T. 2011. Antarctic Silverfish: Life Strategies of a Key Species in the High-Antarctic Ecosystem. *Fish and Fisheries*, 13, 241-266.

3845 La Mesa, M., Piepenburg, D., Pineda-Metz, S. E. A., Riginella, E. & Eastman, J. T. 2019. 3846 Spatial Distribution and Habitat Preferences of Demersal Fish Assemblages in the Southeastern Weddell Sea (Southern Ocean). Polar Biology, 42, 1025-1040. 3847 3848 Ladds, M. A., Sibanda, N., Arnold, R. & Dunn, M. R. 2018. Creating Functional Groups 3849 of Marine Fish from Categorical Traits. PeerJ, 6, e5795. 3850 Laigle, I., Aubin, I., Digel, C., Brose, U., Boulangeat, I. & Gravel, D. 2017. Species Traits as Drivers of Food Web Structure. Oikos, 127, 316-326. 3851 Lavin, C. P., Gordó-Vilaseca, C., Stephenson, F., Shi, Z. & Costello, M. J. 2022. Warmer 3852 3853 Temperature Decreases the Maximum Length of Six Species of Marine Fishes, 3854 Crustacean, and Squid in New Zealand. Environmental Biology of Fishes. Laws, R. M. 1977. Seals and Whales of the Southern Ocean. Philosophical Transactions 3855 3856 of the Royal Society of London. B, Biological Sciences, 279, 81-96. 3857 Layman, C. A., Giery, S. T., Buhler, S., Rossi, R., Penland, T., Henson, M. N., Bogdanoff, A. K., Cove, M. V., Irizarry, A. D., Schalk, C. M. & Archer, S. K. 2015. A Primer on the 3858 History of Food Web Ecology: Fundamental Contributions of Fourteen Researchers. Food 3859 3860 Webs, 4, 14-24. 3861 Lazzaro, X., Lacroix, G., Gauzens, B., Gignoux, J. & Legendre, S. 2009. Predator Foraging Behaviour Drives Food-Web Topological Structure. Journal of Animal Ecology, 3862 78, 1307-1317. 3863 3864 Lefort, S., Aumont, O., Bopp, L., Arsouze, T., Gehlen, M. & Maury, O. 2015. Spatial and 3865 Body-Size Dependent Response of Marine Pelagic Communities to Projected Global 3866 Climate Change. Global Change Biology, 21, 154-164. 3867 Lemoine, N. P., Drews, W. A., Burkepile, D. E. & Parker, J. D. 2013. Increased Temperature Alters Feeding Behavior of a Generalist Herbivore. Oikos, 122, 1669-1678. 3868 3869 Liermann & Hilborn 2001. Depensation: Evidence, Models and Implications. Fish and 3870 *Fisheries*, 2, 33-58. Liu, K., Lin, H., He, X., Huang, Y., Li, Z., Lin, J., Mou, J., Zhang, S., Lin, L., Wang, J. & 3871 3872 Sun, J. 2019. Functional Trait Composition and Diversity Patterns of Marine 3873 Macrobenthos across the Arctic Bering Sea. Ecological Indicators, 102, 673-685. 3874 Lleonart, J., Salat, J. & Torres, G. J. 2000. Removing Allometric Effects of Body Size in 3875 Morphological Analysis. J Theor Biol, 205, 85-93. 3876 Lockyer, C. 1976. Body Weights of Some Species of Large Whales. ICES Journal of 3877 Marine Science, 36, 259-273. 3878 Lombarte, A., Olaso, I. & Bozzano, A. 2003. Ecomorphological Trends in the 3879 Artedidraconidae (Pisces: Perciformes: Notothenioidei) of the Weddell Sea. Antarctic Science, 15, 211-218. 3880

- López-López, L., Genner, M. J., Tarling, G. A., Saunders, R. A. & O'gorman, E. J. 2021.
  Ecological Networks in the Scotia Sea: Structural Changes across Latitude and Depth. *Ecosystems*.
- Lucey, S. M., Aydin, K. Y., Gaichas, S. K., Cadrin, S. X., Fay, G., Fogarty, M. J. & Punt,
  A. 2021. Evaluating Fishery Management Strategies Using an Ecosystem Model as an
  Operating Model. *Fisheries Research*, 234, 105780.
- Lucey, S. M., Gaichas, S. K. & Aydin, K. Y. 2020. Conducting Reproducible Ecosystem
  Modeling Using the Open Source Mass Balance Model Rpath. *Ecological Modelling*,
  427, 109057.
- Luiz, O. J., Crook, D. A., Kennard, M. J., Olden, J. D., Saunders, T. M., Douglas, M. M.,
  Wedd, D. & King, A. J. 2019. Does a Bigger Mouth Make You Fatter? Linking
  Intraspecific Gape Variability to Body Condition of a Tropical Predatory Fish. *Oecologia*,
  191, 579-585.
- Macnuson, J. J. & Heitz, J. G. 1971. Gill Raker Apparatus and Food Selectivity among
  Mackerels, Tunas, and Dolphins. *Fishery Bulletin*, 69.
- Main, C. E. & Collins, M. A. 2011. Diet of the Antarctic Starry Skate Amblyraja
  Georgiana (Rajidae, Chondrichthyes) at South Georgia (Southern Ocean). *Polar Biology*,
  34, 389-396.
- Main, C. E., Collins, M. A., Mitchell, R. & Belchier, M. 2009. Identifying Patterns in the
  Diet of Mackerel Icefish (Champsocephalus Gunnari) at South Georgia Using
  Bootstrapped Confidence Intervals of a Dietary Index. *Polar Biology*, 32, 569-581.
- Maldonado, M. T., Surma, S. & Pakhomov, E. A. 2016. Southern Ocean Biological Iron
  Cycling in the Pre-Whaling and Present Ecosystems. *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences*, 374, 20150292.
- Marina, T. I., Salinas, V., Cordone, G., Campana, G., Moreira, E., Deregibus, D., Torre,
  L., Sahade, R., Tatián, M., Barrera Oro, E., De Troch, M., Doyle, S., Quartino, M. L.,
  Saravia, L. A. & Momo, F. R. 2018. The Food Web of Potter Cove (Antarctica):
  Complexity, Structure and Function. *Estuarine, Coastal and Shelf Science*, 200, 141-151.
- Marina, T. I., Saravia, L. A. & Kortsch, S. 2024. New Insights into the Weddell Sea
  Ecosystem Applying a Quantitative Network Approach. *Ocean Science*, 20, 141-153.
- MarLIN 2006. Biotic Biological Traits Information Catalogue. Marine Biological
   Association of the United Kingdom. Available from <www.marlin.ac.uk/biotic>.
- 3913 Martinez, N. D. 1993. Effects of Resolution on Food Web Structure. *Oikos*, 66, 403-412.
- 3914 Martinez, N. D. 2020. Allometric Trophic Networks from Individuals to Socio-
- 3915 Ecosystems: Consumer–Resource Theory of the Ecological Elephant in the Room.
  3916 Frontiers in Ecology and Evolution, 8.
- Massom, R. A. & Stammerjohn, S. E. 2010. Antarctic Sea Ice Change and Variability –
  Physical and Ecological Implications. *Polar Science*, 4, 149-186.

3919 3920 3921 3922 3923 3924	Matsuoka, K., Skoglund, A., Roth, G., De Pomereu, J., Griffiths, H., Headland, R., Herried, B., Katsumata, K., Le Brocq, A., Licht, K., Morgan, F., Neff, P. D., Ritz, C., Scheinert, M., Tamura, T., Van De Putte, A., Van Den Broeke, M., Von Deschwanden, A., Deschamps-Berger, C., Van Liefferinge, B., Tronstad, S. & Melvær, Y. 2021. Quantarctica, an Integrated Mapping Environment for Antarctica, the Southern Ocean, and Sub-Antarctic Islands. <i>Environmental Modelling &amp; Software</i> , 140, 105015.
3925 3926	May, R. M. 1973. <i>Stability and Complexity in Model Ecosystems</i> , Princeton University Press.
3927 3928 3929	Mccann, K. & Hastings, A. 1997. Re–Evaluating the Omnivory–Stability Relationship in Food Webs. <i>Proceedings of the Royal Society of London. Series B: Biological Sciences</i> , 264, 1249-1254.
3930 3931	Mccann, K., Hastings, A. & Huxel, G. R. 1998. Weak Trophic Interactions and the Balance of Nature. <i>Nature</i> , 395, 794-798.
3932	Mccann, K. S. 2000. The Diversity-Stability Debate. Nature, 405, 228-233.
3933 3934	Mccann, K. S., Rasmussen, J. B. & Umbanhowar, J. 2005. The Dynamics of Spatially Coupled Food Webs. <i>Ecology Letters</i> , 8, 513-523.
3935 3936 3937	Mccann, K. S. & Rooney, N. 2009. The More Food Webs Change, the More They Stay the Same. <i>Philosophical Transactions of the Royal Society B: Biological Sciences</i> , 364, 1789-1801.
3938 3939 3940	Mccarthy, A. H., Peck, L. S. & Aldridge, D. C. 2022. Ship Traffic Connects Antarctica's Fragile Coasts to Worldwide Ecosystems. <i>Proceedings of the National Academy of Sciences</i> , 119, e2110303118.
3941 3942 3943 3944	Mccormack, S. A., Melbourne-Thomas, J., Trebilco, R., Blanchard, J. L. & Constable, A. 2020. Alternative Energy Pathways in Southern Ocean Food Webs: Insights from a Balanced Model of Prydz Bay, Antarctica. <i>Deep Sea Research Part II: Topical Studies in Oceanography</i> , 174.
3945 3946 3947 3948	Mccormack, S. A., Melbourne-Thomas, J., Trebilco, R., , Griffith, G., Hill, S. L., Hoover, C., Johnston, N. M., Marina, T. I., Murphy, E. J., Pakhomov, E. A. & Pinkerton, M. 2021a. Southern Ocean Food Web Modelling: Progress, Prognoses, and Future Priorities for Research and Policy Makers. <i>Frontiers in Ecology and Evolution</i> , 626.
3949 3950 3951	Mccormack, S. A., Melbourne-Thomas, J., Trebilco, R., Blanchard, J. L., Raymond, B. & Constable, A. 2021b. Decades of Dietary Data Demonstrate Regional Food Web Structures in the Southern Ocean. <i>Ecology and Evolution</i> , 11, 227-241.
3952 3953	Mckenna, J. E. 1991. Trophic Relationships within the Antarctic Demersal Fish Community of South Georgia Island. <i>Fishery Bulletin</i> , 89, 643-654.
3954 3955 3956 3957	Mcmahon, K. W., Michelson, C. I., Hart, T., Mccarthy, M. D., Patterson, W. P. & Polito, M. J. 2019. Divergent Trophic Responses of Sympatric Penguin Species to Historic Anthropogenic Exploitation and Recent Climate Change. <i>Proceedings of the National Academy of Sciences</i> , 116, 25721-25727.

Melbourne-Thomas, J., Constable, A., Wotherspoon, S. & Raymond, B. 2013. Testing
Paradigms of Ecosystem Change under Climate Warming in Antarctica. *PLoS ONE*, 8,
e55093.

Meredith, M., Sommerkorn, M., Cassotta, S., Derksen, C., Ekaykin, A., Hollowed, A.,
Kofinas, G., Mackintosh, A., Melbourne-Thomas, J., Muelbert, M. M. C., Ottersen, G.,
Pritchard, H. & Schuur, E. a. G. 2019. Polar Regions. *In:* PöRtner H.-O., Roberts, D. C.,
Masson-Delmotte, V., Zhai, P., Tignor, M., Poloczanska, E., Mintenbeck, K., AlegríA, A.,
Nicolai, M., Okem, A., Petzold, J., Rama, B. & Weyer, N. M. (eds.) *Ipcc Special Report on the Ocean and Cryosphere in a Changing Climate*. Cambridge, UK and New York,
NY, USA: Cambridge University Press.

- Meredith, M. P. & King, J. C. 2005. Rapid Climate Change in the Ocean West of the
  Antarctic Peninsula During the Second Half of the 20th Century. *Geophysical Research Letters*, 32, n/a-n/a.
- 3971 Meyer, B., Atkinson, A., Bernard, K. S., Brierley, A. S., Driscoll, R., Hill, S. L.,
- 3972 Marschoff, E., Maschette, D., Perry, F. A., Reiss, C. S., Rombolá, E., Tarling, G. A.,
- Thorpe, S. E., Trathan, P. N., Zhu, G. & Kawaguchi, S. 2020. Successful EcosystemBased Management of Antarctic Krill Should Address Uncertainties in Krill Recruitment,
  Behaviour and Ecological Adaptation. *Communications Earth & Computer Review Processing*, 1.
- Michael, K., Suberg, L. A., Wessels, W., Kawaguchi, S. & Meyer, B. 2021. Facing
  Southern Ocean Warming: Temperature Effects on Whole Animal Performance of
  Antarctic Krill (Euphausia Superba). *Zoology*, 146, 125910.
- Miller, D. G. M. 1991. Exploitation of Antarctic Marine Living Resources: A Brief
  History and a Possible Approach to Managing the Krill Fishery. *South African Journal of Marine Science*, 10, 321-339.
- Mintenbeck, K. 2017. Impacts of Climate Change on the Southern Ocean. *Climate Change Impacts on Fisheries and Aquaculture.*
- Montoya, D., Yallop, M. L. & Memmott, J. 2015. Functional Group Diversity Increases
  with Modularity in Complex Food Webs. *Nature Communications*, 6, 7379.
- Montoya, J. M., Woodward, G., Emmerson, M. C. & Sole, R. V. 2009. Press Perturbations
  and Indirect Effects in Real Food Webs. *Ecology*, 90, 2426-33.
- Morales-Castilla, I., Matias, M. G., Gravel, D. & Araujo, M. B. 2015. Inferring Biotic
  Interactions from Proxies. *Trends Ecol Evol*, 30, 347-56.
- Mori, M. & Butterworth, D. S. 2006. A First Step Towards Modelling the Krill–Predator
  Dynamics of the Antarctic Ecosystem. *CCAMLR Science*, 13, 217-277.
- Morley, S. A., Abele, D., Barnes, D. K. A., Cárdenas, C. A., Cotté, C., Gutt, J., Henley, S.
  F., Höfer, J., Hughes, K. A., Martin, S. M., Moffat, C., Raphael, M., Stammerjohn, S. E.,
  Suckling, C. C., Tulloch, V. J. D., Waller, C. L. & Constable, A. J. 2020. Global Drivers
  on Southern Ocean Ecosystems: Changing Physical Environments and Anthropogenic
  Pressures in an Earth System. *Frontiers in Marine Science*, 7.

3997 Morley, S. A., Belchier, M., Sands, C., Barnes, D. K. A. & Peck, L. S. 2014. Geographic 3998 Isolation and Physiological Mechanisms Underpinning Species Distributions at the Range 3999 Limit Hotspot of South Georgia. Reviews in Fish Biology and Fisheries, 24, 485-492. 4000 Morley, S. A., Griffiths, H. J., Barnes, D. K. A. & Peck, L. S. 2010. South Georgia: A Key 4001 Location for Linking Physiological Capacity to Distributional Changes in Response to Climate Change. Antarctic Science, 22, 774-781. 4002 4003 Mougi, A. 2018. Spatial Compartmentation and Food Web Stability. Scientific Reports, 8. 4004 Murphy, E. J., Cavanagh, R. D., Hofmann, E. E., Hill, S. L., Constable, A. J., Costa, D. P., 4005 Pinkerton, M. H., Johnston, N. M., Trathan, P. N., Klinck, J. M., Wolf-Gladrow, D. A., 4006 Daly, K. L., Maury, O. & Doney, S. C. 2012. Developing Integrated Models of Southern 4007 Ocean Food Webs: Including Ecological Complexity, Accounting for Uncertainty and the 4008 Importance of Scale. Progress in Oceanography, 102, 74-92. 4009 Murphy, E. J., Watkins, J. L., Trathan, P. N., Reid, K., Meredith, M. P., Thorpe, S. E., 4010 Johnston, N. M., Clarke, A., Tarling, G. A., Collins, M. A., Forcada, J., Shreeve, R. S., Atkinson, A., Korb, R., Whitehouse, M. J., Ward, P., Rodhouse, P. G., Enderlein, P., Hirst, 4011 4012 A. G., Martin, A. R., Hill, S. L., Staniland, I. J., Pond, D. W., Briggs, D. R., Cunningham, N. J. & Fleming, A. H. 2007. Spatial and Temporal Operation of the Scotia Sea 4013 4014 Ecosystem: A Review of Large-Scale Links in a Krill Centred Food Web. Philosophical 4015 Transactions of the Royal Society B: Biological Sciences, 362, 113-148. 4016 Nelson, D., Benstead, J. P., Huryn, A. D., Cross, W. F., Hood, J. M., Johnson, P. W., Junker, J. R., Gíslason, G. M. & Ólafsson, J. S. 2020. Thermal Niche Diversity and 4017 Trophic Redundancy Drive Neutral Effects of Warming on Energy Flux through a Stream 4018 4019 Food Web. *Ecology*, 101, e02952. 4020 Newbold, T. 2018. Future Effects of Climate and Land-Use Change on Terrestrial 4021 Vertebrate Community Diversity under Different Scenarios. Proceedings of the Royal Society B: Biological Sciences, 285, 20180792. 4022 4023 Newman, M. E. 2006. Modularity and Community Structure in Networks. PNAS, 103, 4024 8577-8582. 4025 Newman, M. E. & Girvan, M. 2004. Finding and Evaluating Community Structure in 4026 Networks. Phys Rev E Stat Nonlin Soft Matter Phys, 69, 026113. 4027 Nicol, S., Bowie, A., Jarman, S., Lannuzel, D., Meiners, K. M. & Van Der Merwe, P. 4028 2010. Southern Ocean Iron Fertilization by Baleen Whales and Antarctic Krill. Fish and 4029 Fisheries, 11, 203-209. 4030 Nielsen, J. M., Clare, E. L., Hayden, B., Brett, M. T. & Kratina, P. 2017. Diet Tracing in Ecology: Method Comparison and Selection. Methods in Ecology and Evolution, 9, 278-4031 4032 291. 4033 Nikolaou, A. & Katsanevakis, S. 2023. Marine Extinctions and Their Drivers. Regional 4034 Environmental Change, 23.

- Nisbet, R. M., Jusup, M., Klanjscek, T. & Pecquerie, L. 2012. Integrating Dynamic
  Energy Budget (Deb) Theory with Traditional Bioenergetic Models. *Journal of Experimental Biology*, 215, 892-902.
- Niu, J., Huss, M., Vasemägi, A. & Gårdmark, A. 2023. Decades of Warming Alters
  Maturation and Reproductive Investment in Fish. *Ecosphere*, 14.
- 4040 O'connor, M. I., Piehler, M. F., Leech, D. M., Anton, A. & Bruno, J. F. 2009. Warming
  4041 and Resource Availability Shift Food Web Structure and Metabolism. *PLoS Biology*, 7,
  4042 e1000178.
- 4043 O'gorman, E. J. & Emmerson, M. C. 2009. Perturbations to Trophic Interactions and the
  4044 Stability of Complex Food Webs. *Proceedings of the National Academy of Sciences*, 106,
  4045 13393-13398.
- 4046 O'gorman, E. J. & Emmerson, M. C. 2010. Manipulating Interaction Strengths and the
  4047 Consequences for Trivariate Patterns in a Marine Food Web. *Advances in Ecological*4048 *Research*, 42, 301-419.
- O'gorman, E. J., Ólafsson, Ó. P., Demars, B. O. L., Friberg, N., Guðbergsson, G.,
  Hannesdóttir, E. R., Jackson, M. C., Johansson, L. S., Mclaughlin, Ó. B., Ólafsson, J. S.,
  Woodward, G. & Gíslason, G. M. 2016. Temperature Effects on Fish Production across a
  Natural Thermal Gradient. *Global Change Biology*, 22, 3206-3220.
- 4053 O'gorman, E. J., Jacob, U., Jonsson, T. & Emmerson, M. C. 2010. Interaction Strength,
  4054 Food Web Topology and the Relative Importance of Species in Food Webs. *Journal of*4055 *Animal Ecology*, 79, 682-692.
- 4056 O'gorman, E. J., Petchey, O. L., Faulkner, K. J., Gallo, B., Gordon, T. a. C., Neto4057 Cerejeira, J., Ólafsson, J. S., Pichler, D. E., Thompson, M. S. A. & Woodward, G. 2019. A
  4058 Simple Model Predicts How Warming Simplifies Wild Food Webs. *Nature Climate*4059 *Change*, 9, 611-616.
- 4060 O'gorman, E. J., Zhao, L., Pichler, D. E., Adams, G., Friberg, N., Björn, Seeney, A.,
  4061 Zhang, H., Reuman, D. C. & Woodward, G. 2017. Unexpected Changes in Community
  4062 Size Structure in a Natural Warming Experiment. *Nature Climate Change*, 7, 659-663.
- Ockendon, N., Baker, D. J., Carr, J. A., White, E. C., Almond, R. E. A., Amano, T.,
  Bertram, E., Bradbury, R. B., Bradley, C., Butchart, S. H. M., Doswald, N., Foden, W.,
  Gill, D. J. C., Green, R. E., Sutherland, W. J., Tanner, E. V. J. & Pearce-Higgins, J. W.
  2014. Mechanisms Underpinning Climatic Impacts on Natural Populations: Altered
  Species Interactions Are More Important Than Direct Effects. *Global Change Biology*,
  20, 2221-2229.
- 4069 Ojeda, F. P. 1986. Morphological Characterization of the Alimentary Tract of Antarctic
  4070 Fishes and Its Relation to Feeding Habits. *Polar Biology*, 5, 125-128.
- 4071 Olden, J. D., Leroy Poff, N., Douglas, M. R., Douglas, M. E. & Fausch, K. D. 2004.
  4072 Ecological and Evolutionary Consequences of Biotic Homogenization. *Trends Ecol Evol*, 19, 18-24.

4074 Olivier, P., Frelat, R., Bonsdorff, E., Kortsch, S., Kröncke, I., Möllmann, C., Neumann, 4075 H., Sell, A. F. & Nordström, M. C. 2019. Exploring the Temporal Variability of a Food 4076 Web Using Long-Term Biomonitoring Data. Ecography, 42, 2107-2121. 4077 Pacheco, A. S., González, M. T., Bremner, J., Oliva, M., Heilmayer, O., Laudien, J. & 4078 Riascos, J. M. 2011. Functional Diversity of Marine Macrobenthic Communities from 4079 Sublittoral Soft-Sediment Habitats Off Northern Chile. Helgoland Marine Research, 65, 4080 413-424. 4081 Padovani, L. N., Viñas, M. D., Sánchez, F. & Mianzan, H. 2012. Amphipod-Supported 4082 Food Web: Themisto Gaudichaudii, a Key Food Resource for Fishes in the Southern Patagonian Shelf. Journal of Sea Research, 67, 85-90. 4083 4084 Paine, R. T. 1992. Food-Web Analysis through Field Measurement of Per Capita 4085 Interaction Strength. Nature, 355, 73-75. 4086 Park, J., Kuzminov, F. I., Bailleul, B., Yang, E. J., Lee, S., Falkowski, P. G. & Gorbunov, 4087 M. Y. 2017. Light Availability Rather Than Fe Controls the Magnitude of Massive Phytoplankton Bloom in the Amundsen Sea Polynyas, Antarctica. Limnology and 4088 4089 Oceanography, 62, 2260-2276. 4090 Parkinson, C. L. 2019. A 40-Y Record Reveals Gradual Antarctic Sea Ice Increases 4091 Followed by Decreases at Rates Far Exceeding the Rates Seen in the Arctic. Proceedings 4092 of the National Academy of Sciences, 116, 14414-14423. 4093 Pauli, N.-C., Metfies, K., Pakhomov, E. A., Neuhaus, S., Graeve, M., Wenta, P., Flintrop, 4094 C. M., Badewien, T. H., Iversen, M. H. & Meyer, B. 2021. Selective Feeding in Southern 4095 Ocean Key Grazers-Diet Composition of Krill and Salps. Communications Biology, 4. 4096 Pauly, D. 1989. Food Consumption by Tropical and Temperate Fish Populations: Some Generalizations. Journal of Fish Biology, 35, 11-20. 4097 4098 Peck, L. S., Morley, S. A., Richard, J. & Clark, M. S. 2014. Acclimation and Thermal 4099 Tolerance in Antarctic Marine Ectotherms. Journal of Experimental Biology, 217, 16-22. 4100 Pellissier, L., Albouy, C., Bascompte, J., Farwig, N., Graham, C., Loreau, M., Maglianesi, 4101 M. A., Melián, C. J., Pitteloud, C., Roslin, T., Rohr, R., Saavedra, S., Thuiller, W., 4102 Woodward, G., Zimmermann, N. E. & Gravel, D. 2018. Comparing Species Interaction 4103 Networks Along Environmental Gradients. Biological Reviews, 93, 785-800. Pérez-Matus, A., Ospina-Alvarez, A., Camus, P., Carrasco, S., Fernandez, M., Gelcich, S., 4104 4105 Godoy, N., Ojeda, F., Pardo, L., Rozbaczylo, N., Subida, M., Thiel, M., Wieters, E. & 4106 Navarrete, S. 2017. Temperate Rocky Subtidal Reef Community Reveals Human Impacts 4107 across the Entire Food Web. Marine Ecology Progress Series, 567, 1-16. Petchey, O. L. 2000. Prey Diversity, Prey Composition, and Predator Population 4108 4109 Dynamics in Experimental Microcosms. J Anim Ecol, 69, 874-882. 4110 Petchey, O. L., Beckerman, A. P., Riede, J. O. & Warren, P. H. 2008. Size, Foraging, and 4111 Food Web Structure. Proceedings of the National Academy of Sciences, 105, 4191-4196.

4112 Petrik, C. M., Stock, C. A., Andersen, K. H., Van Denderen, P. D. & Watson, J. R. 2020. Large Pelagic Fish Are Most Sensitive to Climate Change Despite Pelagification of 4113 4114 Ocean Food Webs. Frontiers in Marine Science, 7. 4115 Piatkowski, U., Rodhouse, P. G., White, M. G., Bone, D. G. & Symon, C. 1994. Nekton 4116 Community of the Scotia Sea as Sampled by the Rmt 25 During Austral Summer. marine 4117 Ecology Progress Series, 112, 13-28. 4118 Pichler, M., Boreux, V., Klein, A. M., Schleuning, M. & Hartig, F. 2020. Machine Learning Algorithms to Infer Trait-Matching and Predict Species Interactions in 4119 Ecological Networks. Methods in Ecology and Evolution, 11, 281-293. 4120 4121 Pietzsch, B. W., Schmidt, A., Groeneveld, J., Bahlburg, D., Meyer, B. & Berger, U. 2023. The Impact of Salps (Salpa Thompsoni) on the Antarctic Krill Population (Euphausia 4122 Superba): An Individual-Based Modelling Study. Ecological Processes, 12. 4123 4124 Pigot, A. L., Trisos, C. H. & Tobias, J. A. 2016. Functional Traits Reveal the Expansion 4125 and Packing of Ecological Niche Space Underlying an Elevational Diversity Gradient in Passerine Birds. Proceedings of the Royal Society B: Biological Sciences, 283, 20152013. 4126 4127 Pineda-Metz, S. E. A. 2020. Benthos-Pelagos Interconnectivity: Antarctic Shelf Examples. Springer International Publishing. 4128 4129 Pineda Metz, S. E. 2019. Benthic Communities of the Weddell Sea: Past, Present and 4130 Future. 4131 Pinheiro, J., Bates, D. & Team, R. C. 2023. Nlme: Linear and Nonlinear Mixed Effects 4132 Models. R Package Version 3.1-162. https://CRAN.R-project.org/package=nlme. Pinkas, L., Oliphant, M. S. & Iverson, I. L. K. 1970. Food Habits of Albacore, Bluefin 4133 4134 Tuna, and Bonito in California Waters. Fish Bulletin 152. 4135 Pinkerton, M. & Bradford-Grieve, J. M. 2010. A Balanced Model of the Food Web of the Ross Sea, Antarctica. CCAMLR Science, 17, 1-31. 4136 4137 Pinkerton, M. H., Boyd, P. W., Deppeler, S., Hayward, A., Höfer, J. & Moreau, S. 2021. 4138 Evidence for the Impact of Climate Change on Primary Producers in the Southern Ocean. Frontiers in Ecology and Evolution, 9. 4139 4140 Pinkerton, M. H. & Bradford-Grieve, J. M. 2014. Characterizing Foodweb Structure to 4141 Identify Potential Ecosystem Effects of Fishing in the Ross Sea, Antarctica. ICES Journal 4142 of Marine Science, 71, 1542-1553. Pinnegar, J. K., Blanchard, J. L., Mackinson, S., Scott, R. D. & Duplisea, D. E. 2005. 4143 4144 Aggregation and Removal of Weak-Links in Food-Web Models: System Stability and 4145 Recovery from Disturbance. Ecological Modelling, 184, 229-248. 4146 Plagányi, É. E., Blamey, L. K., Rogers, J. G. D. & Tulloch, V. J. D. 2022. Playing the Detective: Using Multispecies Approaches to Estimate Natural Mortality Rates. Fisheries 4147 4148 Research, 249.

4149 Plagányi, É. E. & Butterworth, D. S. 2004. A Critical Look at the Potential of Ecopath 4150 with Ecosim to Assist in Practical Fisheries Management. African Journal of Marine Science, 26, 261-287. 4151 Podder, A., Panja, S., Chaudhuri, A., Roy, A., Biswas, M. & Homechaudhuri, S. 2021. 4152 Patterns of Morphological Traits Shaping the Feeding Guilds in the Intertidal Mudflat 4153 Fishes of the Indian Sundarbans. Journal of Fish Biology, 99, 1010-1031. 4154 4155 Polazzo, F., Hermann, M., Crettaz-Minaglia, M. & Rico, A. 2023. Impacts of Extreme 4156 Climatic Events on Trophic Network Complexity and Multidimensional Stability. 4157 Ecology, 104. 4158 Post, D. M. 2002. The Long and Short of Food-Chain Length. Trends in Ecology & 4159 Evolution, 17, 269-277. 4160 Potapov, A. M., Brose, U., Scheu, S. & Tiunov, A. V. 2019. Trophic Position of 4161 Consumers and Size Structure of Food Webs across Aquatic and Terrestrial Ecosystems. 4162 Am Nat, 194, 823-839. 4163 Pouvreau, S., Bourles, Y., Lefebvre, S., Gangnery, A. & Alunno-Bruscia, M. 2006. 4164 Application of a Dynamic Energy Budget Model to the Pacific Oyster, Crassostrea Gigas, Reared under Various Environmental Conditions. Journal of Sea Research, 56, 156-167. 4165 Queirós, J. P., Borras-Chavez, R., Friscourt, N., Groß, J., Lewis, C. B., Mergard, G. & 4166 O'brien, K. 2024. Southern Ocean Food-Webs and Climate Change: A Short Review and 4167 4168 Future Directions. PLOS Climate, 3, e0000358. 4169 Queiros, Q., Mckenzie, D. J., Dutto, G., Killen, S., Saraux, C. & Schull, Q. 2024. Fish 4170 Shrinking, Energy Balance and Climate Change. Science of The Total Environment, 906, 4171 167310. 4172 Quetin, L. B., Ross, R. M., Fritsen, C. H. & Vernet, M. 2007. Ecological Responses of 4173 Antarctic Krill to Environmental Variability: Can We Predict the Future? Antarctic 4174 Science, 19, 253-266. 4175 Rall, B. C., Brose, U., Hartvig, M., Kalinkat, G., Schwarzmüller, F., Vucic-Pestic, O. & 4176 Petchey, O. L. 2012. Universal Temperature and Body-Mass Scaling of Feeding Rates. 4177 Philosophical Transactions of the Royal Society B: Biological Sciences, 367, 2923-2934. 4178 Rall, B. C., Vucic-Pestic, O., Ehnes, R. B., Emmerson, M. & Brose, U. 2009. Temperature, Predator-Prey Interaction Strength and Population Stability. Global Change 4179 4180 Biology, 16, 2145-2157. 4181 Ramírez, F., Davenport, T. L. & Mojica, J. I. 2015. Dietary–Morphological Relationships of Nineteen Fish Species from an Amazonian Terra Firme Blackwater Stream in 4182 4183 Colombia. Limnologica, 52, 89-102. 4184 Ratnarajah, L., Bowie, A. R., Lannuzel, D., Meiners, K. M. & Nicol, S. 2014. The 4185 Biogeochemical Role of Baleen Whales and Krill in Southern Ocean Nutrient Cycling. 4186 *PLoS ONE*, 9, e114067.

4187 4188 4189 4190	Ratnarajah, L., Melbourne-Thomas, J., Marzloff, M. P., Lannuzel, D., Meiners, K. M., Chever, F., Nicol, S. & Bowie, A. R. 2016. A Preliminary Model of Iron Fertilisation by Baleen Whales and Antarctic Krill in the Southern Ocean: Sensitivity of Primary Productivity Estimates to Parameter Uncertainty. <i>Ecological Modelling</i> , 320, 203-212.
4191 4192	R Core Team 2023. R: Language and Environment for Statistical Computing. <i>R Foundation for Statistical Computing, Vienna, Austria.</i> https://www.R-project.org/.
4193 4194 4195 4196	Reid, K., Hill, S. L., Diniz, T. C. D. & Collins, M. A. 2005. Mackerel Icefish Champsocephalus Gunnari in the Diet of Upper Trophic Level Predators at South Georgia: Implications for Fisheries Management. <i>Marine Ecology Progress Series</i> , 305, 153-161.
4197 4198 4199	Reid, W. D. K., Clarke, S., Collins, M. A. & Belchier, M. 2007. Distribution and Ecology of Chaenocephalus Aceratus (Channichthyidae) around South Georgia and Shag Rocks (Southern Ocean). <i>Polar Biology</i> , 30, 1523-1533.
4200 4201 4202 4203	Reisinger, R. R., Landman, M., Mgibantaka, N., Smale, M. J., Bester, M. N., De Bruyn, P. J. N. & Pistorius, P. A. 2018. Overlap and Temporal Variation in the Diets of Sympatric Antarctic and Subantarctic Fur Seals (Arctocephalus Spp.) at Marion Island, Prince Edward Islands. <i>Polar Research</i> , 37, 1451142.
4204 4205 4206	Retelletti Brogi, S., Jung, J. Y., Ha, S. Y. & Hur, J. 2019. Seasonal Differences in Dissolved Organic Matter Properties and Sources in an Arctic Fjord: Implications for Future Conditions. <i>Sci Total Environ</i> , 694, 133740.
4207 4208 4209	Reum, J. C. P., Holsman, K. K., Aydin, K. Y., Blanchard, J. L. & Jennings, S. 2019. Energetically Relevant Predator-Prey Body Mass Ratios and Their Relationship with Predator Body Size. <i>Ecology and Evolution</i> , 9, 201-211.
4210 4211 4212	Rezende, E. L., Albert, E. M., Fortuna, M. A. & Bascompte, J. 2009. Compartments in a Marine Food Web Associated with Phylogeny, Body Mass, and Habitat Structure. <i>Ecol Lett</i> , 12, 779-88.
4213 4214 4215 4216	Richardson, J., Wood, A. G., Neil, A., Nowacek, D. & Moore, M. 2012. Changes in Distribution, Relative Abundance, and Species Composition of Large Whales around South Georgia from Opportunistic Sightings: 1992 to 2011. <i>Endangered Species Research</i> , 19, 149-156.
4217 4218 4219	Riemer, K., Anderson-Teixeira, K. J., Smith, F. A., Harris, D. J. & Ernest, S. K. M. 2018. Body Size Shifts Influence Effects of Increasing Temperatures on Ectotherm Metabolism. <i>Global Ecology and Biogeography</i> , 27, 958-967.
4220 4221 4222	Rivera-Hutinel, A., Bustamante, R. O., Marín, V. H. & Medel, R. 2012. Effects of Sampling Completeness on the Structure of Plant–Pollinator Networks. <i>Ecology</i> , 93, 1593-1603.
4223 4224 4225	Rodriguez, I. D., Marina, T. I., Schloss, I. R. & Saravia, L. A. 2022. Marine Food Webs Are More Complex but Less Stable in Sub-Antarctic (Beagle Channel, Argentina) Than in Antarctic (Potter Cove, Antarctic Peninsula) Regions. <i>Mar Environ Res</i> , 174, 105561.

4226 Roman, J., Estes, J. A., Morissette, L., Smith, C., Costa, D., Mccarthy, J., Nation, J., 4227 Nicol, S., Pershing, A. & Smetacek, V. 2014. Whales as Marine Ecosystem Engineers. 4228 Frontiers in Ecology and the Environment, 12, 377-385. 4229 Rooney, N. & Mccann, K. S. 2012. Integrating Food Web Diversity, Structure and 4230 Stability. Trends in Ecology & Evolution, 27, 40-46. 4231 Rooney, N., Mccann, K. S. & Moore, J. C. 2008. A Landscape Theory for Food Web 4232 Architecture. Ecology Letters, 11, 867-881. 4233 Rose, K. A., Holsman, K., Nye, J. A., Markowitz, E. H., Banha, T. N., Bueno-Pardo, J., 4234 Deslauriers, D., Fulton, E. A., Huebert, K. B. & Huret, M. 2024. Advancing 4235 Bioenergetics-Based Modeling to Improve Climate Change Projections of Marine Ecosystems. Marine Ecology Progress Series, 732, 193-221. 4236 4237 Rossi, L., Sporta Caputi, S., Calizza, E., Careddu, G., Oliverio, M., Schiaparelli, S. & 4238 Costantini, M. L. 2019. Antarctic Food Web Architecture under Varying Dynamics of Sea 4239 Ice Cover. Scientific Reports, 9. 4240 Rubalcaba, J. G., Verberk, W. C. E. P., Hendriks, A. J., Saris, B. & Woods, H. A. 2020. 4241 Oxygen Limitation May Affect the Temperature and Size Dependence of Metabolism in Aquatic Ectotherms. Proceedings of the National Academy of Sciences, 117, 31963-4242 4243 31968. 4244 Ruckelshaus, M., Klinger, T., Knowlton, N. & Demaster, D. P. 2008. Marine Ecosystem-4245 Based Management in Practice: Scientific and Governance Challenges. BioScience, 58, 4246 53-63. 4247 Rudolf, V. H. W. 2012. Seasonal Shifts in Predator Body Size Diversity and Trophic 4248 Interactions in Size-Structured Predator-Prey Systems. Journal of Animal Ecology, 81, 4249 524-532. 4250 Rutterford, L. A., Simpson, S. D., Bogstad, B., Devine, J. A. & Genner, M. J. 2023. Sea 4251 Temperature Is the Primary Driver of Recent and Predicted Fish Community Structure across Northeast Atlantic Shelf Seas. Global Change Biology, 29, 2510-2521. 4252 Ruzicka, J. J., Steele, J. H., Ballerini, T., Gaichas, S. K. & Ainley, D. G. 2013. Dividing 4253 4254 up the Pie: Whales, Fish, and Humans as Competitors. Progress in Oceanography, 116, 4255 207-219. 4256 Saba, G., Bockus, A., Shaw, C. & Seibel, B. 2021. Combined Effects of Ocean 4257 Acidification and Elevated Temperature on Feeding, Growth, and Physiological Processes 4258 of Antarctic Krill Euphausia Superba. Marine Ecology Progress Series, 665, 1-18. Salvatteci, R., Schneider, R. R., Galbraith, E., Field, D., Blanz, T., Bauersachs, T., Crosta, 4259 X., Martinez, P., Echevin, V., Scholz, F. & Bertrand, A. 2022. Smaller Fish Species in a 4260 4261 Warm and Oxygen-Poor Humboldt Current System. Science, 375, 101-104. Sambilay, V. C. 1990. Interrelationships between Swimming Speed, Caudal Fin Aspect 4262 Ratio and Body Length of Fishes. Fishbyte, 8, 16-20. 4263

- 4264 Sánchez-Hernández, J., Nunn, A. D., Adams, C. E. & Amundsen, P.-A. 2019. Causes and
  4265 Consequences of Ontogenetic Dietary Shifts: A Global Synthesis Using Fish Models.
  4266 *Biological Reviews*, 94, 539-554.
- Sanchez-Martinez, P., Ackerly, D. D., Martínez-Vilalta, J., Mencuccini, M., Dexter, K. G.
  & Dawson, T. E. 2024. A Framework to Study and Predict Functional Trait Syndromes
  Using Phylogenetic and Environmental Data. *Methods in Ecology and Evolution*, 15, 666681.
- Saunders, R. A., Collins, M. A., Shreeve, R., Ward, P., Stowasser, G., Hill, S. L. &
  Tarling, G. A. 2018. Seasonal Variation in the Predatory Impact of Myctophids on
  Zooplankton in the Scotia Sea (Southern Ocean). *Progress in Oceanography*, 168, 123144.
- 4275 Saunders, R. A., Hill, S. L., Tarling, G. A. & Murphy, E. J. 2019. Myctophid Fish (Family
  4276 Myctophidae) Are Central Consumers in the Food Web of the Scotia Sea (Southern
  4277 Ocean). Frontiers in Marine Science, 6.
- Saunders, R. A. & Tarling, G. A. 2018. Southern Ocean Mesopelagic Fish Comply with
  Bergmann's Rule. *The American Naturalist*, 191, 343-351.
- Savoca, M. S., Czapanskiy, M. F., Kahane-Rapport, S. R., Gough, W. T., Fahlbusch, J. A.,
  Bierlich, K. C., Segre, P. S., Di Clemente, J., Penry, G. S., Wiley, D. N., Calambokidis, J.,
  Nowacek, D. P., Johnston, D. W., Pyenson, N. D., Friedlaender, A. S., Hazen, E. L. &
  Goldbogen, J. A. 2021. Baleen Whale Prey Consumption Based on High-Resolution
  Foraging Measurements. *Nature*, 599, 85-90.
- Scharf, F. S., Juanes, F. & Rountree, R. A. 2000. Predator Size-Prey Size Relationships of
  Marine Fish Predators: Interspecific Variation and Effects of Ontogeny and Body Size on
  Trophic-Niche Breadth. *Marine Ecology Progress Series*, 208, 229-248.
- Schmid, B., Balvanera, P., Cardinale, B. J., Godbold, J., Pfisterer, A. B., Raffaelli, D.,
  Solan, M. & Srivastava, D. S. 2009. Consequences of Species Loss for Ecosystem
  Functioning: Meta-Analyses of Data from Biodiversity Experiments. *In:* Shahid Naeem,
  D. E. B., Andy Hector, Michel Loreau, Charles Perrings (ed.) *Biodiversity, Ecosystem Functioning, and Human Wellbeing: An Ecological and Economic Perspective.* Oxford
  Scholarship Online: Oxford University Press.
- Schmidt, K., Atkinson, A., Steigenberger, S., Fielding, S., Lindsay, M. C. M., Pond, D.
  W., Tarling, G. A., Klevjer, T. A., Allen, C. S., Nicol, S. & Achterberg, E. P. 2011. Seabed
  Foraging by Antarctic Krill: Implications for Stock Assessment, Bentho-Pelagic
  Coupling, and the Vertical Transfer of Iron. *Limnology and Oceanography*, 56, 14111428.
- Schoener, T. W. 1970. Nonsynchronous Spatial Overlap of Lizards in Patchy Habitats. *Ecology*, 51, 408-418.
- 4301 Schoener, T. W. 1982. The Controversy over Interspecific Competition: Despite Spirited
  4302 Criticism, Competition Continues to Occupy a Major Domain in Ecological Thought.
  4303 American Naturalist, 70, 586-595.

4304 4305 4306	Schram, J., Schoenrock, K., Mcclintock, J., Amsler, C. & Angus, R. 2016. Seawater Acidification More Than Warming Presents a Challenge for Two Antarctic Macroalgal-Associated Amphipods. <i>Marine Ecology Progress Series</i> , 554, 81-97.
4307 4308	Schuckel, S., Sell, A. F., Kroncke, I. & Reiss, H. 2012. Diet Overlap among Flatfish Species in the Southern North Sea. <i>J Fish Biol</i> , 80, 2571-94.
4309 4310	Sentis, A., Binzer, A. & Boukal, D. S. 2017. Temperature-Size Responses Alter Food Chain Persistence across Environmental Gradients. <i>Ecology Letters</i> , 20, 852-862.
4311 4312 4313	Shackell, N. L., Frank, K. T., Fisher, J. a. D., Petrie, B. & Leggett, W. C. 2010. Decline in Top Predator Body Size and Changing Climate Alter Trophic Structure in an Oceanic Ecosystem. <i>Proceedings of the Royal Society B: Biological Sciences</i> , 277, 1353-1360.
4314 4315 4316 4317	Sheehy, J. M., Taylor, N. L., Zwerschke, N., Collar, M., Morgan, V. & Merayo, E. 2022. Review of Evaluation and Valuation Methods for Cetacean Regulation and Maintenance Ecosystem Services with the Joint Cetacean Protocol Data. <i>Frontiers in Marine Science</i> , 9.
4318 4319 4320 4321	Siqueira, T., Hawkins, C. P., Olden, J. D., Tonkin, J., Comte, L., Saito, V. S., Anderson, T. L., Barbosa, G. P., Bonada, N. & Bonecker, C. C. 2024. Understanding Temporal Variability across Trophic Levels and Spatial Scales in Freshwater Ecosystems. <i>Ecology</i> , 105, e4219.
4322	Smetacek, V. & Duarte, C. M. 2008. Impacts of Global Warming on Polar Ecosystems.
4323 4324 4325 4326	Sohlström, E. H., Archer, L. C., Gallo, B., Jochum, M., Kordas, R. L., Rall, B. C., Rosenbaum, B. & O'gorman, E. J. 2021. Thermal Acclimation Increases the Stability of a Predator–Prey Interaction in Warmer Environments. <i>Global Change Biology</i> , 27, 3765- 3778.
4327 4328 4329	Spitz, J., Ridoux, V. & Brind'amour, A. 2014. Let's Go Beyond Taxonomy in Diet Description: Testing a Trait-Based Approach to Prey–Predator Relationships. <i>Journal of Animal Ecology</i> , 83, 1137-1148.
4330 4331 4332	Srinivas, T., Sukumaran, S., Neetu, S. & Ramesh Babu, K. 2020. Diversity and Functional Patterns of Benthic Amphipods in the Coralline Intertidal Zones of a Marine National Park, India. <i>Frontiers in Marine Science</i> , 7.
4333 4334 4335	Stachowicz, J. J., Bruno, J. F. & Duffy, J. E. 2007. Understanding the Effects of Marine Biodiversity on Communities and Ecosystems. <i>Annual Review of Ecology, Evolution, and Systematics</i> , 38, 739-766.
4336 4337	Stearns, S. C. 1983. The Influence of Size and Phylogeny on Patterns of Covariation among Life-History Traits in the Mammals. <i>Oikos</i> , 41, 173-187.
4338 4339	Steenbeek, J., Corrales, X., Platts, M. & Coll, M. 2018. Ecosampler: A New Approach to Assessing Parameter Uncertainty in Ecopath with Ecosim. <i>SoftwareX</i> , 7, 198-204.
4340 4341	Steinacher, M., Joos, F., Frölicher, T. L., Bopp, L., Cadule, P., Cocco, V., Doney, S. C., Gehlen, M., Lindsay, K., Moore, J. K., Schneider, B. & Segschneider, J. 2010. Projected

- 4342 21st Century Decrease in Marine Productivity: A Multi-Model Analysis. *Biogeosciences*,
  4343 7, 979-1005.
- 4344 Stock, A., Murray, C. C., Gregr, E. J., Steenbeek, J., Woodburn, E., Micheli, F.,
  4345 Christensen, V. & Chan, K. M. A. 2023. Exploring Multiple Stressor Effects with
  4346 Ecopath, Ecosim, and Ecospace: Research Designs, Modeling Techniques, and Future
  4347 Directions. *Science of The Total Environment*, 869, 161719.
- 4348Stouffer, D. B. & Bascompte, J. 2011. Compartmentalization Increases Food-Web4349Persistence. *Proc Natl Acad Sci U S A*, 108, 3648-52.
- 4350 Strobl, C., Boulesteix, A.-L., Kneib, T., Augustin, T. & Zeileis, A. 2008. Conditional
  4351 Variable Importance for Random Forests. *BMC Bioinformatics*, 9, 307.
- 4352 Strobl, C., Boulesteix, A.-L., Zeileis, A. & Hothorn, T. 2007. Bias in Random Forest
  4353 Variable Importance Measures: Illustrations, Sources and a Solution. *BMC*4354 *Bioinformatics*, 8, 25.
- Subramaniam, R. C., Corney, S. P., Swadling, K. M. & Melbourne-Thomas, J. 2020.
  Exploring Ecosystem Structure and Function of the Northern Kerguelen Plateau Using a
  Mass-Balanced Food Web Model. *Deep Sea Research Part II: Topical Studies in Oceanography*, 174.
- Subramaniam, R. C., Pinkerton, Matt H.,, Melbourne-Thomas, J., Corney, S. P.,
  Swadling, K. M. & Pruvost, P. 2019. A Mass-Balanced Ecosystem Model for the
  Kerguelen Plateau. *HAL open science*.
- Sullivan, T., Byrne, C., Harman, L., Davenport, J., Mcallen, R. & Regan, F. 2014.
  Determination of Spatial and Temporal Variability of Ph and Dissolved Oxygen
  Concentrations in a Seasonally Hypoxic Semi-Enclosed Marine Basin Using Continuous
  Monitoring. *Anal. Methods*, 6, 5489-5497.
- Surma, S., Pakhomov, E. A. & Pitcher, T. J. 2014. Effects of Whaling on the Structure of
  the Southern Ocean Food Web: Insights on the "Krill Surplus" from Ecosystem
  Modelling. *PLoS One*, 9, e114978.
- Swadling, K. M., Constable, A. J., Fraser, A. D., Massom, R. A., Borup, M. D., Ghigliotti,
  L., Granata, A., Guglielmo, L., Johnston, N. M., Kawaguchi, S., Kennedy, F., Kiko, R.,
  Koubbi, P., Makabe, R., Martin, A., Mcminn, A., Moteki, M., Pakhomov, E. A., Peeken,
  I., Reimer, J., Reid, P., Ryan, K. G., Vacchi, M., Virtue, P., Weldrick, C. K., Wongpan, P.
  & Wotherspoon, S. J. 2023. Biological Responses to Change in Antarctic Sea Ice
  Habitats. *Frontiers in Ecology and Evolution*, 10.
- Tang, K. W., Gladyshev, M. I., Dubovskaya, O. P., Kirillin, G. & Grossart, H.-P. 2014.
  Zooplankton Carcasses and Non-Predatory Mortality in Freshwater and Inland Sea
  Environments. *Journal of Plankton Research*, 36, 597-612.
- Targett, T. E. 1981. Trophic Ecology and Structure of Coastal Antarctic Fish
  Communities. *Marine Ecology Progress Series*, 4, 243-263.
- Tarling, G. A., Stowasser, G., Ward, P., Poulton, A. J., Zhou, M., Venables, H. J., Mcgill,
  R. a. R. & Murphy, E. J. 2012a. Seasonal Trophic Structure of the Scotia Sea Pelagic

4382 4383	Ecosystem Considered through Biomass Spectra and Stable Isotope Analysis. <i>Deep Sea Research Part II: Topical Studies in Oceanography</i> , 59-60, 222-236.
4384 4385 4386	Tarling, G. A., Ward, P., Atkinson, A., Collins, M. A. & Murphy, E. J. 2012b. Discovery 2010: Spatial and Temporal Variability in a Dynamic Polar Ecosystem. <i>Deep Sea Research Part II: Topical Studies in Oceanography</i> , 59-60, 1-13.
4387 4388	Team, R. C. 2023. R: Language and Environment for Statistical Computing. <i>R Foundation for Statistical Computing, Vienna, Austria</i> . https://www.R-project.org/.
4389 4390 4391	Teixeira, C. M. G. L., Sousa, T., Marques, G. M., Domingos, T. & Kooijman, S. a. L. M. 2014. A New Perspective on the Growth Pattern of the Wandering Albatross (Diomedea Exulans) through Deb Theory. <i>Journal of Sea Research</i> , 94, 117-127.
4392 4393	Teng, J. & Mccann, K. S. 2004. Dynamics of Compartmented and Reticulate Food Webs in Relation to Energetic Flows. <i>The American Naturalist</i> , 164, 85-100.
4394 4395 4396	Teschke, K., Brtnik, P., Hain, S., Herata, H., Liebschner, A., Pehlke, H. & Brey, T. 2021. Planning Marine Protected Areas under the Ccamlr Regime–the Case of the Weddell Sea (Antarctica). <i>Marine Policy</i> , 124, 104370.
4397 4398 4399	Tews, J., Brose, U., Grimm, V., Tielbörger, K., Wichmann, M. C., Schwager, M. & Jeltsch, F. 2004. Animal Species Diversity Driven by Habitat Heterogeneity/Diversity: The Importance of Keystone Structures. <i>Journal of Biogeography</i> , 31, 79-92.
4400 4401	Thébault, E. & Fontaine, C. 2010. Stability of Ecological Communities and the Architecture of Mutualistic and Trophic Networks. <i>Science</i> , 329, 853-856.
4402 4403 4404	Thomalla, S. J., Nicholson, SA., Ryan-Keogh, T. J. & Smith, M. E. 2023. Widespread Changes in Southern Ocean Phytoplankton Blooms Linked to Climate Drivers. <i>Nature Climate Change</i> , 13, 975-984.
4405 4406 4407 4408	Thomas, K. N., Gower, D. J., Bell, R. C., Fujita, M. K., Schott, R. K. & Streicher, J. W. 2020. Eye Size and Investment in Frogs and Toads Correlate with Adult Habitat, Activity Pattern and Breeding Ecology. <i>Proceedings of the Royal Society B: Biological Sciences</i> , 287, 20201393.
4409 4410 4411	Thomas, P. G. & Green, K. 1988. Distribution of Euphausia Crystallorophias within Prydz Bay and Its Importance to the Inshore Marine Ecosystem. <i>Polar Biology</i> , 8, 327-331.
4412 4413 4414 4415	Thompson, R. M., Brose, U., Dunne, J. A., Hall, R. O., Hladyz, S., Kitching, R. L., Martinez, N. D., Rantala, H., Romanuk, T. N., Stouffer, D. B. & Tylianakis, J. M. 2012. Food Webs: Reconciling the Structure and Function of Biodiversity. <i>Trends in Ecology &amp; Evolution</i> , 27, 689-697.
4416 4417	Thompson, R. M. & Townsend, C. R. 2005. Energy Availability, Spatial Heterogeneity and Ecosystem Size Predict Food-Web Structure in Streams. <i>Oikos</i> , 108, 137-148.
4418 4419	Tilman, D. & Downing, J. A. 1994. Biodiversity and Stability in Grasslands. <i>Nature</i> , 367, 363-365.

4422 Tilman, D., Reich, P. B. & Knops, J. M. H. 2006. Biodiversity and Ecosystem Stability in a Decade-Long Grassland Experiment. Nature, 441, 629-632. 4423 4424 Townsend, C. R., Begon, M. & Harper, J. L. 2008. Essentials of Ecology, Blackwell 4425 publishing. 4426 Trathan, P. N. & Hill, S. L. 2016. The Importance of Krill Predation in the Southern 4427 Ocean. In: Siegel, V. (ed.) Biology and Ecology of Antarctic Krill. Cham: Springer 4428 International Publishing. 4429 Tsai, C. H., Hsieh, C. H. & Nakazawa, T. 2016. Predator-Prey Mass Ratio Revisited: 4430 Does Preference of Relative Prey Body Size Depend on Individual Predator Size? 4431 Functional Ecology, 30, 1979-1987. 4432 Tucker, M. A. & Rogers, T. L. 2014. Examining Predator-Prey Body Size, Trophic Level and Body Mass across Marine and Terrestrial Mammals. Proceedings of the Royal Society 4433 4434 *B: Biological Sciences*, 281, 20142103. Tulloch, V. J. D., Plaganyi, E. E., Brown, C., Richardson, A. J. & Matear, R. 2019. Future 4435 4436 Recovery of Baleen Whales Is Imperiled by Climate Change. Glob Chang Biol, 25, 1263-4437 1281. 4438 Ullah, H., Nagelkerken, I., Goldenberg, S. U. & Fordham, D. A. 2018. Climate Change 4439 Could Drive Marine Food Web Collapse through Altered Trophic Flows and 4440 Cyanobacterial Proliferation. PLOS Biology, 16, e2003446. 4441 Van De Putte, A. P., Griffiths, H. J., Brooks, C., Bricher, P., Sweetlove, M., Halfter, S. & 4442 Raymond, B. 2021. From Data to Marine Ecosystem Assessments of the Southern Ocean: 4443 Achievements, Challenges, and Lessons for the Future. Frontiers in Marine Science, 8. 4444 Van Der Meer, J., Hin, V., Van Oort, P. & Van De Wolfshaar, K. E. 2022. A Simple Deb-4445 Based Ecosystem Model. Conservation Physiology, 10. 4446 Van Der Meer, J., Van Donk, S., Sotillo, A. & Lens, L. 2020. Predicting Post-Natal 4447 Energy Intake of Lesser Black-Backed Gull Chicks by Dynamic Energy Budget 4448 Modeling. Ecological Modelling, 423. 4449 Violle, C., Navas, M.-L., Vile, D., Kazakou, E., Fortunel, C., Hummel, I. & Garnier, E. 4450 2007. Let the Concept of Trait Be Functional! Oikos, 116, 882-892. 4451 Voronina, N. M. 1998. Comparative Abundance and Distribution of Major Filter-Feeders 4452 in the Antarctic Pelagic Zone. Journal of Marine Systems, 17, 375-390. 4453 Vucic-Pestic, O., Rall, B. C., Kalinkat, G. & Brose, U. 2010. Allometric Functional 4454 Response Model: Body Masses Constrain Interaction Strengths. Journal of Animal Ecology, 79, 249-256. 4455

Tilman, D., Isbell, F. & Cowles, J. M. 2014. Biodiversity and Ecosystem Functioning.

Annual Review of Ecology, Evolution, and Systematics, 45, 471-493.

4420

4456 Wainwright, P., Carroll, A. M., Collar, D. C., Day, S. W., Higham, T. E. & Holzman, R. A. 4457 2007. Suction Feeding Mechanics, Performance, and Diversity in Fishes. Integr Comp 4458 *Biol*, 47, 96-106. 4459 Wallis, J. R., Kawaguchi, S., Matsuno, K. & Swadling, M. 2019. Big Things Come in 4460 Small Packages. Biomass Contribution of the Krill Thysanoessa Macrura to the Marine 4461 Ecosystem in the Kerguelen Plateau Region. Second Kerguelen Plateau Symposium: 4462 marine ecosystem and fisheries, 55-58. Waluda, C. M., Hill, S. L., Peat, H. J. & Trathan, P. N. 2017. Long-Term Variability in the 4463 Diet and Reproductive Performance of Penguins at Bird Island, South Georgia. Marine 4464 4465 Biology, 164. Wang, R., Zhang, R., Miao, X., Li, H., Song, P., Li, Y. & Lin, L. 2024. Demersal Fish 4466 4467 Community in the near-Shelf Zone of the Cosmonaut Sea, Southern Ocean. Diversity, 16, 4468 156. 4469 Wang, T., Zhang, P., Molinos, J. G., Xie, J., Zhang, H., Wang, H., Xu, X., Wang, K., Feng, 4470 M. & Cheng, H. 2023. Interactions between Climate Warming, Herbicides, and 4471 Eutrophication in the Aquatic Food Web. Journal of Environmental Management, 345, 118753. 4472 4473 Ward-Campbell, B. M. S., Beamish, F. W. H. & Kongchaiya, C. 2005. Morphological 4474 Characteristics in Relation to Diet in Five Coexisting Thai Fish Species. Journal of Fish 4475 Biology, 67, 1266-1279. 4476 Ward, P., Atkinson, A. & Tarling, G. 2012. Mesozooplankton Community Structure and 4477 Variability in the Scotia Sea: A Seasonal Comparison. Deep Sea Research Part II: Topical Studies in Oceanography, 59-60, 78-92. 4478 4479 Ward, P., Tarling, G., Shreeve, R. & Ten Hoopen, P. 2020. Epipelagic Mesozooplankton 4480 Distribution and Abundance in Southern Ocean Atlantic Sector and the North Atlantic and 4481 Arctic 1996-2013 (Version 1.0). . UK Polar Data Centre, Natural Environment Research 4482 Council, UK Research & Innovation. . 4483 Watters, G. M., Hill, S. L., Hinke, J. T., Matthews, J. & Reid, K. 2013. Decision-Making 4484 for Ecosystem-Based Management: Evaluating Options for a Krill Fishery with an 4485 Ecosystem Dynamics Model. Ecological Applications, 23, 710-725. Wege, M., Salas, L. & Larue, M. 2021. Ice Matters: Life-History Strategies of Two 4486 4487 Antarctic Seals Dictate Climate Change Eventualities in the Weddell Sea. Global Change 4488 Biology, 27, 6252-6262. 4489 Weigel, B. & Bonsdorff, E. 2018. Trait-Based Predation Suitability Offers Insight into 4490 Effects of Changing Prey Communities. PeerJ, 6, e5899. 4491 Weiss, A. I., King, J. C., Lachlan-Cope, T. A. & Ladkin, R. S. 2012. Albedo of the Ice 4492 Covered Weddell and Bellingshausen Seas. The Cryosphere, 6, 479-491. 4493 White, E. P., Ernest, S. K. M., Kerkhoff, A. J. & Enquist, B. J. 2007. Relationships 4494 between Body Size and Abundance in Ecology. Trends in Ecology & amp; Evolution, 22, 4495 323-330.

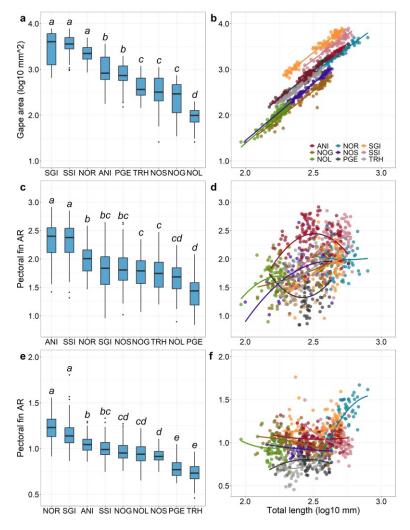
- Whitehouse, G. A. & Aydin, K. Y. 2020. Assessing the Sensitivity of Three Alaska Marine
  Food Webs to Perturbations: An Example of Ecosim Simulations Using Rpath. *Ecological Modelling*, 429, 109074.
- Whitehouse, M. J., Meredith, M. P., Rothery, P., Atkinson, A., Ward, P. & Korb, R. E.
  2008. Rapid Warming of the Ocean around South Georgia, Southern Ocean, During the
  20th Century: Forcings, Characteristics and Implications for Lower Trophic Levels. *Deep*Sea Research Part I: Oceanographic Research Papers, 55, 1218-1228.
- Winkler, N. S., Paz-Goicoechea, M., Lamb, R. W. & Pérez-Matus, A. 2017. Diet Reveals
  Links between Morphology and Foraging in a Cryptic Temperate Reef Fish. *Ecology and Evolution*, 7, 11124-11134.
- Wood, S. A., Russell, R., Hanson, D., Williams, R. J. & Dunne, J. A. 2015. Effects of
  Spatial Scale of Sampling on Food Web Structure. *Ecology and Evolution*, 5, 3769-3782.
- Woodward, G., Blanchard, J., Lauridsen, R. B., Edwards, F. K., Jones, J. I., Figueroa, D.,
  Warren, P. H. & Petchey, O. L. 2010. Chapter 6 Individual-Based Food Webs: Species
  Identity, Body Size and Sampling Effects. *In:* Woodward, G. (ed.) *Advances in Ecological Research*. Academic Press.
- Woodward, G., Ebenman, B., Emmerson, M., Montoya, J. M., Olesen, J. M., Valido, A. &
  Warren, P. H. 2005. Body Size in Ecological Networks. *Trends in Ecology & Evolution*,
  20, 402-409.
- Wootton, K. L., Curtsdotter, A., Roslin, T., Bommarco, R. & Jonsson, T. 2021. Towards a
  Modular Theory of Trophic Interactions. *Functional Ecology*, 37, 26-43.
- Wootton, K. L. & Stouffer, D. B. 2016. Many Weak Interactions and Few Strong; FoodWeb Feasibility Depends on the Combination of the Strength of Species' Interactions and
  Their Correct Arrangement. *Theoretical Ecology*, 9, 185-195.
- 4520 Yachi, S. & Loreau, M. 1999. Biodiversity and Ecosystem Productivity in a Fluctuating
  4521 Environment: The Insurance Hypothesis. *Proceedings of the National Academy of*4522 Sciences, 96, 1463-1468.
- Yang, G., Atkinson, A., Hill, S. L., Guglielmo, L., Granata, A. & Li, C. 2020. Changing
  Circumpolar Distributions and Isoscapes of Antarctic Krill: Indo-Pacific Habitat Refuges
  Counter Long-Term Degradation of the Atlantic Sector. *Limnology and Oceanography*,
  66, 272-287.
- Yang, G., Atkinson, A., Pakhomov, E. A., Hill, S. L. & Racault, M. F. 2022. Massive
  Circumpolar Biomass of Southern Ocean Zooplankton: Implications for Food Web
  Structure, Carbon Export, and Marine Spatial Planning. *Limnology and Oceanography*,
  67, 2516-2530.
- Zerbini, A. N., Adams, G., Best, J., Clapham, P. J., Jackson, J. A. & Punt, A. E. 2019.
  Assessing the Recovery of an Antarctic Predator from Historical Exploitation. *Royal Society Open Science*, 6, 190368.

- Zhao, L., Zhang, H., O'gorman, E. J., Tian, W., Ma, A., Moore, J. C., Borrett, S. R. &
  Woodward, G. 2016. Weighting and Indirect Effects Identify Keystone Species in Food
- 4536 Webs. *Ecology Letters*, 19, 1032-1040.
- Zhao, L., Zhang, H., Tian, W. & Xu, X. 2017. Identifying Compartments in Ecological
  Networks Based on Energy Channels. *Ecology and Evolution*, 8, 309-318.

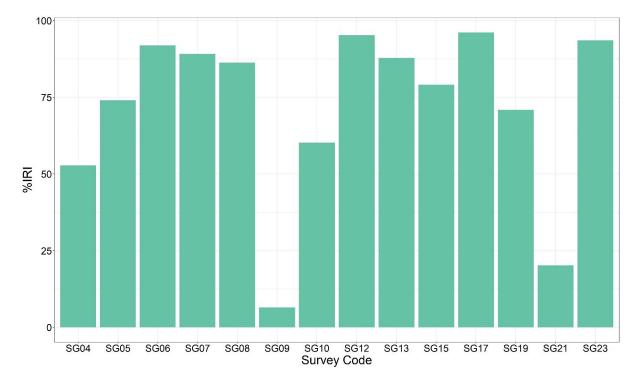
# 4540 Appendix A: Supplementary material for chapter 2

### 4541 *A1: Supplementary figures*

4542



4543 Figure A1: Panels a, c, and e are boxplots of the distribution of mouth gape area, caudal fin 4544 aspect ratio (AR) and pectoral fin AR, respectively, in 9 South Georgia demersal fish species. 4545 Boxplots are ordered by decreasing median value, with letters indicating groupings assigned 4546 by a Dunn's test with Bonferroni correction (groups with a letter in common are not 4547 significantly different). Panels b, d, and f display the relationship between total length and 4548 mouth gape area, caudal fin aspect ratio (AR) and pectoral fin AR, respectively. Regression 4549 lines represent first-order (panel b) and second-order (panels d and f) polynomial fits from 4550 linear regression models for each species. Species codes are: ANI, Champsocephalus 4551 gunnari; SSI, Chaenocephalus aceratus; SGI, Pseudochaenichthys georgianus; NOR, 4552 Notothenia rossii; TRH, Trematomus hansoni; NOS, Lepidonotothen squamifrons; NOL, 4553 Lepidonotothen larseni; NOG, Gobionotothen gibberifrons; PGE, Parachaenichthys 4554 georgianus.





4556 Figure A2: Barplot comparing the total percentage Index of Relative Importance (%IRI) of

- 4557 krill (all Euphausiidae) in the stomachs of Mackerel icefish (Champsocephalus gunnari)
- 4558 during each survey since 2004, including the values calculated from the 2023 data (SG23).

### *A2: Supplementary tables*

Package	Reference	Usage
stats	R Core Team (2023)	Cluster analysis and PCA
factoextra	Kassambara & Mundt (2020)	PCA visualisation
vegan	Oksanen et al. (2022)	SIMPER analysis
nlme	Pinheiro et al. (2023)	Linear mixed effects modelling
FSA	Ogle, et al. (2023)	Dunn's tests
party	Hothorn et al. (2005)	Random Forest modelling
ggplot2	Wickham (2016)	Plotting
RColorBrewer	· Neuwirth (2022)	Plotting
tidyverse	Wickham et al. (2019)	Data handling

4560 Table A1: Packages and functions used during analyses:

4566 Table A2: Outputs of SIMPER analysis with 99 permutations indicating the contribution of

4567 each prey group to the overall dissimilarities in diet composition (based on Index of Relative

4568 Importance, %IRI) between feeding guilds. 'Average' represents the contribution to average

4569 between-group dissimilarity, and 'SD' indicates the standard deviation of this contribution.

4570 The p-value indicates the probability of getting a larger or equal average contribution across

4571 the random permutations.

Krill feeders - Benthos feeders								
Average SD p-value								
Krill	0.1749	0.07372	0.84					
Isopod	0.1008	0.06872	0.01					
Misc. benthos	0.09615	0.01213	0.01					
Fish	0.04096	0.06526	1					
Themisto	0.03667	0.03613	0.99					
Other								
amphipods	0.03464	0.02752	0.37					
Mysid	0.00275	0.00329	1					
Benthic shrimps	0	0	1					
	eeders - Fish	feeders						
	Average	SD	p-value					
Krill	0.3945	0.07424	0.01					
Fish	0.296	0.10198	0.01					
Other								
amphipods	0.0556	0.0822	0.04					
Themisto	0.0495	0.02517	1					
Mysid	0.0268	0.04226	0.95					
Isopod	0.0034	0.00416	1					
Benthic shrimps	0.0006	0.00075	0.99					
Misc. benthos	0.0001	0.00024	1					
Krill feeders	- Themisto	& krill feed	ders					
	Average	SD	p-value					
Krill	0.1966	0.07348	0.55					
Themisto	0.1492	0.06747	0.01					
Fish	0.0677	0.0739	1					
Other								
amphipods	0.01775	0.03051	0.93					
Misc. benthos	0.01188	0.0201	0.88					
Isopod	0.00817	0.01385	0.84					
Mysid	0.00311	0.00324	1					
Benthic shrimps	0.00044	0.00076	0.99					
Krill feeder	s - Benthic s	hrimp feed						
Average SD p-value								
Krill	0.3911	0.07511	0.01					
Benthic shrimps	0.2566	0.10619	0.01					
Mysid	0.2018	0.07327	0.01					
Fish	0.043	0.06098	1					

Other amphipods $0.0002$ $0.00023$ 1           Isopod         0 $0.00003$ 1           Misc. benthos         0         0         1           Misc. benthos         0         0         1           Misc. benthos         0         0         1           Misc. benthos         0.07915         0.01           Krill         0.2196         0.04635         0.29           Isopod         0.0974         0.0691         0.01           Misc. benthos         0.096         0.01217         0.01           Misc. benthos         0.096         0.02343         0.99           Mysid         0.0267         0.04271         0.79           Benthic shrimps         0.0006         0.00076         0.85           Misc. benthos feeders - <i>Themisto &amp;</i> krill feeders         Average         SD         p-value           Themisto         0.16459         0.04883         0.01           Isopod         0.05016         0.07652         0.99           Krill         0.05016         0.07652         0.99           Krill         0.02071         0.46         Mysid           0.00205         0.00166         0.99<	Themisto	0.0317	0.04903	0.99					
amphipods         0.0002         0.00023         1           Isopod         0         0         1           Misc. benthos         6eders - Fish feeders         -           Average         SD         p-value           Fish         0.3365         0.07915         0.01           Krill         0.2196         0.04635         0.29           Isopod         0.0974         0.0691         0.01           Misc. benthos         0.096         0.01217         0.01           Other         -         -         -           amphipods         0.0625         0.06394         0.19           Themisto         0.03         0.02343         0.99           Mysid         0.0267         0.04271         0.79           Benthic shrimps         0.0006         0.00076         0.85           Misc. benthos         0.16459         0.04883         0.01           Isopod         0.09264         0.0706         0.01           Misc. benthos         0.08427         0.02371         0.01           Fish         0.03479         0.02771         0.46           Mysid         0.02025         0.0016         0.99           Ben									
Isopod         0         0.00003         1           Misc. benthos         0         0         1           Misc. benthos         feeders - Fish feeders         SD         p-value           Fish         0.3365         0.07915         0.01           Krill         0.2196         0.04635         0.29           Isopod         0.0974         0.0691         0.01           Misc. benthos         0.096         0.01217         0.01           Other		0.0002	0.00023	1					
Misc. benthos         0         0         1           Misc. benthos feeders - Fish feeders         SD         p-value           Fish         0.3365         0.07915         0.01           Krill         0.2196         0.04635         0.29           Isopod         0.0974         0.0691         0.01           Misc. benthos         0.096         0.01217         0.01           Other				1					
Misc. benthos feeders - Fish feeders           Average         SD         p-value           Fish $0.3365$ $0.07915$ $0.01$ Krill $0.2196$ $0.04635$ $0.29$ Isopod $0.0974$ $0.0691$ $0.01$ Misc. benthos $0.096$ $0.01217$ $0.01$ Other	*	-							
AverageSDp-valueFish $0.3365$ $0.07915$ $0.01$ Krill $0.2196$ $0.04635$ $0.29$ Isopod $0.0974$ $0.0691$ $0.01$ Misc. benthos $0.096$ $0.01217$ $0.01$ Other									
Fish $0.3365$ $0.07915$ $0.01$ Krill $0.2196$ $0.04635$ $0.29$ Isopod $0.0974$ $0.0691$ $0.01$ Misc. benthos $0.096$ $0.01217$ $0.01$ Other $0.0625$ $0.06394$ $0.19$ Themisto $0.03$ $0.02343$ $0.99$ Mysid $0.0267$ $0.04271$ $0.79$ Benthic shrimps $0.0006$ $0.00076$ $0.85$ Misc. benthos feeders - Themisto & krill feedersAverageSDp-valueThemisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Misc. benthos $0.08427$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Krill $0.004143$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.0044$ $0.00077$ $0.86$ Misc. benthos $0.09615$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10811$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Mysid $0.28544$ $0.00382$ $0.98$ Fish $0.02854$ $0.1093$ $0.01$ Krill $0.13458$ $0.05342$ $0.01$ M			1						
Krill $0.2196$ $0.04635$ $0.29$ Isopod $0.0974$ $0.0691$ $0.01$ Misc. benthos $0.096$ $0.01217$ $0.01$ Other $amphipods$ $0.0625$ $0.06394$ $0.19$ Themisto $0.03$ $0.02343$ $0.99$ Mysid $0.0267$ $0.04271$ $0.79$ Benthic shrimps $0.0006$ $0.00076$ $0.85$ Misc. benthos feeders - Themisto & krill feedersAverageSDp-valueThemisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.0044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.22077$ $0.749$ $0.01$ Isopod $0.1081$ $0.07034$ $0.01$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.1081$ $0.07034$ $0.01$ Mysid $0.02644$ $0.00832$ $0.98$ Fish $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.02854$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0$	Fish	<u> </u>							
Isopod $0.0974$ $0.0691$ $0.01$ Misc. benthos $0.096$ $0.01217$ $0.01$ Otheramphipods $0.0625$ $0.06394$ $0.19$ Themisto $0.03$ $0.02343$ $0.99$ Mysid $0.0267$ $0.04271$ $0.79$ Benthic shrimps $0.0006$ $0.00076$ $0.85$ Misc. benthos feeders - Themisto & krill feedersAverageSDp-valueThemisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Misc. benthos $0.08427$ $0.02743$ 1Other $0.00205$ $0.00166$ $0.99$ Krill $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00444$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDMysid $0.22027$ $0.0749$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Mysid $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$									
Misc. benthos $0.096$ $0.01217$ $0.01$ Other amphipods $0.0625$ $0.06394$ $0.19$ Themisto $0.03$ $0.02343$ $0.99$ Mysid $0.0267$ $0.04271$ $0.79$ Benthic shrimps $0.0006$ $0.00076$ $0.85$ Misc. benthos feeders - Themisto & krill feedersAverageSD $p$ -valueThemisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ 1Other $m$ $m$ amphipods $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.0044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSD $p$ -valueBenthic shrimps $0.22663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Mysid $0.02854$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.0$									
Other amphipods $0.0625$ $0.06394$ $0.19$ Themisto $0.03$ $0.02343$ $0.99$ Mysid $0.0267$ $0.04271$ $0.79$ Benthic shrimps $0.0006$ $0.00076$ $0.85$ Misc. benthos feeders - Themisto & krill feeders         Average         SD         p-value           Themisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Misc. benthos $0.08427$ $0.02743$ $1$ Other         amphipods $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.0044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feeders $Average$ SD         p-value           Benthic shrimps $0.22077$ $0.0749$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.22027$ $0.0749$ <t< td=""><td>· · · · ·</td><td></td><td></td><td></td></t<>	· · · · ·								
amphipods $0.0625$ $0.06394$ $0.19$ Themisto $0.03$ $0.02343$ $0.99$ Mysid $0.0267$ $0.04271$ $0.79$ Benthic shrimps $0.0006$ $0.00076$ $0.85$ Misc. benthos feeders - Themisto & krill feedersAverageSDp-valueThemisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ $1$ Otheramphipods $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ OtherImphipods $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ 1Fish feeders - Themisto & krill feedersAverageSDp-valueFish feeders - Themisto & krill feedersFish feeders - Themisto & 0.07484 $0.0015$ Ot									
Themisto $0.03$ $0.02343$ $0.99$ Mysid $0.0267$ $0.04271$ $0.79$ Benthic shrimps $0.0006$ $0.00076$ $0.85$ Misc. benthos feeders - Themisto & krill feedersAverageSDp-valueThemisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ $1$ Otheramphipods $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Otheramphipods $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ 1Fish feeders - Themisto & krill feedersFish feeders - Themisto & krill feedersFish feeders - Themisto & 0.0595 $0.0$		0.0625	0.06394	0.19					
Mysid $0.0267$ $0.04271$ $0.79$ Benthic shrimps $0.0006$ $0.00076$ $0.85$ Misc. benthos feeders - Themisto & krill feedersAverageSDp-valueThemisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ $1$ Otheramphipods $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Misc. benthos $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.02778$ $0.015$ 1Fish feeders - Themisto & krill feedersFish feeders - Themisto & krill feedersFish feeders - Themisto & 0.0595 $0.07484$ $0.06$ Mysid $0.02678$ $0.0421$ $0.89$ Misc. bent									
Benthic shrimps $0.0006$ $0.0076$ $0.85$ Misc. benthos feeders - Themisto & krill feeders         Average         SD         p-value           Themisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ 1           Other         amphipods $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.0044$ $0.00771$ $0.46$ Mysid $0.02055$ $0.0166$ $0.99$ Benthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.22027$ $0.0749$ $0.01$ Krill $0.22027$ $0.0749$ $0.01$ Isopod $0.10081$ $0.0734$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Mis									
Misc. benthos feeders - Themisto & krill feedersAverageSDp-valueThemisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ $1$ Otheramphipods $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.0044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Otheramphipods $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ $1$ Fish feeders - Themisto & krill feedersAverageSDp-valueFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other </td <td></td> <td></td> <td></td> <td></td>									
AverageSDp-valueThemisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ $1$ Other $amphipods$ $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.0044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSD $p$ -valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Other $amphipods$ $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ $1$ Fish feeders - Themisto & krill feeders $Average$ SDFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $amphipods$ $0.02678$ $0.0421$ Mysid $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$									
Themisto $0.16459$ $0.04883$ $0.01$ Isopod $0.09264$ $0.0706$ $0.01$ Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ $1$ Other $amphipods$ $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $amphipods$ $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ $1$ Fish feeders - Themisto & krill feedersAverageSDp-valueFish $0.28654$ $0.1093$ $0.01$ Krill $0.13458$ $0.05342$ $0.01$ Other $amphipods$ $0.0595$ $0.07484$ $0.06$ Mysid $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$									
Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ $1$ Other $amphipods$ $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSD $p$ -valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $amphipods$ $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ $1$ Fish feeders - Themisto & krill feedersAverageSD $p$ -valueFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $amphipods$ $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$	Themisto	0	0.04883						
Misc. benthos $0.08427$ $0.02371$ $0.01$ Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ $1$ Other $amphipods$ $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSD $p$ -valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $amphipods$ $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ $1$ Fish feeders - Themisto & krill feedersAverageSD $p$ -valueFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $amphipods$ $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$	Isopod	0.09264	0.0706	0.01					
Fish $0.05016$ $0.07652$ $0.99$ Krill $0.04143$ $0.02743$ 1Other $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $amphipods$ $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ 1Fish feeders - Themisto & krill feedersAverageSDp-valueFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $amphipods$ $0.0595$ $0.07484$ $0.06$ Mysid $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$									
Krill $0.04143$ $0.02743$ 1Other	Fish			0.99					
Other amphipods $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ 1Fish feeders - Themisto & krill feeders $Average$ SDP-valueFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $amphipods$ $0.0595$ $0.07484$ $0.06$ Mysid $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$									
amphipods $0.03479$ $0.02771$ $0.46$ Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ 1Fish feeders - Themisto & krill feeders $x$ AverageSD $p$ -valueFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $a$ $a$ $a$ Misc. benthos $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$	Other								
Mysid $0.00205$ $0.00166$ $0.99$ Benthic shrimps $0.00044$ $0.00077$ $0.86$ Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Otheramphipods $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ 1Fish feeders - Themisto & krill feedersFish $0.28654$ $0.1093$ $0.01$ Krill $0.13458$ $0.05342$ $0.01$ Otheramphipods $0.0595$ $0.07484$ $0.06$ Mysid $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$	amphipods	0.03479	0.02771	0.46					
Misc. benthos feeders - Benthic shrimp feedersAverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ $1$ Fish feeders - Themisto & krill feeders $Average$ SDFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $amphipods$ $0.0595$ $0.07484$ $0.06$ Mysid $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$		0.00205	0.00166	0.99					
AverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ $1$ Fish feeders - Themisto & krill feeders $Average$ SDFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $amphipods$ $0.0595$ $0.07484$ $0.06$ Mysid $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$		0.00044	0.00077						
AverageSDp-valueBenthic shrimps $0.25663$ $0.10869$ $0.01$ Krill $0.21624$ $0.04803$ $0.28$ Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ $1$ Fish feeders - Themisto & krill feeders $Average$ SDFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $amphipods$ $0.0595$ $0.07484$ $0.06$ Mysid $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$	Misc. benthos fe	eders - Bent	hic shrimp	feeders					
Krill0.216240.048030.28Mysid0.202070.07490.01Isopod0.100810.070340.01Misc. benthos0.096150.012410.01Otheramphipods0.034440.028160.37Themisto0.015840.003820.98Fish0.004780.00151Fish feeders - Themisto & krill feedersAverageSDp-valueFish0.286540.10930.01Krill0.19790.046220.61Themisto0.134580.053420.01Otheramphipods0.026780.04210.89Misc. benthos0.011840.020090.83									
Mysid $0.20207$ $0.0749$ $0.01$ Isopod $0.10081$ $0.07034$ $0.01$ Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ $1$ Fish feeders - Themisto & krill feedersAverageSDFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $0.0595$ $0.07484$ $0.06$ Mysid $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$	Benthic shrimps	0.25663	0.10869	0.01					
Isopod         0.10081         0.07034         0.01           Misc. benthos         0.09615         0.01241         0.01           Other         0.03444         0.02816         0.37           Themisto         0.01584         0.00382         0.98           Fish         0.00478         0.0015         1           Fish feeders - Themisto & krill feeders         Average         SD         p-value           Fish         0.28654         0.1093         0.01           Krill         0.1979         0.04622         0.61           Themisto         0.13458         0.05342         0.01           Other	Krill	0.21624	0.04803	0.28					
Isopod         0.10081         0.07034         0.01           Misc. benthos         0.09615         0.01241         0.01           Other         0.03444         0.02816         0.37           Themisto         0.01584         0.00382         0.98           Fish         0.00478         0.0015         1           Fish feeders - Themisto & krill feeders         Average         SD         p-value           Fish         0.28654         0.1093         0.01           Krill         0.1979         0.04622         0.61           Themisto         0.13458         0.05342         0.01           Other	Mysid	0.20207	0.0749	0.01					
Misc. benthos $0.09615$ $0.01241$ $0.01$ Other $0.03444$ $0.02816$ $0.37$ amphipods $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ $1$ Fish feeders - Themisto & krill feedersAverageSDp-valueFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$				0.01					
Other amphipods $0.03444$ $0.02816$ $0.37$ Themisto $0.01584$ $0.00382$ $0.98$ Fish $0.00478$ $0.0015$ 1Fish feeders - Themisto & krill feedersAverageSDp-valueFish $0.28654$ $0.1093$ $0.01$ Krill $0.1979$ $0.04622$ $0.61$ Themisto $0.13458$ $0.05342$ $0.01$ Other amphipods $0.02678$ $0.0421$ $0.89$ Misc. benthos $0.01184$ $0.02009$ $0.83$				0.01					
Themisto         0.01584         0.00382         0.98           Fish         0.00478         0.0015         1           Fish feeders - Themisto & krill feeders         Average         SD         p-value           Fish         0.28654         0.1093         0.01           Krill         0.1979         0.04622         0.61           Themisto         0.13458         0.05342         0.01           Other           amphipods         0.02678         0.0421         0.89           Misc. benthos         0.01184         0.02009         0.83         0.03         0.01									
Themisto         0.01584         0.00382         0.98           Fish         0.00478         0.0015         1           Fish feeders - Themisto & krill feeders         Average         SD         p-value           Fish         0.28654         0.1093         0.01           Krill         0.1979         0.04622         0.61           Themisto         0.13458         0.05342         0.01           Other           amphipods         0.02678         0.0421         0.89           Misc. benthos         0.01184         0.02009         0.83         0.03         0.01	amphipods	0.03444	0.02816	0.37					
Fish feeders - Themisto & krill feeders           Average         SD         p-value           Fish         0.28654         0.1093         0.01           Krill         0.1979         0.04622         0.61           Themisto         0.13458         0.05342         0.01           Other									
AverageSDp-valueFish0.286540.10930.01Krill0.19790.046220.61Themisto0.134580.053420.01Otheramphipods0.05950.074840.06Mysid0.026780.04210.89Misc. benthos0.011840.020090.83	Fish	0.00478	0.0015	1					
Fish0.286540.10930.01Krill0.19790.046220.61Themisto0.134580.053420.01Otheramphipods0.05950.074840.06Mysid0.026780.04210.89Misc. benthos0.011840.020090.83	Fish feeders	- Themisto	& krill feed	lers					
Fish0.286540.10930.01Krill0.19790.046220.61Themisto0.134580.053420.01Otheramphipods0.05950.074840.06Mysid0.026780.04210.89Misc. benthos0.011840.020090.83		Average	SD	p-value					
Themisto0.134580.053420.01Other	Fish	Ŭ	0.1093	•					
Other amphipods0.05950.074840.06Mysid0.026780.04210.89Misc. benthos0.011840.020090.83	Krill	0.1979	0.04622	0.61					
amphipods0.05950.074840.06Mysid0.026780.04210.89Misc. benthos0.011840.020090.83	Themisto		0.05342	0.01					
Mysid0.026780.04210.89Misc. benthos0.011840.020090.83	Other								
Misc. benthos 0.01184 0.02009 0.83	amphipods	0.0595	0.07484	0.06					
	Mysid	0.02678	0.0421	0.89					
Isopod 0.00971 0.01173 0.78	Misc. benthos	0.01184	0.02009	0.83					
	Isopod	0.00971	0.01173	0.78					

Benthic shrimps	0.00074	0.00079	0.93				
Fish feeders - Benthic shrimp feeders							
	Average	SD	p-value				
Fish	0.3318	0.07917	0.01				
Benthic shrimps	0.256	0.10658	0.01				
Mysid	0.1771	0.08545	0.01				
Other							
amphipods	0.0556	0.08279	0.21				
Themisto	0.0459	0.02313	0.91				
Krill	0.0398	0.02724	1				
Isopod	0.0034	0.0042	0.94				
Misc. benthos	0.0001	0.00024	0.99				
Themisto & krill	feeders - Ben	thic shrim	p feeders				
	Average SD						
Benthic shrimps	0.2562	0.10692	0.01				
Mysid	0.20156	0.07369	0.01				
Krill	0.19455	0.04741	0.48				
Themisto	0.18043	0.04869	0.01				
Fish	0.04998	0.07353	0.99				
Other							
amphipods	0.01773	0.03076	0.78				
Misc. benthos	0.01188	0.02033	0.79				
Isopod	0.00817	0.01401	0.77				

.....

- 4584 Table A3: Model selection for optimum random effects and variance weighting structure for
- 4585 the linear mixed effects model describing the relationship between prey body mass  $(\log_{10} g)$
- 4586 and the predictors predator body mass  $(\log_{10} g)$  and predator feeding guild plus their
- 4587 interaction. The final model structure selected based on Bayesian Information Criterion (BIC)
- 4588 and level of parsimony is highlighted in bold.

6	Random effects structure		Variance structure				BIC	
None	~1   Prey taxon	$\sim$ Predator mass   Prey taxon	varldent(~1  Feeding group)	varIdent(~1  Prey taxon)	varFixed(~Predator mass)	varExp(~Predator mass)	varConst (~Predator mass)	
X	x							1419.92 1071.11
	А	х						1080.51
			х					1080.09
				Х				1092.52
			х	Х				1110.26
					х			1055.98
						х		1060.82
			x		х		Х	1066.71 1076.93
			л Х		л	х		1070.93
			X				х	1086.63
				х	х			1080.09
				Х		х		1085.93
				Х			Х	1092.23

- 4597 Table A4: Model selection for optimum fixed effects structure for the linear mixed effects
- 4598 model describing the relationship between prey body mass  $(\log_{10} g)$  and the predictors
- 4599 predator body mass (log<sub>10</sub> g) and predator feeding guild. Each model includes the optimal
- 4600 random effects and variance weighting structure identified in Table A3. The most
- 4601 parsimonious model structure based on Bayesian Information Criterion (BIC) and retaining
- 4602 only significant fixed effects is highlighted in bold.

Fixed effects structure	BIC
Predator mass * Feeding guild	1054.76
Predator mass + Feeding guild	1029.92
Predator mass	1037.52
Feeding guild	1157.13
Null	1144.12

4604 Table A5: Linear mixed effects model estimates of the relationship between prey mass and

4605 the additive combination of predator mass and feeding guild. The reference level is the krill-

4606 feeding guild.

Coefficient	Estimate	SE	df	t-value	p-value
Intercept	-1.245	0.215	599	-5.777	<0.001
Predator mass (log <sub>10</sub> g)	0.562	0.045	599	12.471	<0.001
Benthos	-0.260	0.066	599	-3.921	<0.001
Fish	-0.414	0.080	599	-5.144	<0.001
Themisto and krill	-0.267	0.061	599	-4.389	<0.001
Benthic shrimps	-0.123	0.141	599	-0.868	0.386

4607

4608 Table A6: Results of Kruskal-Wallis tests of differences in the distribution of trait values4609 between feeding guilds.

Trait	chi-squared	df	p-value
Gape area	224.47	4	< 0.001
Caudal AR	171.54	4	< 0.001
Pectoral AR	262.96	4	< 0.001

4610

4611

4612

4614 Table A7: Results of pairwise Dunns tests with Bonferroni correction for differences in the

	Gape_area		Cauda	ıl AR	Pector	al AR
Feeding guild pairing	Z	P.adj	Z	P.adj	Z	P.adj
Krill - Benthos	-8.486	< 0.001	-4.994	< 0.001	-4.631	< 0.001
Krill - Fish	7.174	< 0.001	2.184	0.289	-0.966	1
Krill – Themisto & krill	4.282	< 0.001	3.897	< 0.001	10.744	< 0.001
Krill - Benthic shrimps	-0.311	1	-10.753	< 0.001	-12.968	< 0.001
Benthos - Fish	-13.138	< 0.001	-6.174	< 0.001	-3.370	0.008
Benthos - Themisto & krill	-4.334	< 0.001	-1.493	1	4.340	< 0.001
Benthos - Benthic shrimps	6.314	< 0.001	-5.042	< 0.001	-7.299	< 0.001
Fish - Themisto & krill	9.947	< 0.001	5.290	< 0.001	8.486	< 0.001
Fish - Benthic shrimps	-5.472	< 0.001	-11.412	< 0.001	-11.021	< 0.001
Themisto & krill - Benthic shrimps	2.814	0.049	-6.926	< 0.001	-3.743	0.002

4615 distribution of traits between feeding guilds.

- 4617 Table A8: Axis loadings for a Principle Components Analysis (PCA) of the trait values for
- 4618 each individual fish.

Dimension	Eigenvalue	% variance	Cumulative % variance
PC1	1.660	55.327	55.327
PC2	0.778	25.928	81.256
PC3	0.562	18.745	100

- 4620 Table A9: Variable loadings for a Principle Components Analysis (PCA) of the trait values for
- 4621 each individual fish.

Trait	PC1	PC2	PC3
Gape area	0.624	-0.218	-0.750
Caudal AR	0.510	0.842	0.179
Pectoral	0.592	-0.494	0.636
AR			

### 4631 Supplementary references:

- Hothorn, T., Bühlmann, P., Dudoit, S., Molinaro, A. & Van Der Laan, M. J. 2005. Survival
  Ensembles. Biostatistics, 7, 355-373.
- 4634 Kassambara, A., Mundt, F. (2020). factoextra: Extract and Visualize the Results of
- 4635 Multivariate Data Analyses. R package version 1.0.7, https://CRAN.R-
- 4636 project.org/package=factoextra.
- 4637 Neuwirth E (2022). RColorBrewer: ColorBrewer Palettes. R package version 1.1-3,
- 4638 https://CRAN.R-project.org/package=RColorBrewer.
- 4639 Ogle, D. H., Doll, J. C., Wheeler, A. P. & Dinno, A. 2023. Fsa: Simple Fisheries Stock
  4640 Assessment Methods. R Package Version 0.9.4. https://CRAN.R-project.org/package=FSA.
- 4641 Oksanen, J., Simpson G., Blanchet F., Kindt R., Legendre P., Minchin P., O'hara R., Solymos
- 4642 P., Stevens M., Szoecs E., Wagner H., Barbour M., Bedward M., Bolker B., Borcard D.,
- 4643 Carvalho G., Chirico M., De Caceres M., Durand S., Evangelista H., Fitzjohn R., Friendly
- 4644 M., Furneaux B., Hannigan G., Hill M., Lahti L., Mcglinn D, Ouellette M., Ribeiro Cunha E.,
- 4645 Smith T., Stier A., Ter Braak C. & J., W. 2022. Vegan: Community Ecology Package. R
- 4646 Package Version 2.6-4. https://CRAN.R-project.org/package=vegan
- 4647 Pinheiro J., Bates D. & Team, R. C. 2023. Nlme: Linear and Nonlinear Mixed Effects
- 4648 Models. R Package Version 3.1-162. https://CRAN.R-project.org/package=nlme.
- 4649 R Core Team 2023. R: Language and Environment for Statistical Computing. R Foundation
  4650 for Statistical Computing, Vienna, Austria. https://www.R-project.org/.
- Wickham, H. (2016). ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag New
  York. ISBN 978-3-319-24277-4, https://ggplot2.tidyverse.org.
- 4653 Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L.D., François, R., Grolemund,
- 4654 G., Hayes, A., Henry, L., Hester, J., Kuhn, M., Pedersen, T.L., Miller, E., Bache, S.M.,
- 4655 Müller, K., Ooms, J., Robinson, D., Seidel, D.P., Spinu, V., Takahashi, K., Vaughan, D.,
- 4656 Wilke, C., Woo, K., Yutani, H. (2019). "Welcome to the tidyverse." Journal of Open Source
- 4657 Software, 4(43), 1686. doi:10.21105/joss.01686, https://doi.org/10.21105/joss.01686.

## 4659 Appendix B: Supplementary material for chapter 3

- 4660 *B1: Supplementary methods and results*
- 4661
- 4662 Supplementary methods
- 4663 *Overview of functional traits.*

Here we detail the functional traits compiled for this study, including a description of theirecological relevance and an overview of how they were coded for analyses.

Body mass data were already available for each food web, having been compiled through a combination of direct measurement in the field and the compilation of values from the literature (Brose et al. 2019 ; López-López et al. 2021). Upon inspection of the body mass data, a small number of discrepancies within and between the food webs were identified, and for consistency these were corrected using published mass data sources (see Supplementary Data). Values were then subject to log10 transformation for analyses.

4672 Foraging habitat represents the physical space in which the organisms are primarily found, and 4673 therefore plays a role in determining which trophic interactions are feasible due to species 4674 overlap. Broad habitat categories were identified for each food web, and species were assigned 4675 to these based on literature review, with those found across multiple habitats assigned to a 4676 combined habitat category. For the Scotia Sea food web, three pelagic habitat categories 4677 ("epipelagic", "mesopelagic" and "benthopelagic") plus possible combinations of these ("epi-4678 mesopelagic", "epi-meso-bathypelagic", "meso-bathypelagic") were taken from the original 4679 publication (López-López et al. 2021). In the Weddell Sea, habitats were assigned as "benthic", 4680 "lower pelagic" and "upper pelagic", in addition to the combinations of "upper and lower pelagic", "upper and lower pelagic and benthic", and "lower pelagic and benthic". For Lough 4681 4682 Hyne and Kongsfjorden, habitats were assigned as "benthic", "intertidal" and "pelagic", in 4683 addition to "benthic and intertidal", "benthic and pelagic", "pelagic and intertidal", and "benthic, intertidal and pelagic", to reflect possible habitat overlaps. 4684

4685 Mobility represents the primary propulsive method used by each species, which will influence 4686 how likely predators and prey are to come into contact, and how easily consumers can capture 4687 resources or resources can evade their consumers. We assigned a scale of increasing mobility: 4688 0 (sessile, attached); 1 (passive drifter, no substantial self-locomotion); 2 (crawler); 3
4689 (swimming by cilia/flagella or appendages); 4 (jet propulsion); 5 (lift-based swimming).

4690 Prey-capture strategy represents how active the predator is in capturing prey. The method 4691 employed will influence how likely different consumers and resources are to come into contact, 4692 and which types of resource can be consumed. We assigned a scale of increasing activity: 0 4693 (Producer, no resource capture involved); 1 (Passive, no action by the consumer until after 4694 contact with resource has been made); 2 (Ambush, consumer is relatively inactive but selects 4695 and actively captures resources when in range); 3 (Active suspension/detritus feeder, displays 4696 limited activity such as pumping of water or sifting through sediment which may involve some 4697 locomotion, but prey selection occurs after contact); 4 (Active search, consumer moves actively 4698 searching for resources either as a browsing herbivore or hunting predator). In some cases, the 4699 primary capture strategy was not documented in the literature, in which case it was inferred 4700 from diet where possible (e.g., consumers of sessile prey must employ an active searching 4701 strategy).

4702 Capture appendages represent any external appendage (i.e. tentacles/arms, legs) which could 4703 reasonably be considered to play a role in grasping and manipulating prey. Organisms which 4704 lack such appendages must engulf their prey to capture them. Cilia and flagella were not 4705 considered capture appendages due to their primary role in locomotion and producing simple 4706 feeding currents.

Body robustness represents the general body type of the organism, which will influence which
consumers it has. We assigned a scale of increasing robustness: 0 (gelatinous); 1 (soft-bodied,
no internal skeleton/shell); 2 (soft-bodied with internal skeleton/shell); 3 (external skeleton); 4
(external hard shell).

4711 Spines are expected to deter certain consumers as they can cause physical damage and may 4712 make consumption difficult. The presence or absence of spines was determined based on image 4713 assessment. Spines were only considered present if their role could be confidently assigned to 4714 defence (e.g., rostral/dorsal spines in copepods and amphipods).

The translucency trait represents how visible organisms are within the water column and is expected to play a role in both prey capture and predator avoidance. This trait was assigned based on assessment of images. 4718 Traits were assigned to all nodes except for difficult to define basal groups (detritus and4719 sediment).

#### 4720 *Robustness of module assignments*

4721 The stochastic element to the Simulated Annealing algorithm used to identify food web 4722 modules means the final results of different runs can vary both in terms of the number of 4723 modules identified and the partitioning of nodes between modules (i.e. the identity of species 4724 assigned to the same module could differ between runs). It was therefore important to ensure 4725 that the final modularity partition reported for each network was representative of the spread 4726 of results obtained across different runs of the algorithm. To select an appropriate result for 4727 each food web we first identified the number of modules in each Simulated Annealing output 4728 and then randomly selected a result displaying the most representative number of modules 4729 across the runs.

88% of runs for the Scotia Sea web identified three modules while the remaining 12% identified
two modules, and 95% of runs for the Weddell Sea identified three modules while 5% identified
four modules. A result with three modules was therefore randomly selected for these two webs.
For Lough Hyne, 81% of runs identified five modules, and the remaining 19% found four,
while for Kongsfjorden 83% of runs identified five, 16% identified four, and 1% identified six
modules. A result with five modules was randomly selected for both networks.

4736 The next step was to identify how consistent the partitioning of species into different modules 4737 in the selected result was across the remaining Simulated Annealing runs. We followed the 4738 methods of Rezende et al. (2009) in focusing on the distribution of interactions within modules. 4739 For each interacting species pair assigned to the same module in the chosen Simulated 4740 Annealing run, we calculated the number of times that pair co-occurred in a module across all 4741 the remaining runs. The results suggest that there is extremely low total variability in the 4742 module membership of interacting pairs, as the overwhelming majority of within-module 4743 interactions were consistent across runs: 96.6% in the Scotia Sea; 95.3% in the Weddell Sea; 4744 99.2% in Lough Hyne; 99.5% in Kongsfjorden. These results support the robustness of the 4745 partitioning of the focal food webs by the Simulated Annealing algorithm.

Finally, we investigated the variability in trophic level (TL) and log<sub>10</sub> body mass of each module for each network, across Simulated Annealing runs. For each run, we investigated the proportion of pairwise differences in the distribution of TL and body masses between modules 4749 that were significant. The results of our randomly selected outputs were generally consistent 4750 with those of the remaining Simulated Annealing runs. In the Scotia Sea, the average proportion 4751 of significant pairwise differences was 94.0% ( $\pm$  1.3% standard error) for TL, and 88.0% ( $\pm$ 4752 2.6%) for body mass. In the Weddell Sea, these proportions were 99.0% ( $\pm$  0.4%) for TL and 98.0% ( $\pm$  0.7%) for mass. In Lough Hyne, the average proportion of significant pairwise 4753 4754 differences was 72.0% ( $\pm 0.5\%$ ) for TL, and 31.0% ( $\pm 0.2\%$ ) for mass. For comparison, in the 4755 randomly chosen run, 70.0% of pairwise comparisons were significantly different for TL, while 4756 30.0% were significantly different for mass. In Kongsfjorden, the values were 34.5% ( $\pm 1.0\%$ ) 4757 for TL and 45.1% ( $\pm$  1.1%) for mass. In the randomly chosen run for this network, 30.0% of 4758 pairwise comparisons were significantly different for TL, while 40.0% were significantly different for mass. When plotting the average TL and body mass in each module for all the 4759 4760 runs, the distribution of values in the randomly chosen run was generally consistent with the 4761 distribution of values from the other runs, though in a minority of cases, differences in the total number of modules or changes to the distribution of species across modules resulted in outliers 4762 4763 (Figure B1 & B2.). These changes would be unlikely to alter the key findings of this study, as 4764 the overall pattern of modules in each network is maintained (modules in the Scotia Sea and 4765 Weddell Sea encompass distinct trophic clusters and body mass distributions while many of 4766 those in Lough Hyne and Kongsfjorden are not significantly different).

### 4767 Supplementary results

#### 4768 *Differences in trophic level variance within and between modules*

4769 A potential issue with comparing the mean difference in trophic level between modules in each 4770 network is that this metric could be sensitive to both the maximum trophic level of a given 4771 network and the number of modules identified. We therefore calculated two metrics; the 4772 variance in trophic level for all pairwise combinations of species from the same module 4773 (within-variance), and the variance in trophic level for all pairwise combinations of species from separate modules (between-variance). The ratio of the within-variance and between-4774 4775 variance provides a metric that is independent of network size and maximum trophic level and 4776 describes the organisation of modules: larger values indicate that modules encompass a wide 4777 range of trophic levels but have a large overlap with one-another; smaller values indicate that 4778 modules have less overlap in trophic levels. The results using this metric support the 4779 conclusions drawn using the mean trophic level differences: modules in the Scotia Sea and

4780 Weddell Sea food webs have lower values than those in Lough Hyne and Kongsfjorden (0.49,

4781 SE = 0.001 and 0.58, SE = 0.001 versus 0.90, SE = 0.001 and 1.26, SE = 0.001, respectively).

### 4782 Distribution of node metrics across modules

4783 There is a significant decline in average vulnerability from modules 1 to 3 in both the Scotia 4784 Sea and Weddell Sea food webs (Figure B3). This likely results from the organisation of 4785 modules by trophic level, as organisms occupying the highest trophic levels (i.e. those in 4786 module 3) are the least vulnerable as they have relatively few predators. Similarly, the 4787 increase in omnivory with trophic level in both networks also fits with the organisation of 4788 modules by trophic level, as species found higher in the feeding hierarchy have a greater 4789 variety of organisms of different trophic levels available to feed upon (Thompson et al. 4790 (2007; Figure B4). The contrast in the change in generality across modules in these two 4791 networks (an increase in the Scotia Sea and a decrease in the Weddell Sea; Figure B5) 4792 suggests that top predators in the Scotia Sea feed on a greater number of prey species than 4793 those lower in the food web, while the inverse is true of top predators in the Weddell Sea. It is 4794 not clear what the underlying driver(s) of this difference in the level of dietary specialisation 4795 of higher predators in the two networks might be, but it is interesting that despite these 4796 contrasts both food webs display very similar structuring of modularity by trophic levels. In 4797 the Lough Hyne and Kongsfjorden food webs there is a lack of consistent differences in these 4798 node-level metrics across modules. This fits the description of modules in these networks as 4799 semi-isolated energy channels, as most modules contain an assemblage of species centred 4800 around distinct basal resources and therefore are not structured by trophic level in the same 4801 way as those in the Scotia Sea and Weddell Sea.

4802

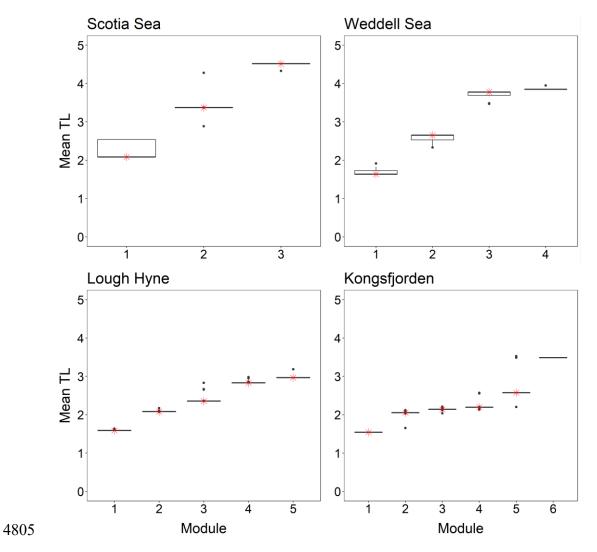


Figure B1: The distribution of mean trophic level of each module across all runs of the Simulated Annealing algorithm for each network. Red stars indicate the values for the randomly selected outputs used in later analyses. Note that the Weddell Sea had 4 modules in only 5% of runs, whilst Kongsfjorden had 6 modules in only 1% of runs

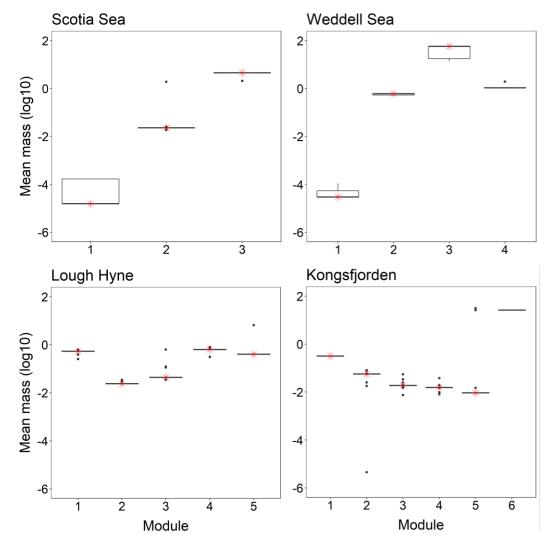
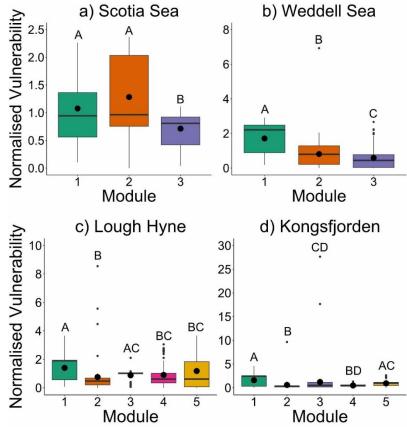


Figure B2: The distribution of mean body mass of each module across all runs of the Simulated
Annealing algorithm for each network. Red stars indicate the values for the randomly selected
outputs used in later analyses. Note that the Weddell Sea had 4 modules in only 5% of runs,
whilst Kongsfjorden had 6 modules in only 1% of runs





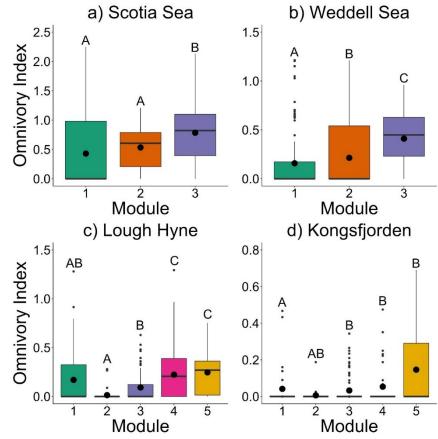
4815 4816 Figure B3: Boxplots of node normalised vulnerability across compartments within each food

web. Large black points indicate the mean, thick horizontal lines represent the median, boxes 4817

indicate the  $25^{\text{th}} - 75^{\text{th}}$  percentile range, whiskers are  $1.5 \times$  the interquartile range, and 4818

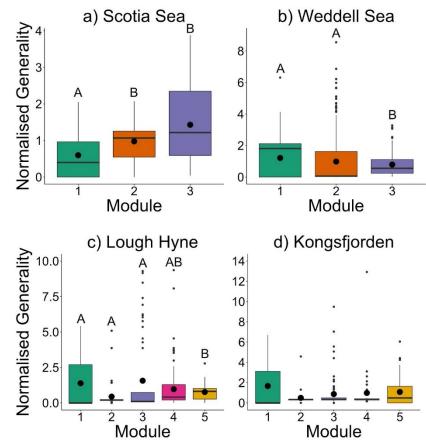
4819 outliers beyond this range are indicated as small black points. Boxes not sharing a common

4820 letter are significantly different from one another (Dunn's test, p < 0.05).





4821 4822 Figure B4: Boxplots of node omnivory index values across modules within each food web. 4823 Large black points indicate the mean, thick horizontal lines represent the median, boxes 4824 indicate the 25th -75th percentile range, whiskers are  $1.5 \times$  the interquartile range, and 4825 outliers beyond this range are indicated as small black points. Boxes not sharing a common 4826 letter are significantly different from one another (Dunn's test, p < 0.05). Note varying axis 4827 scales.





4828 4829 Figure B5: Boxplots of node normalised generality across modules within each food web.

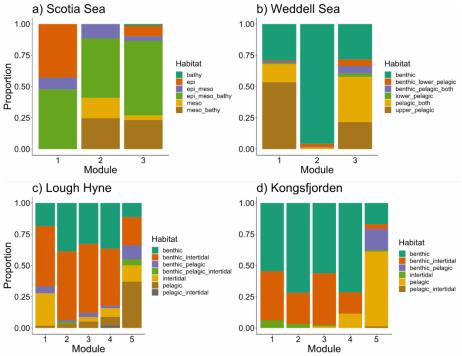
4830 Large black points indicate the mean, thick horizontal lines represent the median, boxes

4831 indicate the 25th -75th percentile range, whiskers are  $1.5 \times$  the interquartile range, and

outliers beyond this range are indicated as small black points. Boxes not sharing a common 4832

4833 letter are significantly different from one another (Dunn's test, p < 0.05). No significant

4834 differences were found for Kongsfjorden. Note varying axis scales.

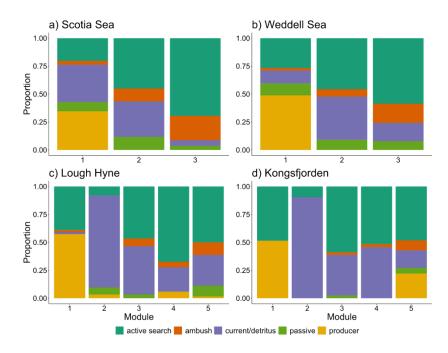


4835 Module Module Module Module Module 4836 Figure B6: Bar plots displaying the proportion of species within each food web module which

4837 were assigned to each foraging habitat. In a), "bathy" represents bathypelagic, "epi"

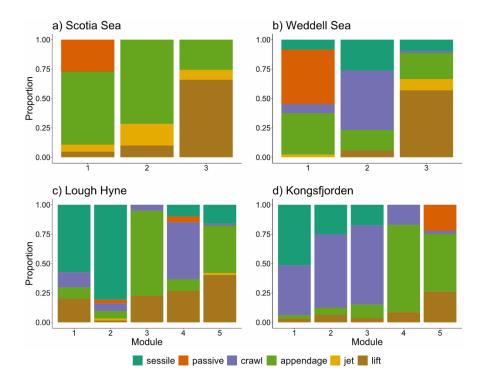
4838 represents epipelagic and "meso" represents mesopelagic. In all plots, habitats joined by "\_"

4839 indicate that the species forage across multiple habitats.



4840

4841 Figure B7: Bar plots displaying the proportion of species within each food web module which4842 were assigned to each prey-capture strategy.





4845 Figure B8: Bar plots displaying the proportion of species within each food web module which

4846 were assigned to each movement method.

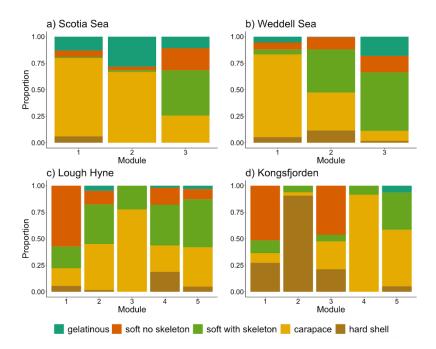
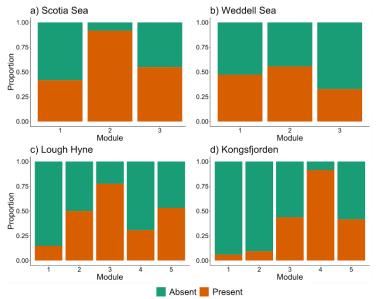




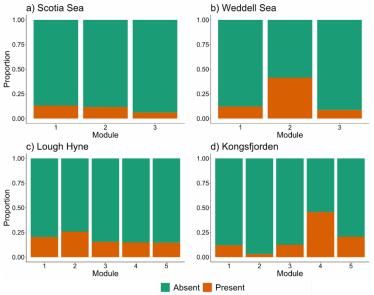
Figure B9: Bar plots displaying the proportion of species within each food web module whichwere assigned to each body robustness category.



4851
4852 Figure B10: Bar plots displaying the proportion of species within each food web module

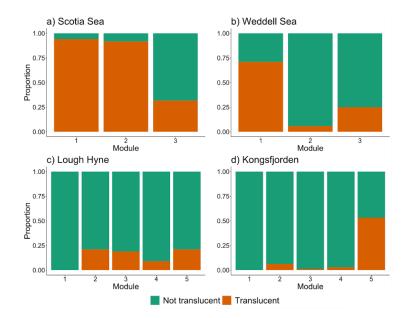
4853 which were considered to have feeding appendages capable of grasping and manipulating

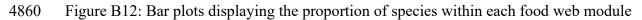
4854 prey.



4855
4856 Figure B11: Bar plots displaying the proportion of species within each food web module

4857 which were considered to have defensive spines.





4861 which were considered to be largely translucent.

### 4862 **Supplementary references**:

Brose, U., Archambault, P., Barnes, A. D., Bersier, L.-F., Boy, T., Canning-Clode, J., Conti, E.,
Dias, M., Digel, C., Dissanayake, A., Flores, A. a. V., Fussmann, K., Gauzens, B., Gray, C.,
Häussler, J., Hirt, M. R., Jacob, U., Jochum, M., Kéfi, S., Mclaughlin, O., Macpherson, M. M.,
Latz, E., Layer-Dobra, K., Legagneux, P., Li, Y., Madeira, C., Martinez, N. D., Mendonça, V.,
Mulder, C., Navarrete, S. A., O'gorman, E. J., Ott, D.,

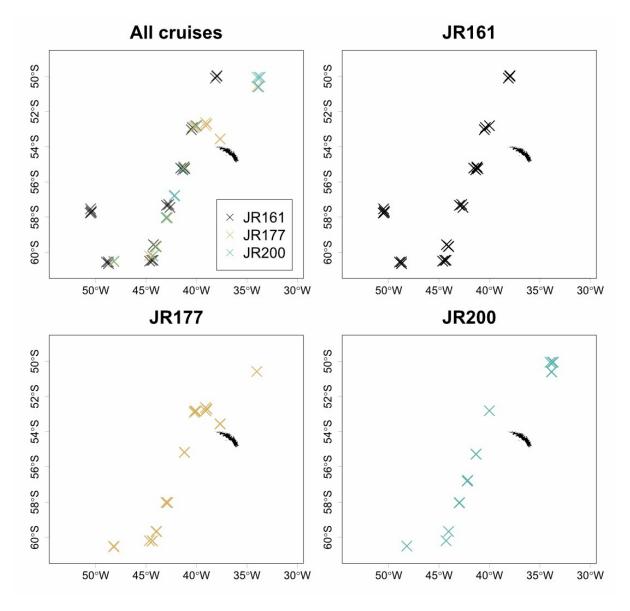
4868 López-López, L., Genner, M. J., Tarling, G. A., Saunders, R. A. & O'gorman, E. J. 2021.
4869 Ecological Networks in the Scotia Sea: Structural Changes across Latitude and Depth.
4870 *Ecosystems*.

4871 Rezende, E. L., Albert, E. M., Fortuna, M. A. & Bascompte, J. 2009. Compartments in a Marine

- Food Web Associated with Phylogeny, Body Mass, and Habitat Structure. *Ecol Lett*, 12, 779-88.
- 4874 Thompson, R. M., Hemberg, M., Starzomski, B. M. & Shurin, J. B. 2007. Trophic Levels and
- 4875 Trophic Tangles: The Prevalence of Omnivory in Real Food Webs. Ecology, 88, 612-617.
- 4876

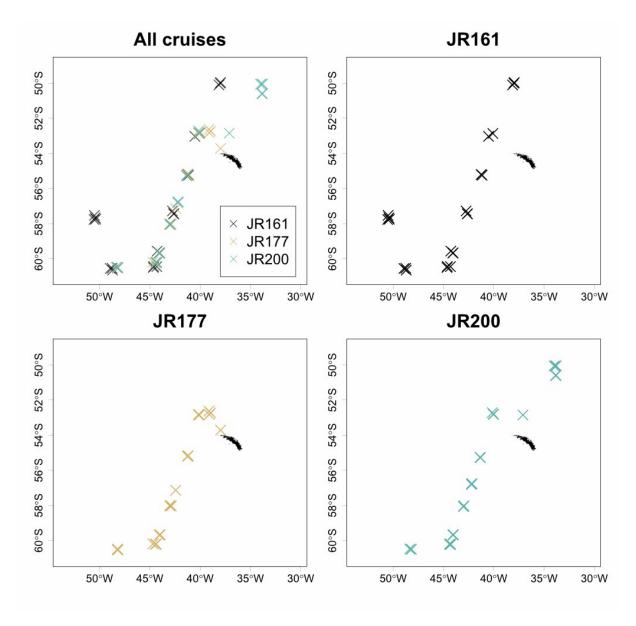
# 4877 Appendix C: Supplementary material for chapter 4

# 4878 *C1: Supplementary figures*

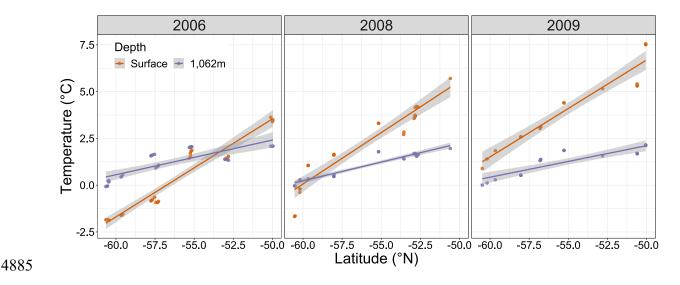




4880 Figure C1: Distribution of myctophid sampling stations from each cruise.



4883 Figure C2: Distribution of zooplankton sampling stations from each cruise.



4886 Figure C3: Comparison of temperatures at different depths across the sampling sites.

4887 Relationship between temperature (both at the surface and at 1,062m depth) and latitude at

4888 each haul location, split by sampling year (n = 27, 25, and 18 sites in 2006, 2008 and 2009,

respectively). Lines represent model predicted values and shading represents 95% confidenceintervals.

4891

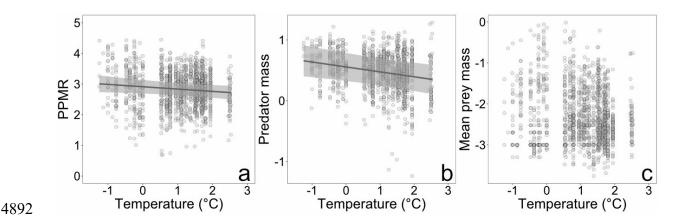
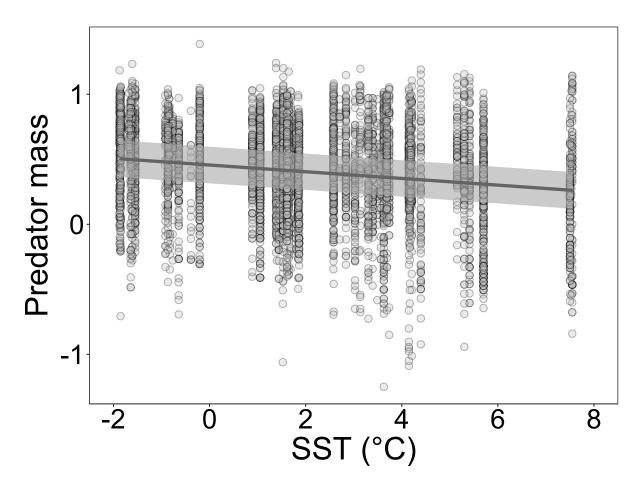
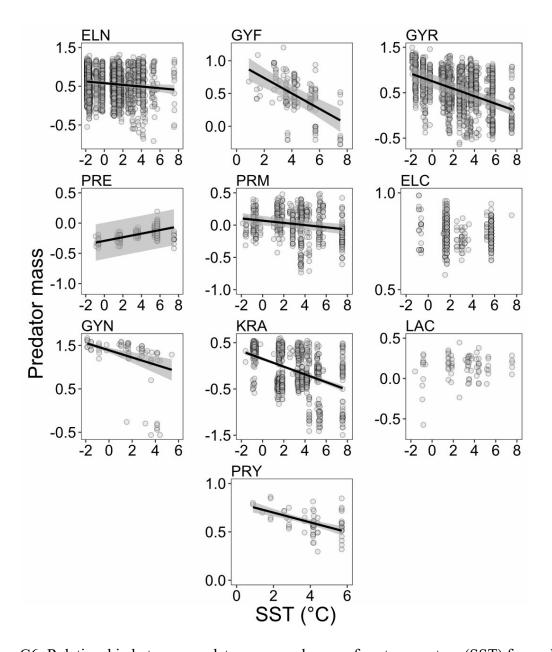


Figure C4: Effects of temperature at 1,062 m depth on predator and prey body mass (n = 1576 fish). (a) partial residual plot from a linear mixed model of the effect of temperature (at 1,062m depth) on prey-averaged predator-prey mass ratio (PPMR); (b) partial residual plot from a linear mixed model of the effect of temperature on predator body mass; (c) scatterplot of the relationship between temperature and abundance-weighted average prey mass in predator stomachs. Y-axis values are in log<sub>10</sub> g. Lines represent predicted values at each SST. Shading represents 95% confidence intervals.



4903Figure C5: Effects of temperature on predator body mass from a larger dataset. Partial4904residual plot from a linear mixed model of the effect of sea-surface temperature (SST) on4905predator body mass using a larger dataset of myctophid body sizes (n = 6,143). Y-axis values4906are in  $log_{10}$  g. Line represents predicted values of predator mass at each temperature. Shading4907represents 95% confidence intervals.



4909 Figure C6: Relationship between predator mass and sea surface temperature (SST) for each

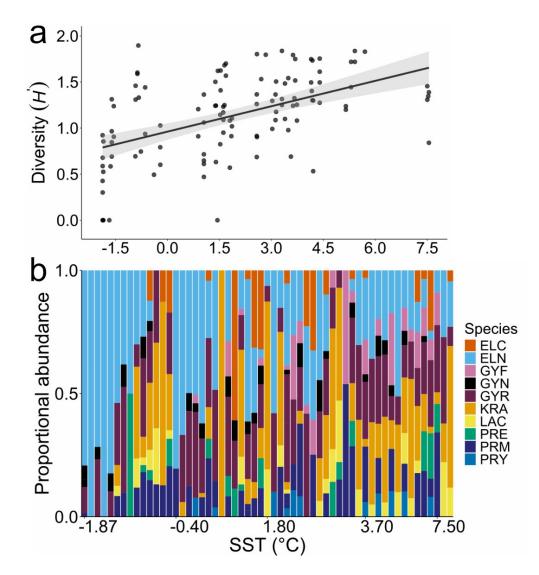
4910 species. Panels ELN, GYF, GYR, PRE and PRM are partial residuals plots, the remainder are

4911 scatterplots of the raw data with regression lines indicating predicted values from a

4912 Generalised Least Squares model. Y-axis values are in log<sub>10</sub> g. Panels with no regression lines

4913 indicate species for which no significant trend in size with SST was identified. Shading

- 4914 represents 95% confidence intervals. ELC = E. carlsbergi (n = 486 fish), ELN = E. antarctica
- 4915 (n = 2,101), GYF = G. fraseri (n = 143), GYN = G. nicholsi (n = 68), GYR = G. braueri (n = 143)
- 4916 1448), KRA = K. and erssoni (n = 944), LAC = N. achirus (n = 76), PRE = P. tension (n = 160)
- 4917 217), PRM = *P. bolini* (*n* = 596), PRY = *P. choriodon* (*n* = 64).





4919 Figure C7: Effects of temperature on myctophid community diversity and the relative 4920 abundance of each species. a) Generalised Least Squares regression model predicted values 4921 of species diversity (Shannon-Wiener index) versus sea-surface temperature (SST). Shading 4922 represents 95% confidence interval. b) stacked bar plot showing the change in proportional 4923 sqrt-transformed abundance of each species across SST. Species codes are ELC = E. 4924 *carlsbergi* (n = 26 individual abundance estimates), ELN = E. *antarctica* (n = 101), GYF = G. 4925 fraseri (n = 27), GYN = G. nicholsi (n = 36), GYR = G. braueri (n = 91), KRA = K. 4926 and ersson (n = 64), LAC = N. achirus (n = 32), PRE = P. tension (n = 16), PRM = P. bolini 4927 (n = 52), PRY = P. choriodon (n = 15). A clear shift in species composition can be seen with 4928 increasing temperature, from communities dominated by the relatively large-bodied E. 4929 antarctica at low temperatures to ones with a greater proportion of smaller species like K. 4930 anderssoni under warmer conditions. Note the discrete x-axis scale for panel b.

## *C2: Supplementary tables*

- 4933 Table C1: Length-Mass relationships used to estimate body mass of each individual
- 4934 myctophid species. The regressions used to convert standard length (SL, mm) to wet mass
- 4935 (WM, g) follow the equation  $WM = a * SL^b$ . Lower and upper 95% confidence intervals are
- 4936 provided for each coefficient, along with the overall  $R^2$  for the relationship.

Species	а	Lower	Upper	b	Lower	Upper	<b>R</b> <sup>2</sup>
E. carlsbergi	2.09 x10 <sup>-05</sup>	9.51 x10 <sup>-06</sup>	4.59 x10 <sup>-05</sup>	2.90	2.72	3.08	0.7214
E. antarctica	3.72 x10 <sup>-06</sup>	3.22 x10 <sup>-06</sup>	4.30X10 <sup>-06</sup>	3.27	3.24	3.31	0.9599
G. fraseri	3.53 x10 <sup>-06</sup>	1.31 x10 <sup>-06</sup>	9.51 x10 <sup>-06</sup>	3.24	3.00	3.47	0.8811
G. nicholsi	2.87 x10 <sup>-06</sup>	2.02 x10 <sup>-06</sup>	4.08 x10 <sup>-06</sup>	3.25	3.18	3.33	0.9936
G. braueri	4.58 x10 <sup>-06</sup>	3.60 x10 <sup>-06</sup>	5.82 x10 <sup>-06</sup>	3.11	3.06	3.17	0.9326
K. anderssoni	9.05 x10 <sup>-06</sup>	7.49 x10 <sup>-06</sup>	1.09 x10 <sup>-05</sup>	3.02	2.97	3.07	0.9599
N. achirus	8.14 x10 <sup>-06</sup>	5.17x 10 <sup>-07</sup>	1.28 x10 <sup>-02</sup>	2.49	1.45	3.54	0.4259
P. tenisoni	1.39 x10 <sup>-05</sup>	9.74x 10 <sup>-06</sup>	1.97 x10 <sup>-05</sup>	2.94	2.84	3.03	0.9589
P. bolini	1.98 x10 <sup>-05</sup>	1.34 x10 <sup>-05</sup>	2.92 x10 <sup>-05</sup>	2.88	2.77	2.98	0.8926
P. choriodon	1.27 x10 <sup>-05</sup>	3.24 x10 <sup>-06</sup>	4.94 x10 <sup>-05</sup>	2.98	2.66	3.30	0.8779

4938 Table C2: Number of stomachs collected for each myctophid species during each cruise.

Species	JR161	JR177	JR200	TOTAL
Electrona carlsbergi	80	34	27	141
Electrona antarctica	152	178	112	442
Gymnoscopelus fraseri	11	60	27	98
Gymnoscopelus nicholsi	22	11	7	40
Gymnoscopelus braueri	143	94	109	346
Krefftichthys anderssoni	132	44	22	198
Nannobranchium achirus	23	0	0	23
Protomyctophum tenisoni	27	17	0	44
Protomyctophum bolini	106	76	26	208
Protomyctophum choriodon	0	36	0	36
TOTAL	696	550	330	1576

- 4946 Table C3: Number of individual body mass measurements for each myctophid species during
- 4947 each cruise, from a larger dataset of myctophid sizes (from which only a subset were kept for
- 4948 stomach contents analyses).

Species	JR161	JR177	JR200	TOTAL
E. carlsbergi	195	248	43	486
E. antarctica	568	1023	510	2101
G. fraseri	12	90	41	143
G. nicholsi	30	30	8	68
G. braueri	443	576	429	1448
K. anderssoni	590	162	192	944
N. achirus	43	9	24	76
P. tenisoni	79	98	40	217
P. bolini	261	177	158	596
P. choriodon	0	50	14	64
TOTAL	2221	2463	1459	6143

- 4950 Table C4: Mean abundances for each broad zooplankton taxon, averaged across sampling
- 4951 sites.

Taxon	Mean density (ind./m <sup>2</sup> )	Proportion of total density
Copepoda	13,822.938	0.743
Polychaeta & Chaetognatha	2,645.003	0.142
Pteropoda	1,076.652	0.058
Ostracoda	780.023	0.042
Euphausidae	109.152	0.006
Cnidaria	107.666	0.006
Tunicata	41.255	0.002
Amphipoda	7.266	<0.001
Decapoda	3.692	<0.001
Cephalopoda	0.013	<0.001
Isopoda	0.003	<0.001
Mysidae	0.001	< 0.001

4960 Table C5: Identification of the optimal random effects and variance weighting structure for

- 4961 the models involving predator-prey mass ratio (PPMR). The table displays the various
- 4962 random effects and variance weighting structures for the linear mixed effects models
- 4963 describing the relationship between PPMR and the predictors sea-surface temperature (SST)
- 4964 and surface chlorophyl-a concentration (Chl-a), plus their interaction. The most parsimonious
- 4965 model structure based on Akaike's Information Criterion (AIC) is highlighted in grey. NC
- 4966 indicates models with no convergence.

]	Rando	m effe	ects st	ructur	e			Va	riance	struct	ture			AIC
~1 Species	$\sim \! 1  $ Year	$\sim$ SST Species	~Chl-a Species	$\sim$ SST Year	~Chl-a Year	varIdent(~1 Species)	varIdent( $\sim$ 1 Year)	varFixed(~SST)	varFixed(~Chl-a)	varExp(~SST)	varExp(~Chl-a)	varConst(~SST)	varConst(~Chl-a)	
x x x x x x x x x x x	x x x x x x x x x x x x x	X X	x x	x x x x x	X X X X X									3174.23 2920.04 3167.52 2862.79 2923.84 2921.24 2926.99 3156.83 3158.10 3156.01 NC NC NC
X	X	х		11	1									2852.11
X X X X X X	X X X X X	x x	X X X X	x x	x									2856.31 3000.85 NC NC NC
X	X	X			X									2856.11 2852.11
X X	X X	X X				х								2746.44
x	x	x					х							2837.77
х	х	х				х	Х							2734.96
х	х	х						х						3413.57
х	Х	Х							х					2823.82
х	х	х								х				2849.13
х	х	Х									х			2825.19
х	х	х										х		NC
Х	Х	Х											Х	2815.18

4967

Table C6: The results of a Moran's I test on the residuals from each selected linear mixedeffects model with sea-surface temperature (SST).

Response	Moran's I	P value
Predator-prey mass ratio	0.003	0.339
Predator mass	-0.002	0.656
Prey mass in the diet	-0.001	0.061

Table C7: Identification of the optimal fixed effects structure for the models involving
predator-prey mass ratio (PPMR). The table shows the various fixed effects structures for the
linear mixed effects models describing the relationship between PPMR and the predictors
sea-surface temperature (SST) and surface chlorophyl-a concentration (Chl-a). Each model
includes the optimal random effects and variance weighting structure identified in Table C5.
The most parsimonious model structure based on Akaike's Information Criterion (AIC) and
retaining only significant fixed effects is highlighted in grey.

Fixed effects structure	AIC
SST*Chl-a	2719.88
SST+Chl-a	2718.18
Chl-a	2726.46
SST	2717.16
Null	2725.11

Table C8: Identification of the optimal random effects and variance weighting structure for
the models involving predator body mass. The table shows the random effects and variance
weighting structures for the linear mixed effects models describing the relationship between
predator mass and the predictors sea-surface temperature (SST) and surface chlorophyl-a
concentration (Chl-a), plus their interaction. The most parsimonious model structure based on
Akaike's Information Criterion (AIC) is highlighted in grey.

I	Rando	m effe	ects st	ructur	e		Variance structure							AIC
~1 Species	$\sim$ 1 Year	$\sim$ SST Species	~Chl-a Species	~SST Year	~Chl-a Year	varIdent(~1 Species)	varIdent(~1 Year)	varFixed(~SST)	varFixed(~Chl-a)	varExp(~SST)	varExp(~Chl-a)	varConst(~SST)	varConst(~Chl-a)	
x x	X X													1714.07 807.08 1666.15 733.51
х		Х												759.16
X			X											NC 762.26
х	v	Х	Х	v										762.26 1662.24
	X X			Х	х									1651.06
	X			х	X									1656.18
х	X			X	Λ									737.41
x	x			21	х									737.51
x	x			х	x									743.41
х	х	х												709.89
х	х		х											713.01
х	х	Х	Х											1498.84
х	х		х	х										NC
х	х		Х		Х									716.87
х	х	Х		Х										713.89
X	X	X			X									713.88
Х	х	Х												709.89
Х	х	Х				х								167.11
Х	х	Х					х							650.81
Х	Х	Х				Х	Х							126.01
X	X	X						х						864.08
X	X	X							х	77				729.57 693.14
x x	X X	X X								х	х			706.43
X X	x x	X X									л	х		700.45 NC
X	x x	X X										л	x	700.55

4989Table C9: Identification of the optimal fixed effects structure for the models involving

- 4990 predator mass. The table displays the fixed effects structures for the linear mixed effects
- 4991 models describing the relationship between predator mass and the predictors sea-surface
- 4992 temperature (SST) and surface chlorophyl-a concentration (Chl-a). Each model includes the
- 4993 optimal random effects and variance weighting structure identified in Table C8. The most
- 4994 parsimonious model structure based on Akaike's Information Criterion (AIC) and retaining
- 4995 only significant fixed effects is highlighted in grey.

Fixed effects structure	AIC
SST*Chl-a	105.79
SST+Chl-a	103.83
Chl-a	108.06
SST	102.39
Null	106.49

- 5015 Table C10: Identification of the optimal random effects and variance weighting structure for
- the models involving dietary prey body mass. The table shows the random effects and
- 5017 variance weighting structures for linear mixed effects models describing the relationship
- 5018 between abundance-weighted prey mass in predator diets and the predictors sea-surface
- 5019 temperature (SST) and surface chlorophyl-a concentration (Chl-a), plus their interaction. The
- 5020 most parsimonious model structure based on Akaike's Information Criterion (AIC) is
- 5021 highlighted in grey. NC indicates models with no convergence.

Ι	Rando	m effe	ects st	ructur	e			Va	riance	struct	ure			AIC
~1 Species	~1 Year	$\sim$ SST Species	~Chl-a Species	$\sim$ SST Year	~Chl-a Year	varIdent(~1 Species)	varIdent(~1 Year)	varFixed(~SST)	varFixed(~Chl-a)	varExp(~SST)	varExp(~Chl-a)	varConst(~SST)	varConst(~Chl-a)	
x x x x x	X X	x	x											3227.59 3059.97 3208.49 2909.48 3051.49 NC
x x	X X X X	X	X	X X X	X X									NC 3181.53 3186.04 NC NC
X X X	X X X	X		x	X X									NC NC 2888.69
X X X X	X X X X	X	X X X	x										2909.44 3055.41 NC
X X X	X X X	X X	X	X	x x									NC NC NC
X X X	X X X	X X X				x	X							2888.69 2744.95 2868.50
X X X X	X X X X	X X X X X				X	Х	x	x	x				2732.40 3493.64 2861.30 2869.58
X X X X	X X X X	X X X X								Λ	x	x	x	2869.38 2861.37 2868.64 2851.97

- Table C11: Identification of the optimal fixed effects structure for the models involving
  dietary prey body mass. The table displays the fixed effects structures for the linear mixed
  effects models describing the relationship between abundance-weighted prey mass in
  predator diets and the predictors sea-surface temperature (SST) and surface chlorophyl-a
  concentration (Chl-a). Each model includes the optimal random effects and variance
  weighting structure identified in Table C10. The most parsimonious model structure based on
- 5030 Akaike's Information Criterion (AIC) and excluding non-significant fixed effects is
- 5031 highlighted in grey.

Fixed effects structure	AIC
SST*Chl-a	2717.49
SST+Chl-a	2715.61
Chl-a	2714.59
SST	2713.76
Null	2712.70

Table C12: Identification of the optimal random effects and variance weighting structure for the models involving predator dietary prey size selectivity. The table shows the random effects and variance weighting structures for linear mixed effects models describing the relationship between predator dietary size preference and the interaction between sea-surface temperature (SST) and predator body mass. The most parsimonious model structure based on Akaike's Information Criterion (AIC) is highlighted in grey. NC indicates models with no convergence.

5058		

Ra	ndom	effect	struct	ure	Va	riance	e struc	ture		AIC
~1 Species	~1 Site	~1 Year	~SST Species	~SST Year	varIdent(~1 Species)	varIdent(~1 Year)	varFixed(~SST)	varExp(~SST)	varConst(~SST)	
										288.76
Х										263.06
	Х									268.13
		х								275.76
Х	х									243.02
Х		Х								238.30
	Х	х								266.08
х	х	х								243.80
х			х							264.69
Х				Х						269.55
	х		х							NC
	Х			Х						274.13
Х	Х	х	Х							244.75
х	Х	Х		Х						NC
X	X	X	X	X	+					NC
х		х								238.30
х		х			х					213.95
х		х				х				216.12
Х		Х			х	Х				208.60
х		Х					х			226.08
Х		Х						Х		231.49
х		Х							Х	227.54

Table C13: Identification of the optimal fixed effects structure for the models involving dietary prey size selectivity. The table displays the fixed effects structures for the linear mixed effects models describing the relationship between predator dietary size preference and the interaction between sea-surface temperature (SST) and predator body mass. Each model includes the optimal random effects and variance weighting structure identified in Table C12. The most parsimonious model structure based on Akaike's Information Criterion (AIC) and retaining only significant fixed effects is highlighted in grey.

	Fixed effects structure	AIC	Moran's I	P value
	SST*predator mass	192.34	-0.025	0.515
	SST+predator mass	200.35		
	Predator mass	201.51		
	SST	232.45		
	Null	236.86		
l			-	

5086 Table C14: Identification of the optimal random effects and variance weighting structure for 5087 the models involving predator mass and a larger dataset of myctophid body sizes (n = 6,143). 5088 The table shows the random effects and variance weighting structures for linear mixed effects 5089 models describing the relationship between predator mass and the predictors sea-surface 5090 temperature (SST) and chlorophyll-a concentration (Chl-a), plus their interaction, using the 5091 larger dataset of myctophid body sizes. The most parsimonious model structure based on 5092 Akaike's Information Criterion (AIC) is highlighted in grey. NC indicates models with no 5093 convergence.

Ra	ndon	n effe	ects s	truct	ure			Vari	ance	struc	eture			AIC
Year~1	Species~1	$Year \sim SST$	Species~SST	$Year \sim Chl-a$	Species~Chl-a	varIdent(~1 Year)	varIdent(~1 Species)	varFixed(~SST)	varFixed(~Chl-a)	varExp(~SST)	varExp(~Chl-a)	varConst(~SST)	varConst(~Chl-a)	
														7542.835
х														7286.548 4474.619
v	X													4396.677
X X	Х	х												7178.632
X		Λ		х										7203.550
X		Х		Х										NC
21	х	21	х	21										4307.066
	X				x									NC
	х		х		x									4308.701
х	х	Х												4349.700
x	х			х										7178.632
х	Х	Х		Х										NC
х	Х		Х											4245.069
х	Х				х									4282.719
х	Х		Х		Х									6738.586
х	Х	Х	Х											4249.069
х	Х	Х			X									4283.036
Х	Х		Х	Х										4183.526
X	X			X	X									NC
х	Х		х	Х										4183.526
						Х								4157.655
						-	X							2161.584
						Х	Х							2158.628
								Х	v					5708.168 NC
									Х	v				NC 4129.790
I										Х				4129./90

Х	4184.524
Х	4106.948
Х	4186.680

5095 Table C15: Identification of the optimal fixed effects structure for the models involving

5096 predator mass and a larger dataset of myctophid body sizes (n = 6,143). The table displays the

5097 fixed effects structures for the linear mixed effects model describing the relationship between

5098 predator size and the predictors sea-surface temperature (SST) and surface chlorophyll-a

5099 concentration (Chl-a), plus their interaction, using the larger dataset of myctophid body sizes.

5100 Each model includes the optimal random effects and variance weighting structure identified

5101 in Table C14. The most parsimonious model structure based on Akaike's Information

5102 Criterion (AIC) and retaining only significant fixed effects is highlighted in grey. The result

5103 of a Moran's I test for spatial autocorrelation for the optimal model is also provided.

Fixed effects structure	AIC	Moran's I	P-value
SST*Chl-a	2330.480		
SST+Chl-a	2328.769		
Chl-a	2426.262		
SST	2326.877	0.001	0.161
Null	2424.297		

5104

Table C16: Model statistics for the optimal linear mixed effects model identified in Tables C14-C15 describing the relationship between sea-surface temperature (SST) and predator body mass using a larger dataset of myctophid body sizes (n = 6,143).

	Coefficient	Estimate	SE	DF	t-value	p-value
	Intercept	0.456	0.072	6113	6.351	< 0.0001
	SST	-0.026	0.002	6113	-10.397	< 0.0001
5108						
5109						
5110						
5111						
5112						
5113						
5114						
5115						

Table C17: Identification of the optimal random effects and variance weighting structure for the models involving predator mass for each myctophid species. The table shows the random effects and variance weighting structures for linear mixed effects models describing the relationship between predator mass and sea-surface temperature (SST) for each species, using a larger dataset of myctophid body sizes (n = 6,143). The most parsimonious model based on Akaike's Information Criterion (AIC) is highlighted in grey. NC indicates models with no convergence.

Species	ef	Random effect structure		riance	struc	ture	AIC
	Year~1	Year~ sst	varident(~1 Year)	varfixed(~SST)	varexp(~SST)	varConst(~SST)	
E. carlsbergi	x x	x	x				- 1288.456 - 1287.623 NC - 1318.993
				X	X	X	-1096.769 -1309.503 -1308.141
	x						1620.508 1615.725
E. antarctica	X X	х	x				1619.725 1617.816
	X X X			X	x	v	2093.591 1590.430 1557.973
	Λ					X	51.531
	X						44.042 NC
G. fraseri	X X	Х	x				42.658
	x			х			50.821
	X				х		45.530
	X					X	45.833 119.246
	x						120.740
	х	Х					NC
G. nicholsi			х	v			118.141 94.101
				Х	х		67.816
						x	90.010
G. braueri							1457.564

	I		T				1405 256
	X						1405.256
	х	Х					1407.795
	х		Х				1396.838
	Х			Х			1670.105
	х				Х		1365.444
	X					Х	1370.079
							942.875
	х						944.619
<b>.</b>	х	Х					NC
K. anderssoni			х				840.444
				Х			915.442
					Х		866.609
	ļ					Х	880.390
							-47.476
	Х						-45.476
	Х	Х					-41.476
N. achirus			Х				-47.247
				Х			-2.504
					Х		-57.541
						Х	-55.363
							-136.615
	х						-428.244
	х	Х					NC
P. tenisoni			х				-443.422
				Х			-427.005
					Х		-443.606
						Х	-459.302
							-44.103
	х						-55.174
	х	Х					NC
P. bolini	х		х				-63.431
	х			Х			-21.393
	х				Х		-64.357
	х					Х	-70.513
[							-77.654
	х						-75.654
	x	х					NC
P. choriodon			х				-75.660
				х			-81.037
					Х		-79.060
					-	х	NC
L	1		1				

5125 Table C18: Identification of the optimal fixed effects structure for the models involving

5126 predator mass for each myctophid species. The table shows the fixed effects structures for the

5127 linear mixed effects models of the relationship between predator size and sea-surface

5128 temperature (SST) for each species, using a larger dataset of myctophid body sizes (n =

5129 6,143). Each model includes the optimal random effects and variance weighting structure

5130 identified in Table C17. The results of a Moran's I test for spatial autocorrelation are provided

5131 for the optimal models, along with the optimal correlation structure implemented based on

5132 AIC for any models with significant autocorrelation. No adjustments were made for multiple

5133 comparisons.

Species	Fixed effects	AIC	Moran's I	Moran's I	Autocorrelation
	structure			p-value	structure
E. carlsbergi	SST	-1340.507			
E. curisbergi	Null	-1342.335	< 0.001	0.7174	
E. antarctica	SST	1542.642	0.026	< 0.0001	Rational
E. aniarciica	Null	1569.967			
C fuggari	SST	34.019	0.060	0.0038	Exponential
G. fraseri	Null	68.081			
G. nicholsi	SST	55.946	0.004	0.6894	
G. nicholsi	Null	71.849			
C. harmani	SST	1356.068	0.009	0.0175	Exponential
G. braueri	Null	1366.125			-
V and anna ani	SST	824.964	-0.010	0.0342	Spherical
K. anderssoni	Null	1001.208			
N. achirus	SST	-72.145			
N. achirus	Null	-73.502	-0.104	0.0914	
D ( in i	SST	-469.989	-0.004	0.9498	
P. tenisoni	Null	-442.164			
D halini	SST	-83.925	-0.013	0.0937	
P. bolini	Null	-76.457			
Daharialan	SST	-93.411	-0.021	0.9246	
P. choriodon	Null	-78.834			

5134

- 5136 Table C19: Outputs of the optimal linear mixed effects models identified in Tables C17-C18,
- 5137 describing the relationship between predator mass and sea-surface temperature (SST) for each
- 5138 species, using a larger dataset of myctophid body sizes (n = 6,143).

Species	Coefficient	Estimate	SE	DF	t-value	p-value
E. carlsbergi	Intercept	0.790	0.003	486	298.685	< 0.0001
E motorestica	Intercept	0.579	0.021	2097	27.425	< 0.0001
E. antarctica	SST	-0.022	0.009	2097	-2.479	0.0148
C. funnari	Intercept	0.887	0.121	139	7.353	< 0.0001
G. fraseri	SST	-0.100	0.028	139	-3.536	0.0011
C wish alai	Intercept	1.390	0.030	68	45.721	< 0.0001
G. nicholsi	SST	-0.083	0.019	68	-4.478	< 0.0001
G. braueri	Intercept	0.754	0.079	1444	9.578	< 0.0001
G. bruueri	SST	-0.083	0.007	1444	-11.213	< 0.0001
K. anderssoni	Intercept	0.154	0.019	944	8.287	< 0.0001
K. underssoni	SST	-0.085	0.006	944	-14.086	< 0.0001
N. achirus	Intercept	0.153	0.015	76	9.965	< 0.0001
P. tenisoni	Intercept	-0.294	0.153	213	-1.925	0.0555
F. lenisoni	SST	0.030	0.005	213	6.484	< 0.0001
P. bolini	Intercept	0.069	0.038	592	1.835	0.0670
1.001111	SST	-0.017	0.005	592	-3.406	0.0007
P. choriodon	Intercept	0.800	0.035	64	22.598	< 0.0001
F. Chorlodon	SST	-0.050	0.009	64	-5.464	< 0.0001

- Table C20: Identification of the optimal random effects and variance weighting structure for the models involving myctophid species diversity. The table shows the random effects and variance weighting structures for the linear mixed effects models describing the relationship between myctophid species diversity (Shannon-Wiener index) and the interaction between sea-surface temperature (SST) and surface chlorophyll-a concentration (Chl-a). The most parsimonious model structure based on Akaike's Information Criterion (AIC) is highlighted
- 5160 in grey. NC indicates models with no convergence.

e	ando ffect ructu	s		Variance structure						AIC
Year~1	Year~ SST	Year~ Chl-a	varIdent(~1 Year	varFixed(~SST)	varFixed(~Chl-a)	varExp(~SST)	varExp(~Chl-a)	varConst(~SST)	varConst(~Chl-a)	
										134.843
Х										136.498
Х	Х									137.938
Х		Х								139.111
Х	х	X								NC
			Х							131.819
				Х						169.6902
					х					155.0526
						х				132.8421
							х			132.0601
								х		137.1728
									Х	129.3736

- 5170 Table C21: Identification of the optimal fixed effects structure for the models involving
- 5171 myctophid species diversity. The table displays the fixed effects structures for the linear
- 5172 mixed effects models of the relationship between Shannon-Wiener diversity and the
- 5173 interaction between sea-surface temperature (SST) and surface chlorophyll-a concentration
- 5174 (Chl-a). Each model includes the optimal random effects and variance weighting structure
- 5175 identified in Table C20. The most parsimonious model structure based on Akaike's
- 5176 Information Criterion (AIC) and retaining only significant fixed effects is highlighted in grey.
- 5177 The results of a Moran's I test for spatial autocorrelation is provided for the optimal model.

Fixed effects structure	AIC	Moran's I	P-value
SST*Chl-a	114.7948		
SST+Chl-a	115.1217		
Chl-a	148.1829		
SST	114.1337	0.071	0.103
Null	146.3198		

- 5179 Table C22: Model statistics for the optimal linear mixed effects model identified in Tables
- 5180 C20-C21 describing the relationship between myctophid species diversity and sea-surface
- 5181 temperature (SST).

Coefficient	Estimate	SE	DF	t-value	p-value
Intercept	0.962	0.045	115	21.545	< 0.0001
SST	0.090	0.015	115	6.080	< 0.0001

5182

5183

- 5185 Table C23: Identification of the optimal random effects and variance weighting structure for
- 5186 the models involving predator-prey mass ratio (PPMR) and temperature at 1,062 m depth.
- 5187 The table shows the random effects and variance weighting structures for the linear mixed
- 5188 effects models describing the relationship between PPMR and the predictors temperature at
- 5189 1,062m depth (TAD) and surface chlorophyl-a concentration (Chl-a), plus their interaction.
- 5190 The most parsimonious model structure based on Akaike's Information Criterion (AIC) is
- 5191 highlighted in grey. NC indicates models with no convergence.

I	Rando	m effe	ects st	ructur	e			Va	riance	struct	ure			AIC
Species~1	Year~1	Species~TAD	Species~Chl-a	$Y ear \sim TAD$	Y ear~Chl-a	varIdent(~1 Species)	varIdent( $\sim$ 1 Year)	varFixed(~TAD)	varFixed(~Chl-a)	varExp(~TAD)	varExp(~Chl-a)	varConst(~TAD)	varConst(~Chl-a)	
X X X X X X	X X X X X	x x	X X	X	X									3178.22 2942.28 3173.63 2883.36 2945.36 2943.27 2945.37 3161.19 3161.62 3153.99
x x x	X X X X			x x x	x x x									NC NC NC
X X X X X X	X X X X X X	X X	X X X X X	x	x									2865.27 2877.08 2994.12 NC NC
X X X	X X X	X X X		X	X									NC NC 2865.27
X X X X	X X X X	X X X X				X X	X X	x						2762.27 2852.23 2751.60 6959.02
X X X	X X X	X X X							X	x	x	V		2836.48 2811.59 2839.16
X X	X X	X X										Х	х	2833.85 2828.41

- 5194 Table C24: Identification of the optimal fixed effects structure for the models involving
- 5195 predator-prey mass ratio (PPMR) and temperature at 1,062 m depth. The table displays the
- 5196 fixed effects structures for the linear mixed effects models describing the relationship
- 5197 between PPMR and the predictors temperature at 1,062m depth (TAD) and surface
- 5198 chlorophyl-a concentration (Chl-a). Each model includes the optimal random effects and
- 5199 variance weighting structure identified in Table C23. The most parsimonious model structure
- 5200 based on Akaike's Information Criterion (AIC) and retaining only significant fixed effects is
- 5201 highlighted in grey.

Fixed effects structure	AIC
TAD*Chl-a	2740.15
TAD+Chl-a	2738.21
Chl-a	2740.84
TAD	2739.64
Null	2741.57

5221 Table C25: Identification of the optimal random effects and variance weighting structure for 5222 the models involving predator body mass and temperature at 1,062 m depth. The table shows

- 5223 the random effects and variance weighting structures for the linear mixed effects models
- 5224 describing the relationship between predator mass and the predictors temperature at 1,062m
- 5225 depth (TAD) and surface chlorophyl-a concentration (Chl-a), plus their interaction. The most
- 5226 parsimonious model structure based on Akaike's Information Criterion (AIC) is highlighted
- 5227 in grey. NC indicates models with no convergence.

I	Rando	m effe	ects st	ructur	e			Va	riance	struct	ure			AIC
Species~1	Y ear~1	Species~TAD	Species~Chl-a	$Y ear \sim TAD$	Y ear~Chl-a	varIdent(~1 Species)	varIdent( $\sim$ 1 Year)	varFixed(~TAD)	varFixed(~Chl-a)	varExp(~TAD)	varExp(~Chl-a)	varConst(~TAD)	varConst(~Chl-a)	
X X X X X	X X X X X X	x x	x x	X X	x x									1671.26 822.32 1671.09 734.48 780.53 NC 780.29 NC 1659.41 NC NC
X X	X X			х	х									NC
х	х			х	Х									NC
Х	Х	Х												691.51
X	X		X											712.97 NC
X X	X	Х	X X	v										NC NC
X	X X		X	х	х									NC
X	X	х	л	х	л									NC
X	X	X		Λ	х									NC
X	X	X												691.51
x	X	x				х								155.92
x	x	X					х							636.45
x	Х	х				х	х							113.34
х	х	х						х						3278.38
х	х	х							х					703.20
х	х	х								х				677.66
х	х	х									х			686.63
х	х	х										х		679.64
Х	Х	Х											х	661.70

5228

- 5230 Table C26: Identification of the optimal fixed effects structure for the models involving
- 5231 predator mass and temperature at 1,062 m depth. The table displays the fixed effects
- 5232 structures for the linear mixed effects models describing the relationship between predator
- 5233 mass and the predictors temperature at 1,062m depth (TAD) and surface chlorophyl-a
- 5234 concentration (Chl-a). Each model includes the optimal random effects and variance
- 5235 weighting structure identified in Table C25. The most parsimonious model structure based on
- 5236 Akaike's Information Criterion (AIC) and retaining only significant fixed effects is
- 5237 highlighted in grey.

Fixed effects structure	AIC
TAD*Chl-a	96.84
TAD+Chl-a	95.57
Chl-a	100.62
TAD	95.07
Null	99.88

- 5240 Table C27: Identification of the optimal random effects and variance weighting structure for
- 5241 the models involving dietary prey body mass and temperature at 1,062 m depth. The table
- shows the random effects and variance weighting structures for linear mixed effects models
- 5243 describing the relationship between abundance-weighted prey mass in predator diets and the
- 5244 predictors temperature at 1,062m depth (TAD) and surface chlorophyl-a concentration (Chl-
- a), plus their interaction. The most parsimonious model structure based on Akaike's
- 5246 Information Criterion (AIC) is highlighted in grey. NC indicates models with no
- 5247 convergence.

x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x     x     x     x       x     x     x
x       3057.92         x       3187.10         x       x
x     x     x     2881.77       x     x     x     2913.04
x x x 2913.04
x     x     x     x       x     x     x     x       x     x     x     x       x     x     x     x
<u>x x x x</u> <u>2885.77</u>
x     x     x     2881.77       x     x     x     2741.32       x     x     x     2861.62       x     x     x     2728.55
x x x x x 7247.49
x x x x 2855.30
x x x x 2782.81
x x x x 2855.72
x x x x 2818.68 x x x x x 2845.75

5249

- 5251 Table C28: Identification of the optimal fixed effects structure for the models involving
- 5252 dietary prey mass and temperature at 1,062 m depth. The table shows the fixed effects
- 5253 structures for the linear mixed effects models describing the relationship between abundance-
- 5254 weighted prey mass in predator diets and the predictors temperature at 1,062m depth (TAD)
- 5255 and surface chlorophyl-a concentration (Chl-a). Each model includes the optimal random
- 5256 effects and variance weighting structure identified in Table C27. The most parsimonious
- 5257 model structure based on Akaike's Information Criterion (AIC) and excluding non-significant
- 5258 fixed effects is highlighted in grey.

Fixed effects structure	AIC
TAD*Chl-a	2717.22
TAD +Chl-a	2715.43
Chl-a	2713.89
TAD	2713.79
Null	2712.36

Table C29: Model statistics for the effects of temperature at 1,062 m depth on predator and prey body masses. Output from linear mixed effects models with predator-prey mass ratio (PPMR), predator body mass and abundance-weighted average prey body mass in predator stomachs as response variables. The temperature variable represents temperature at ~1,062m depth (TAD), while R<sup>2</sup>m and R<sup>2</sup>c are the Nakagawa's marginal and conditional model R<sup>2</sup> values, respectively.

Model	Coefficient	Estimate	SE	DF	t-value	p-value
PPMR	Intercept	3.017	0.109	1550	27.717	< 0.0001
	TAD	-0.131	0.050	1550	-2.596	0.0095
$R^2m = 0.025, R^2c = 0.489$						
Predator body mass	Intercept	0.566	0.108	1550	5.257	< 0.0001
	TAD	-0.071	0.036	1550	-1.984	0.0475
$R^2m = 0.012, R^2c = 0.980$						
Mean prey body mass $R^2m < 0.001$ , $R^2c = 0.476$	Intercept	-2.357	0.072	1551	-32.600	< 0.0001

5266

- 5267 Table C30: The results of a Moran's I test on the residuals from each selected linear mixed
- 5268 effects model with the temperature at 1,062m depth as the predictor variable.

Response	Moran's I	p value
Predator-prey mass ratio	0.003	0.359
Predator mass	0.001	0.693
Prey mass in diet	0.003	0.312

# 5270 Appendix D: Supplementary material for chapter 5

5271 D1: Supplementary methods

#### 5272 6.2.5 Re-aggregation of functional groups

5273 Groups were reaggregated using information on biomass and rate parameters available in the 5274 supplementary materials for each model. When aggregating groups, the new group biomass 5275 was determined by a simple sum of the biomasses of the groups being combined. The final 5276 aggregated rate parameters (P/B and Q/B) were averages of the original group-specific 5277 parameters, calculated using the group biomasses as weightings. In cases where groups were 5278 split to form multiple new groups, the appropriate biomass was assigned to each new group 5279 using biomass information in the relevant supplementary material for the published model or 5280 wider literature for that region. Unique rate parameters were assigned to the new groups 5281 where available in the published supporting information, otherwise the new groups retained 5282 the rate estimates from their parent group. Below, we provide an overview of the steps taken 5283 to reaggregate each functional group in each regional model.

#### 5284 Whales:

5285 In each model, we reaggregated whale groups to form four functional groups: Toothed 5286 whales, humpback whales, minke whales and "other baleen whales".

5287 In the Prydz Bay model, minke whales were already separated and the humpback whale

5288 group was extracted from the "baleen whale" group and parameterized using species-level

5289 biomass and Q/B information from the supplementary, while retaining the P/B value

specified for the original "baleen whale" group. Orca and sperm whales were aggregated intoa single toothed whale group.

6 6 I

5292 The Ross Sea model had very similar whale groups to the Prydz Bay model with an

5293 additional "toothed whales" group representing two beaked whale species. The same

5294 reaggregation steps were taken as for the Prydz Bay model, while "toothed whales", sperm

5295 whales and orcas were combined into a single new "toothed whales" group. In this model,

5296 P/B and Q/B estimates were available for each species and were used in the reaggregation.

- 5297 The Prince Edward Islands model represented only orca with no other whale species
- 5298 included, due to their extremely low abundance in the model region. To ensure comparability

with other models, we added minke whale, humpback whale and 'other baleen whale' groupsbut kept their initial biomasses at zero.

The Kerguelen Plateau model included a toothed whale group (representing southern bottlenose whales and hourglass dolphins), orcas, sperm whales and a baleen whale group representing fin and southern right whales. The only change made to this model was to add the humpback and minke whale groups, again with a negligible biomass to reflect their extremely low current abundance in the model region, and to combine the orcas, sperm whales and 'toothed whales' into a single group.

5307 The South Georgia model included two whale groups: toothed and baleen. Humpback and 5308 minke whale groups were extracted from the baleen whale group using biomass and Q/B 5309 estimates for each species available in the supplementary, while they retained the P/B 5310 estimate from the baleen whale group.

5311 The Antarctic Peninsula model represented each whale species as a separate group. Orca and

- 5312 sperm whales were aggregated into the toothed whale group while baleen whales excluding
- 5313 humpbacks and minkes were aggregated into the "Other baleen whales" group.

# 5314 <u>Seals and penguins:</u>

5315 In most models, seals and penguins were represented as individual species. These were

5316 aggregated into a single seal or penguin group in each model. The exception was the

5317 Kerguelen Plateau model, which already aggregated penguins into a single group and

5318 therefore required no changes.

# 5319 Flying birds:

5320 Flying birds were represented to a varying degree of taxonomic resolution across models,

ranging from a single functional group to multiple groups representing different feeding

5322 guilds, taxonomic distinctions (e.g. albatrosses versus other birds) or even individual species.

- 5323 Models which included multiple flying bird groups were reaggregated to have a single
- 5324 "Flying birds" group.

5325 <u>Fish:</u>

5326 All models included multiple fish groups specified to varying levels of taxonomic resolution.

- 5327 In each model, the groups were combined based on their habitat association to generate two
- 5328 broad functional groups representing demersal and pelagic fish.

#### 5329 <u>Squid:</u>

5330 Squid are known to be poorly represented in most Southern Ocean ecosystem models, due to

- a lack of information regarding their regional biomass and ecology (REFS). All models
- 5332 included a squid group, with the Prince Edward Islands model distinguishing this group by
- size. In this latter model the two size classes were aggregated into a single squid group, while
- 5334 no changes were required for the other models.

#### 5335 Zooplankton:

5336 We sought to distinguish between Antarctic krill and all other krill. The Prydz Bay, Ross Sea 5337 and Antarctic Peninsula models already provided separate Antarctic krill and other euphausiid 5338 groups, though the species represented by the latter differed between models. The Kerguelen 5339 Plateau model included a single krill group representing euphausiids other than Euphausia 5340 superba, as this group is not found in any meaningful numbers in the model region (Yang et 5341 al. 2022; Yang et al. 2020). The Prince Edward Islands model did not explicitly model any 5342 krill group but according to Hill et al. (2021) the model authors identified that euphausiids 5343 make up 88% of the "large crustacean zooplankton" and 20% of the "small crustacean 5344 zooplankton" groups. As with the Kerguelen Plateau model, E. superba are not present in the 5345 model region (Yang et al. 2022; Yang et al. 2020), therefore we redistributed the biomass of 5346 the relevant zooplankton groups to produce a single "other krill" group using the above 5347 proportions. The South Georgia model included an Antarctic krill group but included other 5348 euphausiids in the "Carnivorous macrozooplankton". We estimated the average relative 5349 biomass of other euphausiid species in the macrozooplankton from zooplankton samples taken near South Georgia in 2006, 2008 and 2009 during the Discovery cruises (Tarling et al. 5350 5351 2012a). The average proportion of macrozooplankton that was euphausiids was 6.9%, and we 5352 extracted the relevant biomass from the "Carnivorous macrozooplankton" group in the South 5353 Georgia model and assigned it to a new "Other krill" group.

5354 Three models explicitly included salps and therefore required no further reaggregation to

- represent this group. In the Prince Edward Islands model, salps represent 5% of the "other
- 5356 zooplankton" group, while in the Ross Sea, salps are estimated to make up 2.6% of
- 5357 macrozooplankton biomass (Hill et al. 2021). The proportion of salps in the zooplankton
- 5358 groups in the Kerguelen Plateau model was not clear, but the source used by the authors to
- 5359 estimate macrozooplankton biomass identifies that salps display extremely patchy
- 5360 distributions in the region, representing between 0.1% and 7% of total macrozooplankton

5361 biomass (Hunt et al. 2011). In the absence of better estimates, we assumed that 7% of 5362 macrozooplankton biomass was salps. Salp relative abundance is unlikely to be equivalent to 5363 relative biomass as salps are heavier than many other macrozooplankton taxa, but given their 5364 patchy distribution we determined that setting their relative biomass to be the maximum 5365 estimate of their relative abundance would reasonably capture their approximate biomass in 5366 the study region, though with high associated uncertainty. In each of these models, we 5367 extracted the relevant biomass from the original groups and assigned these to a new salp 5368 group.

5369 We grouped all remaining zooplankton into three size-based functional groups:

5370 macrozooplankton, mesozooplankton and microzooplankton. Three models already had this

aggregation scheme, and the only adjustments needed were to subtract the estimated salp

5372 biomass from the Kerguelen Plateau model. The Ross Sea model included macro- and

5373 mesozooplankton groups, but further distinguished flagellates and ice metazoa and protozoa.

5374 We merged the mesozooplankton and ice metazoan groups into an aggregate

5375 mesozooplankton group, and combined the flagellates and ice protozoa with the heterotrophic

5376 microplankton to form a single microzooplankton group. We spit the zooplankton groups in

the Prince Edward Islands model following (Hill et al. 2021): macrozooplankton are 12%

5378 large crustacean zooplankton, 20% small crustacean zooplankton, 48% other zooplankton;

5379 mesozooplankton are 49% small crustacean zooplankton, 43% other zooplankton;

5380 microzooplankton are 11% small crustacean zooplankton, 4% other zooplankton. The South

5381 Georgia model already incorporated macrozooplankton (carnivorous macrozooplankton

5382 minus the biomass representing other krill) and microzooplankton (heterotrophic

5383 microzooplankton), and we aggregated the two remaining zooplankton groups ("herbivorous

5384 mesozooplankton" and "herbivorous and detritivorous copepods") into a single

5385 mesozooplankton group.

5386 <u>Benthos:</u>

All models included at least one benthic functional group. In models that include more than
one such group (Ross Sea – megabenthos, macrobenthos and meiobenthos; Prince Edward

5389 Islands – benthos and benthic decapods) these were combined into a single benthos group.

5390 <u>Bacteria:</u>

5391 Three models (PB, SG, KP) explicitly modelled bacteria as a single functional group.

5392 Another (RS) further split bacteria into ice, water-column and sediment-associated bacterial

- groups, which we aggregated into a single group. We did not add bacterial groups to the tworemaining models (AP and PE).
- 5395 <u>Primary producers:</u>
- 5396 We continued to distinguish between the sea ice algae and other producers in the RS and AP
- 5397 models. All other producer groups were aggregated into a single primary producer group.
- 5398 Detritus:
- 5399 All detrital and carcass groups were aggregated into a single detritus group in each model.
- 5400 6.2.6 Automated balancing routine:

5401 The primary input parameters for the automated balancing algorithm were B, P/B and Q/B5402 and DC. Additionally, the assimilation efficiencies of each group were obtained from 5403 (Pinkerton and Bradford-Grieve 2010) and held constant across model regions and 5404 throughout the balancing process. The balancing algorithm first determines the relative 5405 change to each group's B by a random draw from a normal distribution with mean = 0 and 5406 SD = 0.05. Changes to B are therefore small and within a local region around the values at 5407 the current step. The final biomass values of each group must remain within the bounds of 5408 their pedigree, which was set using the standard Ecopath data pedigree approach (see Table 5409 S4). This information was already provided for some of the models (RS, PE, SG), while for 5410 the remaining models we assigned pedigrees based on a review of the supplementary 5411 information accompanying each published model. The exception to this is the primary producers, for which we set the *EE* values to 0.5 and therefore *B* is calculated based on 5412

5413 consumer demand.

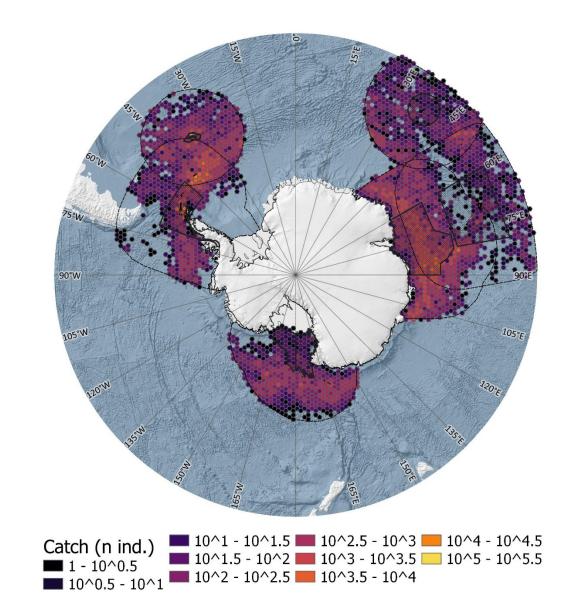
5414 Next, a growth efficiency (GE) value is drawn from a uniform distribution between a 5415 minimum and maximum value determined by group type, based on values in Townsend et al. 5416 (2008): endotherm vertebrate (0.001 - 0.05); ectotherm vertebrate (0.05 - 0.2); invertebrate 5417 (0.1 - 0.4); bacteria (0.2 - 0.5). This step is included to ensure that energetic parameters 5418 comply with general expected ecological relationships. By randomly varying this parameter, 5419 rather than fixing it at some predetermined value, we provide further exploration of the range 5420 of possible model parameterisations. P/B and O/B values are varied within a range of 50% on 5421 either side of the model-averaged input values. Because GE is determined by the combination 5422 of P/B and O/B it is only possible to set one of these energetic parameters directly, with the 5423 other estimated from the combination with GE. To avoid potential bias by only varying one

5424 of these parameters throughout, our algorithm randomly determines, for each group, which of 5425 these energetic parameters is randomly sampled at that step. The relevant P/B or Q/B values 5426 are then sampled randomly from a uniform distribution within their specified bounds and 5427 used in combination with *GE* to estimate the missing parameters. The exception to this was 5428 whale Q/B, which was fixed at the lower values calculated from prior estimates of whale 5429 consumption to ensure consistency in starting whale consumption across model versions and 5430 regions for later analyses.

5431 Random adjustments to the diet composition of each group are then also made. These are 5432 achieved by resampling their input balanced diets from a Dirilichet distribution, which 5433 ensures that the resulting diet compositions sum to one. A further constraint on the diet 5434 resampling is the inclusion of a scaling parameter which restricts the changes to each dietary 5435 component, such that diets do not become completely scrambled (i.e. common prey remain 5436 common and rare prey remain rare). We found a scaling factor of 50 to provide a good 5437 balance between allowing sufficient variation to explore a broad range of plausible prev 5438 compositions and preventing excessive reshuffling of diets. For example, for a prey 5439 representing 50% of a consumer's diet, a scaling factor of 50 generates values which generally lie within 20% of the original input, with minimum and maximum tails of around 5440 5441 40%. Higher values of the scaling factor increasingly restrict the possible dietary changes 5442 (Figure D14).

5443 Once the algorithm has generated a new set of suitable parameter values, the model is 5444 evaluated by calculating an objective function, set as the sum of *EE* for all out of balance at 5445 that step. This model is accepted if the objective function is lower than that of the previous 5446 step. In this case, the biomass values are carried over and used as the basis for determining 5447 biomasses at the next step (all other parameters are varied from scratch at each step and 5448 therefore do not need to be updated). If the objective function for the new parameter set is 5449 higher than that of the previous set, the changes are rejected and the biomass values reset to 5450 those of the previous step. To prevent the algorithm from becoming stuck in local optima, 5451 steps with higher (worse) objective functions are accepted with a specific probability (P), 5452 which is kept constant throughout. The algorithm runs for a specified number of steps (or 5453 until a balanced model is found), and a record of the best model (lowest objective function) is 5454 kept and updated at each step.

- 5455 If no balanced model is identified after the specified number of steps, the algorithm switches 5456 to targeted, primarily small, adjustments to biomasses and diets to achieve balance. The 5457 biomass of the group most out of balance is increased by a small amount and the biomass of 5458 the maximal predator of that group is reduced slightly. The diet matrix of the maximal 5459 predator is adjusted to slightly reduce the contribution of the focal prey group, with the 5460 missing diet contribution randomly redistributed across other prey groups (excluding 5461 cannibalistic interactions). The biomass values for the final balanced model are then checked, 5462 and the model is rejected if any biomass values fall outside the acceptable bounds set by the 5463 pedigree CV. The full balancing process was repeated until a suite of 1000 versions of each
- 5464 regional input model had been generated.

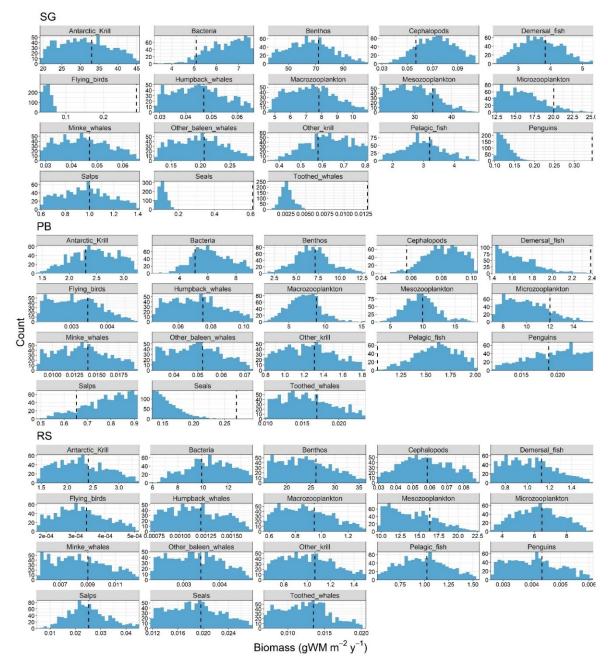


5467

5468 Figure D1: Map of all baleen whale catches (number of individuals) within each of the model

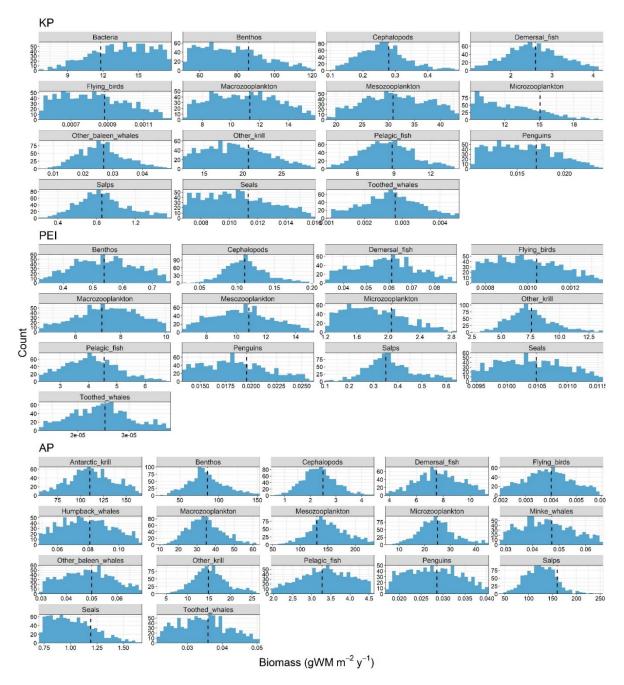
regions (solid polygons) and their 1000km buffers (dashed lines). Catches are on a log scale.

5470 Hex tiles are  $100 \text{km}^2$ .



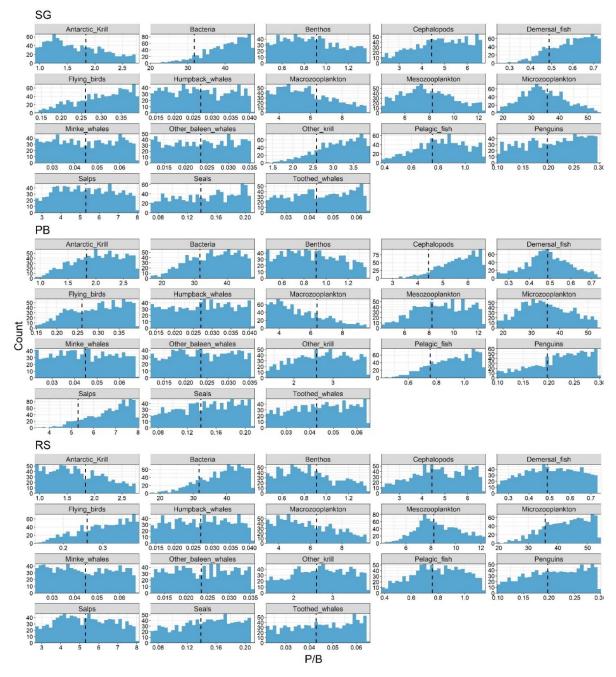
5472 Figure D2: Distribution of biomasses in the balanced versions of each model (*n*=1000), for

- 5473 PB, RS and SG. Vertical dashed lines display the original value for each parameter in the
- 5474 standardised, reaggregated but unbalanced version of each model.



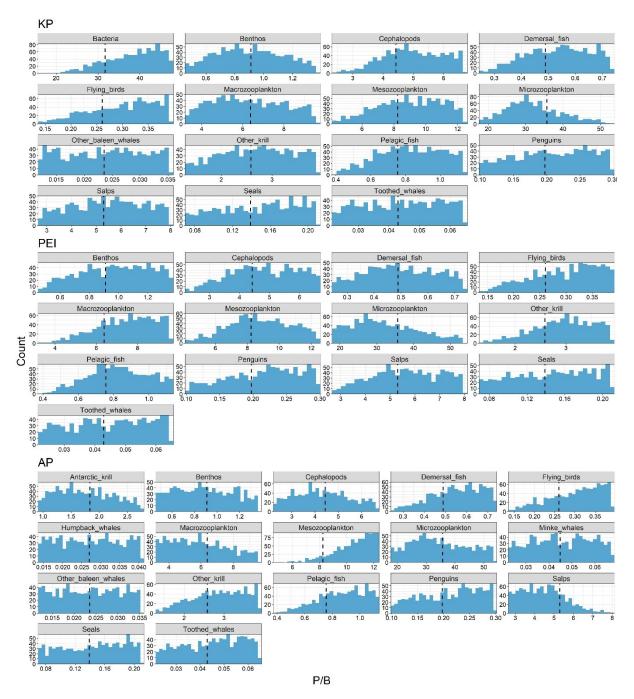
5477 Figure D3: Distribution of biomasses in the balanced versions of each model (*n*=1000), for

- 5478 KP, PE and AP. Vertical dashed lines display the original value for each parameter in the
- 5479 standardised, reaggregated but unbalanced version of each model.



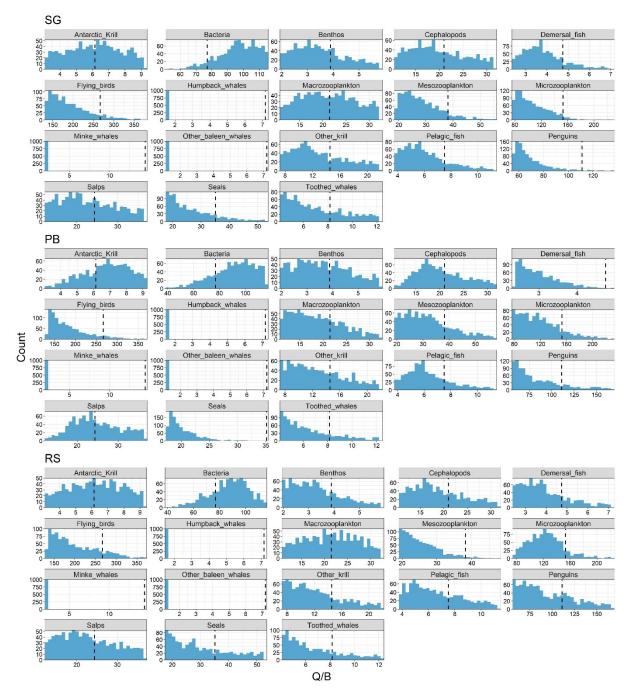
5482 Figure D4: Distribution of production over biomass (P/B) in the balanced versions of each 5483 model (n=1000), for SG, PB and RS. Vertical dashed lines display the original value for each 5484 parameter in the standardised, reaggregated but unbalanced version of each model.

5485



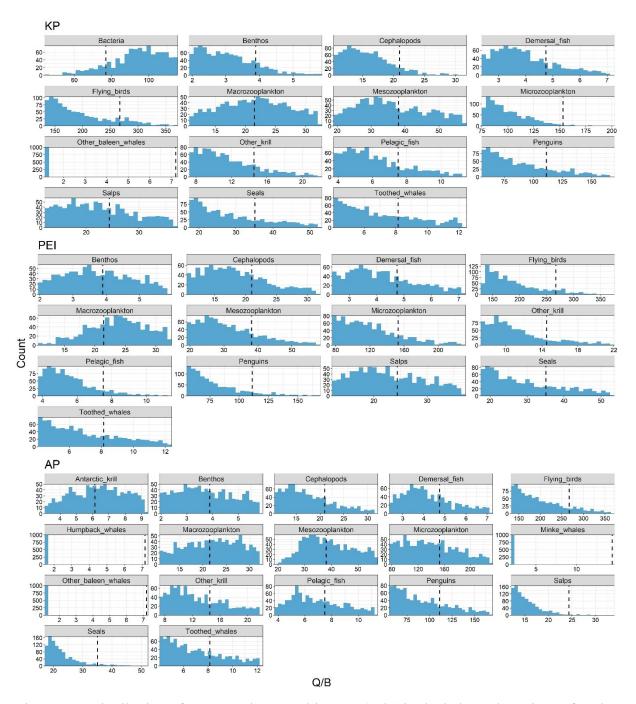
5488 Figure D5: Distribution of production over biomass (*P/B*) in the balanced versions of each

- 5489 model (*n*=1000), for KP, PE and AP. Vertical dashed lines display the original value for each
- 5490 parameter in the standardised, reaggregated but unbalanced version of each model.



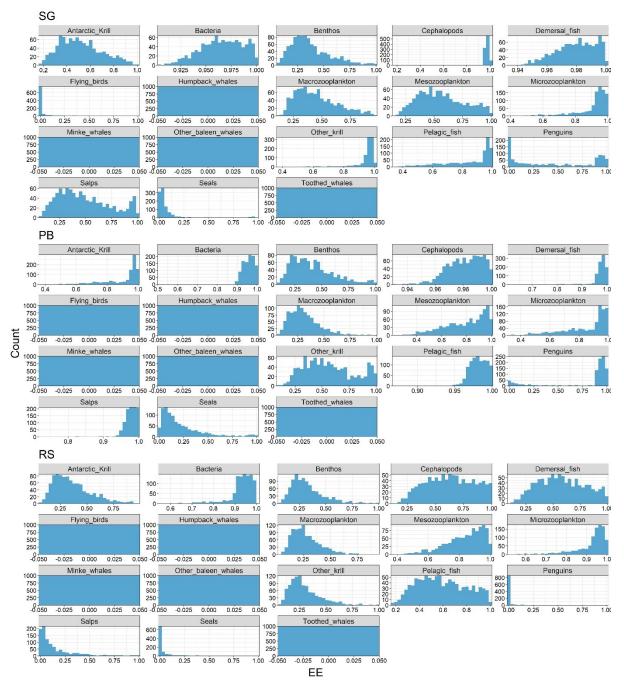
5493 Figure D6: Distribution of consumption over biomass (Q/B) in the balanced versions of each 5494 model (*n*=1000), for SG, PB, RS. Vertical dashed lines display the original value for each 5495 parameter in the standardised, reaggregated but unbalanced version of each model.

5496



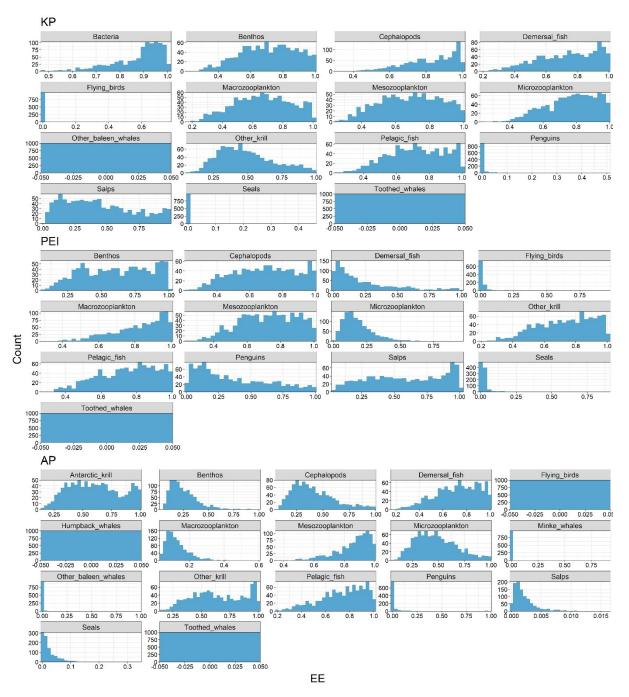
5499 Figure D7: Distribution of consumption over biomass (Q/B) in the balanced versions of each 5500 model (*n*=1000), for KP, PE and AP. Vertical dashed lines display the original value for each

5501 parameter in the standardised, reaggregated but unbalanced version of each model.



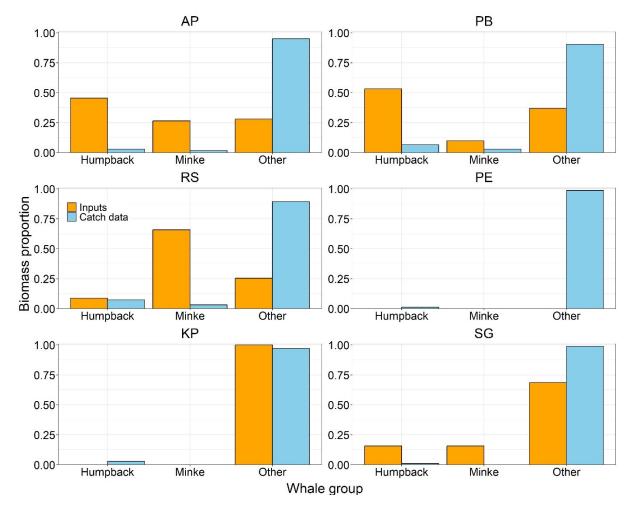
5503 Figure D8: Distribution of ecotrophic efficiency (*EE*) in the balanced versions of each model

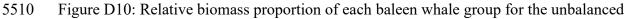
5504 (*n*=1000), for SG, PB and RS.



5507 Figure D9: Distribution of ecotrophic efficiency (*EE*) in the balanced versions of each model

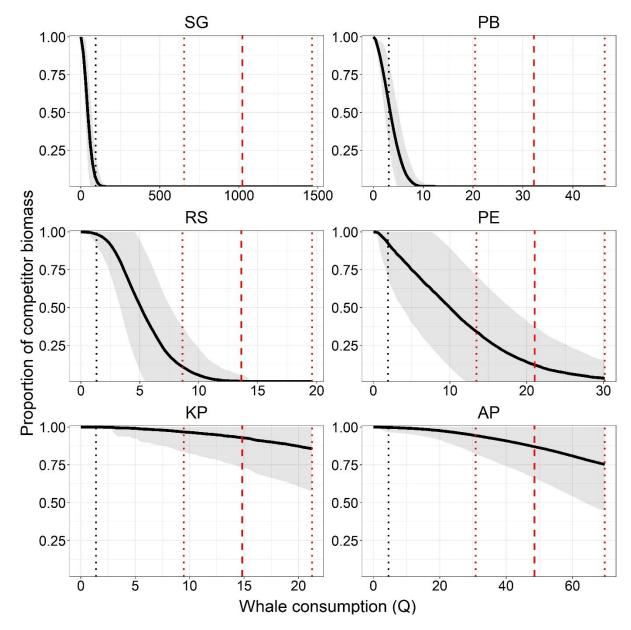
5508 (*n*=1000), for KP, PE and AP.





5511 model inputs and for the IWC total catch-derived biomass estimates from the 1000km buffers

around each model region.



5513

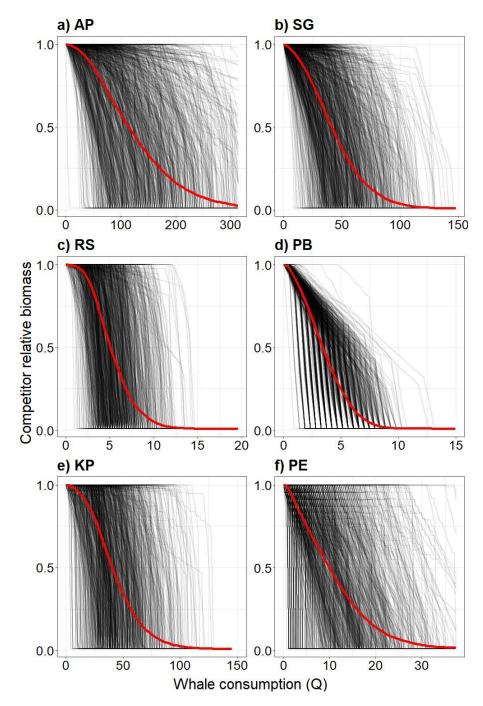
5514 Figure D11: Relationship between competitor biomass proportion and whale consumption

5515 (Q). Solid black line indicates the ensemble average, and shading indicates standard

5516 deviation. Vertical lines indicate the whale Q values which represent combinations of catch-

5517 derived total whale biomass and estimated Q/B values: black dotted = baseline estimates; red

dashed = median estimates from Savoca et al. (2021); red dotted = lower and upper estimates
from Savoca et al. (2021).

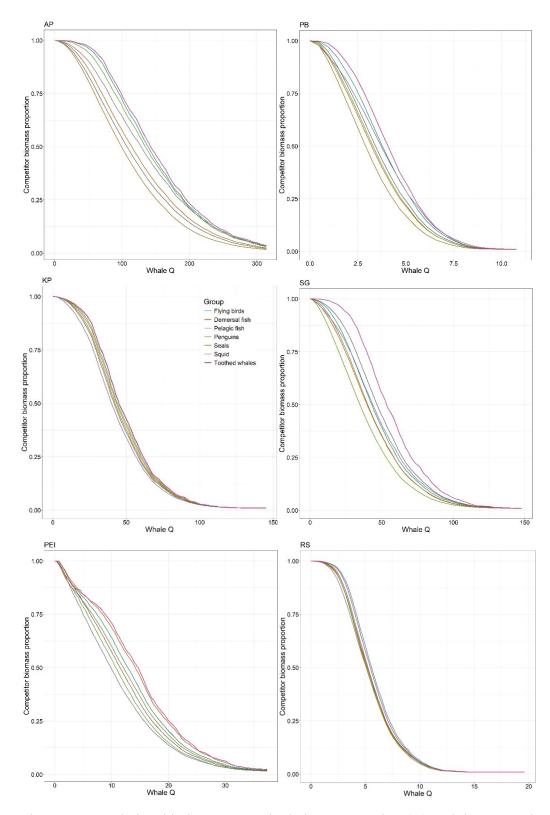




5521 Figure D12: Relationship between competitor biomass proportion and whale consumption

5522 (Q) for each of the model ensembles. Black lines represent individual model runs (n=1000),

5523 while red lines represent the ensemble average. Note varying x axis scales.



5525 Figure D13: Relationship between total whale consumption (Q) and the proportion of 5526 competitor biomass that can be sustained in each model, split by competitor group.

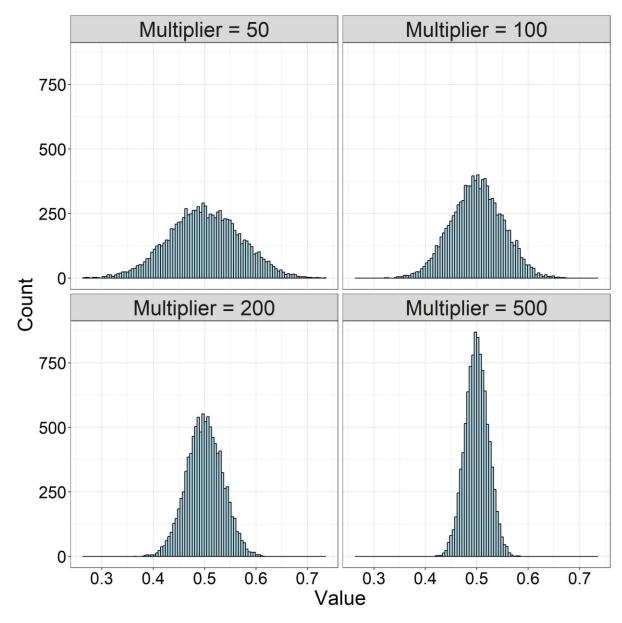


Figure D14: Histograms of the distribution of values generated by resampling the diet of a
predator feeding on two prey in equal proportions, under different scaling factors. Each plot
displays the values generated for one of the prey.

# 5537 D3: Supplementary tables

- 5538 Table D1: Conversion factors used to convert model parameters from to wet weight (gWM m<sup>-</sup>
- $^{2}y^{-1}$ ) to carbon (gC m<sup>-2</sup>y<sup>-1</sup>).

Functional	WM:C	References and notes
group	conversion	
	factor	
Whales	0.105	Sheehy et al. 2022
Seals	0.162	Horn and de la Vega 2016
Birds	0.181	Horn and de la Vega 2016
Cephalopods	0.087	Ikeda 2016.
Demersal fish	0.105	Horn and de la Vega 2016. Average of benthivorous and
		piscivorous demersal fish
Pelagic fish	0.170	Horn and de la Vega 2016. Average of planktivorous and
		piscivorous pelagic fish
Krill	0.107	Kiørboe 2013
Zooplankton	0.049	Kiørboe 2013. Average of ctenophores, tunicates, cnidarians,
		chaetognaths, gastropods, copepods, euphausids and amphipods
Benthos	0.075	Gogina et al. 2022; Pinkerton and Bradford-Grieve 2010.
Denuios	0.075	Based on biomass composition of typical Antarctic benthic
		communities (Gutt et al. 2016; Pineda Metz 2019).
Bacteria	0.024	Bratbak and Dundas 1984; Finlay and Uhlig 1981
Phytoplankton		Ullah et al. 2018; Hatton et al. 2021
Macrophyte	0.460	Hill et al. 2021
Detritus	0.450	Hill et al. 2021
Protozoa	0.154	Kiørboe 2013
Salps	0.007	Kiørboe 2013

....

5549 Table D2: Reaggregated functional groups included in each of the regional models (X

5550 indicates that the group is included in the model, • indicates the group is absent from the

5551 model).

Functional group	SG	RS	PE	KP	PB	AP
Toothed whales	X	X	X	X	X	X
Humpback whales	X	X	X	X	X	X
Minke whales	X	X	X	X	X	X
Other baleen whales	Х	Х	Х	Х	Х	Х
Seals	Х	Х	Х	Х	Х	Х
Penguins	Х	Х	X	Х	Х	X
Flying birds	Х	Х	Х	Х	Х	Х
Demersal fish	Х	Х	Х	Х	Х	Х
Pelagic fish	Х	Х	Х	Х	Х	Х
Cephalopods	Х	Х	Х	Х	Х	Х
Antarctic krill	Х	Х	•	•	Х	Х
Other krill	Х	Х	Х	Х	Х	Х
Salps	Х	Х	Х	Х	Х	Х
Macrozooplankton	Х	Х	Х	Х	Х	Х
Mesozooplankton	Х	Х	Х	Х	Х	Х
Microzooplankton	Х	Х	Х	Х	Х	Х
Benthos	Х	Х	Х	Х	Х	Х
Bacteria	Х	Х	•	Х	Х	•
Phytoplankton	Х	Х	Х	Х	Х	Х
Ice algae	•	Х	•	•	•	Х
Detritus	Х	Х	Х	Х	Х	Х

- Table D3: Standardised diet composition for each whale group in each model. Separate
- 5563 compositions are provided for krill groups as certain krill groups were absent in some
- 5564 models. Abbreviations are: MW = Minke whales; HW = Humpback whales; OBW = Other
- baleen whales; SG = South Georgia; PB = Prydz Bay; RS = Ross Sea; AP = Antarctic
- 5566 Peninsula; KP = Kerguelen Plateau; PE = Prince Edward Islands.

	MW	HW	OBW
Demersal fish	0.029	0.055	0.003
Pelagic_fish	0.128	0.165	0.003
Cephalopods		0.047	
Salps	0.004	0.001	0.003
Macrozooplankton	0.081	0.058	0.083
Mesozooplankton	0.007	0.030	0.160
SG			
Antarctic_krill	0.739	0.634	0.736
Other_krill	0.013	0.011	0.013
PB			
Antarctic_krill	0.479	0.411	0.477
Other_krill	0.272	0.234	0.271
RS			
Antarctic_krill	0.521	0.447	0.519
Other_krill	0.231	0.198	0.230
AP			
Antarctic_krill	0.663	0.569	0.663
Other_krill	0.089	0.076	0.089
КР			
Antarctic_krill	0.000	0.000	0.000
Other_krill	0.752	0.645	0.749
РЕ			
Antarctic_krill	0.000	0.000	0.000
Other_krill	0.752	0.645	0.749

- 5568 Table D4: Constants (a) and metabolic scaling exponents (b) used to calculate the average
- baseline estimates of whale consumption, obtained from Savoca et al. (2021).

a	b
0.1	0.8
0.42	0.67
0.035	1
1.66	0.559
0.123	0.8
0.17	0.773
0.06	0.75

- 5571 Table D5: Overview of the values used to estimate the Q/B for each whale group. Average
- body masses are taken from Greenspoon et al. (2023). Daily rations are from Savoca et al.
- 5573 (2021), multiplied by 90 to estimate annual rations assuming a 90-day feeding period. The

5574 median Q/B for each group in the original published models is also shown.

Group	Median Q/B from publish- ed models	Mean mass (kg)	Baseline daily ration (kg)	Baseline <i>Q/B</i>	Lower daily ration (kg)	Lower Q/B	Median daily ration (kg)	Median Q/B	Upper daily ration (kg)	Upper Q/B
Minke	10.43	6,566	150.78	2.07	362	4.96	685	9.39	1,085	14.87
Humpback	3.50	30,408	502.13	1.49	1,813	5.37	3,151	9.32	4,926	14.58
Other	3.24	79163	1093.75	1.24	7,765. 50	8.69	12,048	13.60	17,057	19.46

5575

5576 Table D6: Pedigree scheme used to identify confidence intervals for the biomass parameters

5577 for each functional group.

Code	Source	Confidence interval (± %)
1	Estimated by Ecopath	80
2	From other model	80
3	Guesstimate	80
4	Approximate/indirect method	50
5	Local sample, low precision	40
6	Local sample, high precision	10

5578

5579

5580

5581

- Table D7: Characteristics of the models. Values represent averages, with brackets indicating
- the standard deviation.

		PB	SG	RS	AP	КР	PE
	Production- weighted	0.658 (0.108)	0.526 (0.132)	0.781 (0.108)	0.584 (0.070)	0.636 (0.125)	0.741 (0.108)
	mean <i>EE</i> of whale prey						
	Schoener's	0.554	0.642	0.573	0.250	0.717	0.458 (0.059)*
	dietary overlap	(0.060)	(0.075)	(0.048)	(0.042)	(0.067)	
5585	*Based on set	ting whale	l biomass to a	an arbritraril	y low value		
5586							
5587							
5588							
5589							
5590							
5591							
5592							
5593							
5594							
5595							
5596							
5597							
5598							
5599							
5600							
5601							
5602							
5603							
5604							
5605							
5606							

- 5607 Table D8: Average total baleen whale biomass (t/km2) across the balanced model ensemble
- 5608 for each region, and total baleen whale catch biomass estimated from the 1000km buffer
- around each model region and applied to the model's spatial area. Also shown are the whale
- 5610 consumption (Q) values estimated from these biomass values when combined with different
- 5611 consumption per unit biomass (Q/B) values: Minimum values derived from prior studies
- 5612 (used as the inputs for balancing the models); the lower estimates from Savoca et al. (2021);
- the upper estimates from Savoca et al. (2021).

	PB	SG	RS	AP	KP	PE
Mean balanced total biomass	0.133	0.288	0.013	0.173	0.026	NA
Total catch biomass	2.44	75.45	1.04	3.62	1.10	1.55
Baseline <i>Q/B</i> with balanced biomass	0.195	0.406	0.023	0.271	0.033	NA
Baseline <i>Q/B</i> with catch biomass	3.132	94.052	1.333	4.586	1.378	1.937
Lower Savoca <i>Q/B</i> with balanced biomass	1.130	2.496	0.110	1.486	0.224	NA
Lower Savoca <i>Q/B</i> with catch biomass	20.397	652.781	8.619	30.846	9.458	13.434
Median savoca <i>Q/B</i> with balanced biomass	1.776	3.908	0.173	2.331	0.351	NA
Median savoca $Q/B$ with catch biomass	32.184	1022.304	13.612	48.482	14.831	21.040
Upper Savoca <i>Q/B</i> with balanced biomass	2.552	5.595	0.249	3.343	0.504	NA
Upper Savoca $Q/B$ with catch biomass	46.362	1464.142	19.622	69.621	21.266	30.137

5615 Table D9: Conversion of average total whale *Q* into biomass at different reference points of

Model	Mean	Catch-	Remaining	Total	Whale	Whale	Whale	Whale
	initial	derived	competitor	whale	biomass	biomass	biomass	biomass
	whale	total	biomass	Q	with	with	with	with
	biomass	whale			baseline	lower	median	upper
		biomass			Q/B	Q/B	Q/B	Q/B
PB	0.133	2.44	99%	0.256	0.202	0.030	0.019	0.013
			75%	2.087	1.649	0.246	0.156	0.109
RS	0.013	1.04	99%	1.159	0.914	0.137	0.087	0.061
			75%	3.652	2.880	0.431	0.274	0.191
KP	0.026	1.10	99%	5.172	4.143	0.599	0.383	0.267
			75%	28.095	22.506	3.256	2.078	1.450
PE	0.000	1.55	99%	0.841	0.675	0.097	0.062	0.043
			75%	5.043	4.049	0.582	0.372	0.260
AP	0.173	3.62	99%	2.103	1.675	0.245	0.156	0.109
			75%	70.255	55.956	8.177	5.213	3.636
SG	0.288	75.45	99%	3.091	2.482	0.357	0.228	0.159
			75%	25.943	20.832	2.994	1.912	1.335

5616 competitor biomass, using multiple estimates of whale Q/B.

5617

5618 Table D10: Primary production required to support the whale consumption (Q) values that

5619 can be achieved by reducing total competitor biomass to either 50% or 10% of starting

values, as identified in Scenario 1. Values are averages across each model ensemble

5621 (n=1000), with standard deviations in brackets.

	AP	PB	SG	RS	КР	PE
Mean starting	19,871.53	2,251.10	4,372.09	1,073.45	4,094.98	1,075.23
primary	(4,246.13)	(669.51)	(926.56)	(318.65)	(933.84)	(216.67)
production						
Mean	19,931.07	2,258.79	4,563.84	1,075.19	4,110.01	1,081.85
primary	(4,240.89)	(669.52)	(974.81)	(318.55)	(935.06)	(217.45)
production at						
75%						
competitor						
biomass						
Mean	19,931.07	2,633.83	28,931.76	1,183.00	4,110.00	1,241.15
primary	(4,240.89)	(716.51)	(9,626.08)	(332.04)	(935.06)	(267.18)
production at						
maximum						
whale $Q$						

5622

5624 Table D11: Details of the relationship between whale consumption (Q) and the proportion of

Model	Max whale Q	Slope
Kerguelen Plateau	21.266	0.000183
Antarctic Peninsula	69.621	0.000046
Prydz Bay	46.362	0.004013
Ross Sea	19.622	0.005649
Prince Edward	30.137	0.005264
Islands		
South Georgia	1464.142	0.000519

5625 extra primary production required to support this increase, for each model.

5626

5627

- - - -